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(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
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Protein-Proteophore complexes

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Protein-Proteophore Complexes

The present invention relates to inclusion compounds comprising a protein and an encapsulating organic compound (EOC), which will also be referred to as proteophore, in a 1:1 stoichiometric. In one embodiment, the EOC is a dendrimer, resulting in a dendrimer-protein inclusion compound (DPIC). In a further embodiment, the EOC is a macrocyclic structure resulting in a macrocycle-protein inclusion compound (MPIC). The encapsulating compounds, as well as the resulting inclusion compounds, are water soluble and lend themselves for the controlled release of the protein to a target, preferably in a living body, in particular a mammal.

Proteins are large and unstable molecules. A large amount of proteins is known which show an important pharmacological activity. Examples include insulin, interferon, growth hormones and blood forming factors. In general, proteins are applied to mammals and humans by injection. To date, it is not possible to apply pharmacologically active proteins orally or transdermally. Following the injection, the proteins are readily attacked and often partially or totally eliminated by the immune system, various enzymes or kidney filtration. In addition, the protein can be toxic or cause allergic reactions.

In order to overcome the in-vivo elimination, several techniques were developed in order to ensure a controlled release of the respective protein. Examples include modification of the protein sequence, pegylation, proteinylation, binding to albumin, glycosylation, formulation into hydrogels or encapsulation by microparticles, nanoparticles, dendrimers.

Supramolecular chemistry is directed towards the synthesis and analysis of inclusion compounds in which two or more components are associated through complete enclosure of one set of molecules in a suitable structure formed by another.

In such chemical host-guest systems, one molecular entity is complementary to a different, second entity. Complementarity can occur in shape or physicochemical properties or in a combination of both. In the case of shape complementarity, the host molecule forms a cavity of size similar to the guest molecule. In such topologically well-defined cases, where the cavity is an inherent structural feature of a single molecule, the host is termed cavitand, and the host-guest aggregate cavitare. Typically, the stoichiometry of the supramolecular system is 1:1. Nevertheless, various types of complex stoichiometries are known. Guest molecules are typically smaller in size than the corresponding host compound.

Examples for small host-guest systems are for instance complexes formed between crown ethers and sodium or potassium ions. Well-known examples for synthetic organic host-guest molecules are the complexation of aromatic compounds such as nitrophenol by cyclodextrin carbohydrates. Cyclodextrins come in various ring sizes, the larger of which can accommodate bicyclic structures such as naphthalene and derivatives. Fullerene molecules are of spherical shape and accommodate free space of a diameter of 0.7 nm.

Biomacromolecules such as starch can form inclusion complexes with small guests such as iodine by filling a channel-like volume in the interior of a helix. DNA double helices are known to accommodate rigid aromatic compounds by means of intercalation. Serum albumin is a well-studied example of a protein that can be loaded with several molecules of fatty acids.

Comparatively little research has been undertaken to investigate the complexation of biomacromolecules such as proteins by synthetic host compounds.

A large number of molecular medicines are based on proteins or peptides of which group insulin, interferons, growth hormones and blood factors are among the most widely used therapeutics.

proteins containing non-human amino acid sequences are highly immunogenic. Antibody formation is even notable for human proteins such as insulin if frequently administered by injection. Biomolecules may be cleared from circulation too fast or too slow for a given therapeutic application and may exhibit a narrow therapeutic window. Proteins can be degraded by endogenous proteases. Most therapeutic biomolecules need to be administered parenterally, often imposing the need for lifetime daily injections on the patient. At this point in time there are no approved protein formulations for oral or pulmonary delivery. Only in a few cases has it been possible to direct the therapeutic protein to the diseased tissue or cell type or to deliver the protein in an intracellular fashion.

Therefore it is highly desirable to develop molecularly defined delivery vehicles to enhance the therapeutic benefit of protein-based medication. Specifically, proteins need to be efficiently encapsulated for protection and released from the encapsulating agent for bioactivity.

Proteins are composed of condensed amino acid sequences that fold into a compact three-dimensional arrangement, often of globular shape. The diameter for globular proteins typically ranges from 1 to 10 nm. Proteins can form complexes comprised of several identical or different subunits, and several proteins can associate to form even larger complexes. A protein encapsulating agent has to provide a well-hydrated internal volume of a similar size and approximately spherical or ellipsoidal or channel-like shape. The water content is an important molecular property, as most biomolecules depend on a hydration shell for bioactivity.

Polymers have proven highly useful in delivery of therapeutic molecular biological material to humans. Linear or branched water soluble polymers can occupy a volume of similar size or greater than a protein molecule.

Polyethyleneglycol (PEG) is a polymer of low toxicity and immunogenicity. Various therapeutic proteins have been covalently conjugated to PEG by a process called PEGylation and successfully applied in molecular therapy (Harris JM, Chess RB, Nature (2003) 214-21).

An advantage of protein PEGylation is the improvement of pharmacokinetic properties of the conjugate in circulation. Unconjugated proteins such as interferon alfa-2a are cleared rapidly within 2.5 h in rats, the corresponding PEGylated interferons circulate with a half-life of $t_{1/2}$ = 3.4 h (linear 5 kDa PEG monoconjugate) up to 23 h (dipegylated with 2 x 20 kDa PEG). This effect is attributed to reduced renal filtration. Kidney filtration is partially a size-exclusion process, and enhancing the hydrodynamic radius of a protein by PEGylation can significantly reduce its rate of clearance. PEG is strongly hydrated (2-3 water molecules per ethyleneoxide unit) and therefore displays a high apparent molecular weight in size exclusion chromatography studies. Due to its high conformational flexibility and hydration, PEG molecules appear 5-10 times as large as proteins of similar molecular mass.

PEG is widely used to render surfaces protein adsorption resistant and to precipitate proteins from aqueous solution, corroborating the notion that PEG does not physicochemically bind to protein.

The property of PEG to fold randomly and to occupy a large molecular volume explains also for the second therapeutic benefit of PEGylation, namely the reduction of immunogenicity of proteins. This effect is most pronounced for proteins of non-human origin and is likely to be achieved by imposing a steric shield in the vicinity of the immunogenic epitope and thereby preventing recognition by the immune system.

The shielding effect may be enhanced by employing branched PEG. Polyethylene glycol with a low degree of branching is known from US 5,643,575 and the 2003 catalogue of Nektar Therapeutics. WO 01/21197 mentions branched monosubstituted insulin-PEG conjugates.

The steric shielding mechanism explains for the observed reduction of bioactivity of PEGylated proteins. Covalent conjugation of protein side chains close to an epitope generally may impair the ability of the protein to bind to its receptor. ~~Conjugation to the amino group of lysine residues is preferred, as it does not affect the protein's structure and function.~~

Steric shielding may also be enhanced by conjugating the protein to more than one PEG molecule. Multiple PEGylation leads to an apparent increase in hydrodynamic volume and serves better to protect the protein from antibody recognition or protease attack. The approach is compromised by loss of bioactivity, loss of therapeutic activity per gram of material and by increasing the risk of protein inactivation by conformational destabilization.

It is a challenge to obtain a homogeneous product from the reaction of protein with PEG reagent. If a PEG reagent is reacted with a given protein under equimolecular conditions or added in slight excess, mono-, bis-, tris- and oligo-conjugations are commonly obtained. The reason is that protein surfaces display various functional groups of similar reactivities. The difficulties in analysis of such mixtures is aggravated by the fact that PEG in itself is a polydisperse molecule. Polydispersity relates to the fact that PEG cannot be obtained as a molecule of precise chain length beyond a degree of polymerization (dp) of 12 ethyleneglycol units. Typical PEGs of MW 5 kDa or 20 kDa exhibit polydispersities of 1.01 up to 1.20 respectively.

Non-water soluble polymers such as poly(lactide-co-glycolide, PLG) may form nano- or microparticles if precipitated from aqueous solution under certain conditions. The formed particles are not water-soluble but are suspended in aqueous solution. Proteins present in the aqueous phase may be entrapped inside these non-covalent assemblies. Proteins are released as the particles degrade. Such hydrogels are successfully used in slow release formulations of therapeutic proteins such as growth hormone (Tracy MA, Biotechnol Prod 14 (1998) 108-15).

Proteins or polypeptides may be incorporated in polymeric material by carrying out the polymerization step in the presence of the biomolecule. Insulin has been loaded to n-butylcyanoacrylate nanoparticles in this fashion (WO 96/31231). Polymerizing monomers are highly reactive molecular species and the process usually requires organic solvents. Biomacromolecules may suffer structural modification or degradation under such conditions.

Typical encapsulation methods involving prepolymerized entities are water-in-oil-in-water (w/o/w) double emulsion/solvent evaporation or the solid-in-oil-in-water (s/o/w) technique. The encapsulation process involves organic solvents such as methylene chloride, heat and

sonication or homogenization and therefore can lead to inactivation of the encapsulated material.

An alternative method is based on polymer crosslinking. Proteins may be permanently entrapped in polymers if the crosslinking step is carried out in the presence of the protein. Protein, monomers and crosslinker are mixed and polymerized. Such polymers are not soluble per se. Crosslinked polymers are constituted of a network of polymer chains. Within this network various pores and cavities and channels exist in a random fashion, some of which may be sufficiently large to allow for diffusion of protein into, through or out of the polymer. The degree of crosslinking has a strong effect on diffusion into and effusion from the polymer. Products from such crosslinking are called hydrogels, as they can be produced from water-soluble, well-hydrated components and exhibit considerable swelling behaviour.

All of these methods of preparation may have severely detrimental effects on the protein integrity and bioactivity. As a portion of the protein material is inactivated during the particle preparation process, and it is difficult to quantify the remaining bioactivity of the entrapped protein sample.

Even after encapsulation, the protein is put under stress as the protein is forced to make tight contact with the more hydrophobic polymer molecules. This again may cause additional denaturing and loss of activity. Additionally, the molecular architecture of the polymer network imposes mechanical and physicochemical stress on the protein. The protein may be dehydrated or denatured by aggregation or contact to internal surfaces, and it is difficult to analyze the protein's bioactivity after encapsulation.

The release of proteins from the entrapment can be achieved by diffusion, a chemical or enzymatic reaction leading to degradation of the polymer or solvent activation (through osmotic or swelling) or a combination of mechanisms. For therapeutic applications, efficient

techniques such as mixing the drug with the lipid in an organic solvent, addition of an aqueous medium and subsequent removal of the organic solvent or dialysis of mixed lipid-detergent micelles are not readily applied to protein encapsulation due to protein denaturation by solvent or detergent. A more suitable approach is lipid film hydration. Liposomes are formed by hydrating and dispensing a previously dried film of lipid. Liposomes are not per se water-soluble but can be homogeneously distributed in water by means of dispersion. If protein is present in the hydration solution it becomes both associated on the surface and entrapped in the interior of the liposomes. The process reduces the exposure of protein to denaturing conditions but is of little encapsulation efficiency.

Dendrimers are well-defined polymeric structures. Dendrimers are based on repeating hyperbranched structures emanating from a central core (US 4,507,466). Typical dendrimers are based on polyamidoamine (PAMAM), polyethylene imine (PEI), polypropylene imine or polylysine. These synthetic macromolecules are assembled in a stepwise fashion, with each reaction cycle adding another layer of branches (dubbed „generation“). Dendrimers are synthetically accessed by stepwise, divergent „bottom-up“ or convergent „top-down“ synthesis. Central structural component is the core unit from which hyperbranched dendrimers extend in a radially symmetric fashion. The core may provide at least two reactive groups for dendrimer conjugation, it may also be of heterofunctional nature and protecting groups may be used. In the latter case, the dendrimer may be assembled, and a guest compound may be subsequently conjugated to an anilin core by means of orthogonal chemistries (WO 88/01180). The core and dendrimers form the interior or backbone of a dendrimer. As a consequence of the spherical symmetry supported by sterical crowding, the terminal groups of the hyperbranches are defining the exterior. In higher generation dendrimers, the terminal branches form rather dense shells and flexible internal voids have been discovered. It is understood, that for a given dendrimer these cavities are filled up by backfolded end groups and tightly coordinated solvent molecules. Dendrimers are related to micelles, similary well suited to complex hydrophobic compounds. But in contrast they exhibit higher structural order because of their monomolecular nature and the absence of a dynamic equilibrium of various species. Synthetic compounds can only diffuse into dendrimers if certain structural requirement such as conformational rigidity and flatness as well as charge distribution such as affinity to tertiary amines are met. Various apolar compounds such as pyrene or naphthalene have been encapsulated in dendrimers, but the

number of trapped guests as well as their molecular interaction with the dendrimer interior are rather undefined and frequently substoichiometric.

In US 5,714,166 and WO 95/24221, dendrimer-protein conjugates are revealed. PAMAM dendrimers of G4 are covalently coupled through their terminal functional groups to insulin, fluorescently labeled insulin, avidin, monoclonal antibodies and bradykinin. The reactive groups used for conjugation are only present at the surface of the dendrimers, and therefore any covalent adduct generated by the taught method will be associated with the dendrimer exterior. Sterical „congestion“ of the dendrimeric terminal groups is a prerequisite to the formation of internal void space. In a scanning transmission electron micrograph study, it was observed that PAMAM dendrimers undergo a morphological change at the G9 stage. Surface congestion created a hollow interior surrounded by a dense rim. The G4 dendrimers used for protein conjugation do not contain such voids. Furthermore it is apparent from molecular size comparison, that a 3 nm sized insulin may not be encapsulated in a dense, 4 nm-sized generation 4 PAMAM dendrimer. Hemoglobin has a diameter of 5.5 nm, and PAMAM dendrimers of G5, G6 and G7 exhibit diameters of 5.3 nm, 6.7 and 8.0 nm respectively. Macromolecules such as peptides and proteins are per se excluded from diffusion through the dense molecular packing and entering the interior of such dendrimers. As the dendrimer surface is rather densely clustered, pore sizes are too small to allow for an entry of a protein into the dendrimer interior. For these reasons, macromolecular protein or polypeptide guests have not been encapsulated in dendrimers, neither has the non-covalent encapsulation of proteins been demonstrated.

PAMAM dendrimers contain free amine groups on their surfaces and readily associate with DNA through electrostatic interactions.

WO 01/07489 details water-soluble polypeptide dendrimers constituted of ornithin and glycineamino acid. The patent application also teaches the non-covalent encapsulation of an

applicable to proteins as substantial side reactions between the maleimido core and the protein will occur, furthermore steric competition will prevent an efficient encapsulation as either full formation of the tri-dendritic structure is prevented or the protein will not be entrapped. The example does not teach how to generate a complex of well-defined stoichiometry.

There is a continuous need for techniques and devices which allow for an effective encapsulation of proteins in order to ensure a controlled delivery and, if appropriate, release of pharmacologically active proteins. The encapsulation should not alter the proteins' structure and properties and should efficiently protect the protein from attacks by the immune system and enzymes of the individual to which the protein is administered. Furthermore, the protein should enable an efficient release of the encapsulated protein, in case this is desired.

This object is attained by a protein encapsulated covalently or non-covalently by an encapsulating organic compound (EOC) wherein the protein and the encapsulating organic compound are present in 1:1 stoichiometry.

Appropriate EOCs are water soluble.

The EOCs contain several, i.e. at least 2, molecule chains of an appropriate length which chains can arrange such that a cavity is formed which can accommodate the protein and protect it from the action of enzymes, antibodies and the like. The molecule chains will hereinafter be referred to as "encapsulating molecular chains" EMC. The EMCs can be directly connected with each other, or via a chemical unit, often one or more so-called branching units (see below).

In the following, EMCs according to the present invention will be defined. This definition applies every time EMCs will be mentioned in the present application in a general form, either in connection with a general formula or in any other context.

The EMCs contain hydrophilic groups, in an appropriate ratio and amount with respect to hydrophobic groups which may be present in the EOC, to render the latter water soluble.

The EMCs are built up from linear, branched or cyclical alkyl chains. To render the hydrophobic alkyl chains more hydrophilic, hetero atoms like but not limited to S, N, O may be present within the chain. Further appropriate groups which can be present in the EMCs include (-S-S)-, amide -C(O)NH- or C(O)NR-, -S-succinimido, amino (-NR-), carboxylic ester (-C(O)O-), sulfonamide (-S(O)₂-NR-, carbamate (-O-C(O)-NR-), carbonate (-O-C(O)-O-

), sulfone ($-S(O)_2-$), ether ($-O-$), oxime ($-CR=N-O-$), hydrazone ($-CR=N-NR-$), urea ($-NR-C(O)-NR-$), thiourea ($-NR-C(S)-NR-$), carbohydrate, glyceryl, phosphate ($-O-P(O)(OR)O-$), phosphonate ($-P(O)(OR)O-$), saturated and nonsaturated (hetero)cyclic compounds. Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms. In addition to the afore-mentioned groups, further groups known to the person skilled in the art can be present in the EMCs.

Example for preferred groups in the EMCs comprise oxyalkylene groups (i.e. oxyethylene ($-OCH_2CH_2-$), oxypropylene groups ($-OCH_2CH(CH_3)-$ and oxybutylene groups) and amide groups ($-C(O)NH-$). It is preferred if the EMCs comprise oxyethylene groups ($-OCH_2CH_2-$) and amide groups ($-C(O)NH-$).

In one embodiment of the present invention, the EMCs comprises at least one amino acid unit in its chain. In the context of the present invention, "amino acid unit" means an amino acid, preferably a naturally occurring amino acid like lysine, which is connected to at least one further binding partner, for example a further amino acid, by its amino and/or its carboxy function. The amino acid may be modified, e.g. carry one or more substituents.

The EMCs can carry one or more substituents (capping groups or modifiers C) on their backbone. Appropriate capping groups are sterically demanding groups. The capping groups will in particular be present if the EMCs require sterically demanding groups forcing them into a certain conformation necessary for the creation of the cavity enclosing the protein. In many cases, the EMCs according to the present invention are not rigid, and the subunits of the EMCs may rotate around the bonds of the chain and occupy a spatial position in accordance with the sterical requirements (which, in general, will be the position with the lowest energy). The capping groups can avoid a too close approaching of the EMCs and an opening of the cavity which may result in an insufficient encapsulating of the protein and an insufficient protection from the attack of enzymes, antibodies or the like. Furthermore, the protein may totally leave the cavity through the gap resulting from the movement of the EMCs.

The capping groups C are built up from linear, branched or cyclical alkyl chains. To render the hydrophobic chain more hydrophilic, it may contain one or more hydrophilic groups.

P(O)(OR)O-), phosphonate (-P(O)(OR)O-), saturated and nonsaturated (hetero)cyclic compounds. Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms. In addition to the aforementioned groups, further groups known to the person skilled in the art can be present in the EMCs.

In one embodiment of the present invention, the capping units comprise at least one amino acid unit in its chain. In the context of the present invention, "amino acid unit" means an amino acid, preferably a naturally occurring amino acid like lysine, which is connected to at least one further binding partner, for example a further amino acid, by its amino and/or its carboxy function. The amino acid may be modified, e.g. carry one or more substituents.

It is preferred if the capping groups comprise oxyalkylene groups (i.e. oxyethylene (-OCH₂CH₂)-, oxypropylene groups (-OCH₂CH(CH₃))- and oxybutylene groups) and amide groups (-C(O)NH-). It is even more preferred if the capping groups comprise oxyethylene groups (-OCH₂CH₂)- and amide groups (-C(O)NH-), in an appropriate ratio and amount, in order to obtain capping groups with the desired hydrophilicity which may be higher or lower than the hydrophilicity of the EMCs.

The capping groups in the EMC can contain one or more functional groups from those cited above. The functional groups present in a given capping group can be identical or different. Each of the cited groups can be present only once or several times. The capping groups present in a given EMC can be identical or different.

In one embodiment of the present invention, the capping groups do not have a high branching degree. This will in particular be the case if the EOCs according to the present invention have a high number of EMCs.

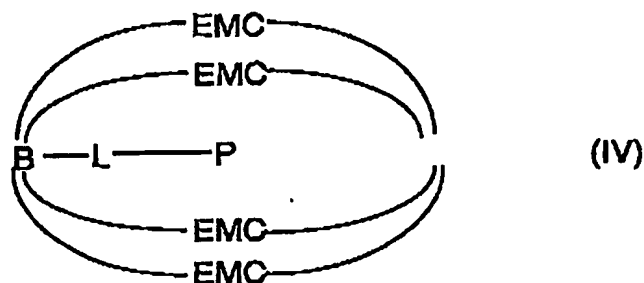
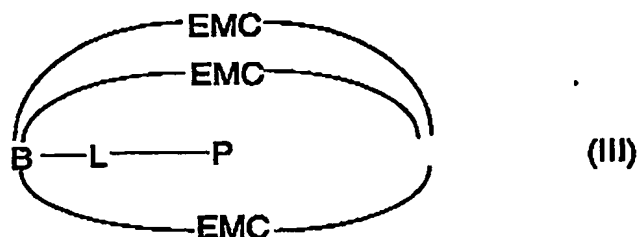
In a further embodiment of the present invention, the capping groups are highly branched molecules having preferably a branching degree of 2, 3, 4, 5 or 6. A branching degree of 2 means that the principal chain connected to the encapsulating unit splits up into 2 subchains, whereas in the case of a branching degree of 3, the main chain splits up into 3 subchains, etc.

The subchains may themselves also be branched. In the context of the present invention, this case will be referred to as "subbranched" (The modifiers are subbranched, i.e. their main chain contains subchains which themselves are branched.) For example, in the case of a branching degree of 2, each of the 2 subchains can be subbranched to a subbranching degree

- L:** linker containing at least one functional group Lfg which is connected to the protein P or capable of connecting with functional groups present on the protein P under the formation of a chemical bond;
- I:** 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12, preferably 3, 4, 5, 6, 7 or 8, in particular 2,3,4,5 or 6;
- P:** pharmacologically active protein.

The EMCs have been defined beforehand. In the following, the groups B, L and P according to the formulae (I) and (II) will be defined. This definition applies every time B, L and P will be mentioned in the present application in a general form, either in connection with a general formula or in any other context.

The EOCs according to the present invention may comprise more than 2 EMCs, for example 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13. In the formulae below, some preferred embodiments are shown.



The branching units B can be regarded as the basic unit or core of the EMCs according to the present invention.

The EMCs are linked by an EMC functional group to at least one branching unit B. B contains at least one branching center Bc. Examples of Bc include units like $>\text{CH}-$ or $>\text{C}<$ and the respective analogues wherein H is replaced by an organic group; $>\text{N}-$; $>\text{P}-$. The centers Bc can directly be linked to the branching functional groups (see below), or can be linked to at least one organic chain.

Examples for appropriate organic chains include linear, branched or cyclical alkyl chains. Hetero atoms like but not limited to S, N, O may be present within the chain. Further appropriate groups which can be present in B include $(-\text{S}-\text{S}-)$, amide $-\text{C}(\text{O})\text{NH}-$ or $\text{C}(\text{O})\text{NR}-$, $-\text{S}-\text{succinimido}$, amino $(-\text{NR}-)$, carboxylic ester $(-\text{C}(\text{O})\text{O}-)$, sulfonamide $(-\text{S}(\text{O})_2-\text{NR}-)$, carbamate $(-\text{O}-\text{C}(\text{O})-\text{NR}-)$, carbonate $(-\text{O}-\text{C}(\text{O})-\text{O}-)$, sulfone $(-\text{S}(\text{O})_2-)$, ether $(-\text{O}-)$, oxime $(-\text{CR}=\text{N}-\text{O}-)$, hydrazone $(-\text{CR}=\text{N}-\text{NR}-)$, urea $(-\text{NR}-\text{C}(\text{O})-\text{NR}-)$, thiourea $(-\text{NR}-\text{C}(\text{S})-\text{NR}-)$, carbohydrate, glyceryl, phosphate $(-\text{O}-\text{P}(\text{O})(\text{OR})\text{O}-)$, phosphonate $(-\text{P}(\text{O})(\text{OR})\text{O}-)$, saturated and nonsaturated (hetero)cyclic compounds. Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms. In addition to the afore-mentioned groups, further groups known to the person skilled in the art can be present in B.

B can contain one or more groups chosen from those cited above. The groups can be identical or different. Each of the cited groups can be present only once or several times. In a preferred embodiment of the present invention, B comprises at least one amino acid unit, preferably of a naturally occurring amino acid like lysine. It is even more preferred if B contains a unit composed of several amino acid units.

In general, B will be a branched structure containing one or more of the above mentioned groups and having a certain length, in accordance with the steric requirements of the protein to be encapsulated. B will comprise one or more branching center. Furthermore, B will contain at least two branching functional groups Bfg allowing for the attachment of the EMCs.

Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms or aryl groups.

As will be apparent to the person skilled in the art from the foregoing, the functional groups present on the EMC and B (Bfg) of the EOC will be transformed (and changed) into a bond species. When, in the context of the present invention, reference is made to "functional group" present on B and the EMC, this refers to the functional groups as such, as well as to the bond species formed by reaction between the groups.

In the context of the present invention, "bond" or "chemical bond" refers to the attraction forces as such between two or more atoms (e.g. a "covalent bond"), whereas "bond species" denotes the chemical bond and the atoms in the vicinity which is involved in the binding process (e.g.-S-S-).

As the bond species depicted beforehand are formed by the reaction between functional groups (which functional groups can be identical or different), the EMCs and B before forming the bond species, both contain functional groups which are capable of reacting with each other under the formation of an appropriate chemical bond, preferably one of the bonds mentioned beforehand:

Examples for appropriate branching functional groups Bfg and EMC functional groups comprise amino (-NRH), carboxylic acid (-C(O)OH) and derivatives, sulfonic acid (-S(O)₂-OH) and derivatives, carbonate (-O-C(O)-O-) and derivatives, hydroxyl (-OH), aldehyde (-CHO), ketone (-CRO), hydrazine (H₂N-NR-), isocyanate (-NCO), isothiocyanate (-NCS), phosphoric acid (-O-P(O)(OR)OH) and derivatives, phosphonic acid (-P(O)(OR)OH) and derivatives, haloacetyl, alkyl halides, maleimide, acryloyl, arylating agents like aryl fluorides, hydroxylamine, disulfides like pyridyl disulfide, vinyl sulfone, vinyl ketone, diazoalkanes, diazoacetyl compounds, epoxide, oxirane, aziridine, Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms or aryl groups.

In a preferred embodiment of the present invention branching functional groups and EMC functional groups comprise amino, carboxylic acid and derivatives, hydrazine, hydroxylamine, thiol, aldehyde, hydroxyl, carbonate, maleimide or haloacetyl groups.

As already mentioned, B can contain two or more Bfgs. This is in particular the case when the EOC comprises more than four, for example 5, 6, 7, 8 or more EMCs.

Likewise B can contain one or more branching centers Bc.

A further essential constituent of the EOCs according to the present invention is the linker L which serves to establish a chemical bond between the protein P and the EOC, by the reaction between appropriate functional groups Lfg on the linker L and the protein. The chemical bond can be a covalent bond or a non-covalent bond, for example a coordinative bond. Preferably, the EOC has 1 or 2 linkers.

In the context of the present invention, several sorts of linkers can be employed:

Non-cleavable linker: a linker containing no selectively cleavable bonds.

Cleavable linker: a linker containing a bond that can be selectively cleaved by a cleavage reagent (TCEP, TFA, DTT, enzyme, or a buffer).

Traceless linker: a linker that upon cleavage releases protein in such a fashion that the protein is not associated with a remaining linker cleavage product.

Prodrug linker: a cleavable linker containing a bond that can be selectively cleaved under in-vivo conditions, for instance in the presence of endogeneous enzymes or other endogeneous reagents, or solely in aqueous buffer.

Traceless prodrug linkers: linkers having both the characteristics of prodrug linkers and traceless linkers.

Depending on the therapeutic application, the protein may need to be permanently

In many cases, the release of the protein from the EOC/EPIC is mandatory for its bioactivity. One example is insulin which, in order to bind to its receptor, must diffuse out of the shielding EOC. Protein release may be achieved by cleaving the covalent tether between protein and EOC.

Cleavable linkers are known in the art (see Hermanson).

The linker L can react with any appropriate functional group Pfg present on the protein P, preferably with those mentioned below.

Examples for suitable bond species formed between the protein P and the linker L include the following: (-S-S)-, S-succinimido, amide -C(O)NH- or C(O)NR-, -S-succinimido, amino (-NR-), carboxylic ester (-C(O)O-), sulfonamide (-S(O)₂-NR-), carbamate (-O-C(O)-NR-), carbonate (-O-C(O)-O-), ether (-O-), oxime (-CR=N-O-), hydrazone (-CR=N-NR-), urea (-NR-C(O)-NR-), thiourea (-NR-C(S)-NR-), phosphate (-O-P(O)(OR)O-), phosphonate (-P(O)(OR)O-). Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms or aryl groups.

In the context of the present invention, "bond" or "chemical bond" refers to the attraction forces as such between two or more atoms (e.g. a "covalent bond"), whereas "bond species" denotes the chemical bond and the atoms in the vicinity which is involved in the binding process (e.g.-S-S-).

As the bond species depicted beforehand are formed by the reaction between functional groups (which functional groups can be identical or different), the EOC, before reacting with the protein, and the protein, before reacting with the EOC, both contain functional groups which are capable of reacting with each other under the formation of an appropriate chemical bond, preferably one of the bonds mentioned beforehand.

Examples for appropriate protein functional groups Pfg which are part of the amino acids forming the natural (i.e. non-modified) protein are amino, thiol, hydroxyl, phenol, imidazole, amide, indole, carboxylic acid and guanidino groups.

In a preferred embodiment of the present invention Pfgs comprise amino, imidazole and thiol groups.

Examples for appropriate linker functional groups comprise amino ($-NH_2$), carboxylic acid ($-C(O)OH$) and derivatives, sulfonic acid ($-S(O)_2-OH$) and derivatives, carbonate ($-O-C(O)-O-$) and derivatives, hydroxyl ($-OH$), aldehyde ($-CHO$), ketone ($-CRO$), isocyanate ($-NCO$), isothiocyanate ($-NCS$), haloacetyl, alkyl halides, maleimide, acryloyl, arylating agents like aryl fluorides, disulfides like pyridyl disulfide, vinyl sulfone, vinyl ketone, diazoalkanes, diazoacetyl compounds, epoxide, oxirane, aziridine, Non-limiting examples of R include H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms or aryl groups.

In a preferred embodiment of the present invention Lfgs comprise carbamate, carbonate, thiol, thioether, succinimidyl, amide and disulfide.

As will be apparent to the person skilled in the art from the foregoing, the functional groups present on the protein P (Pfg) and the linker L (Lfg) of the EOC will be transformed (and changed) into a chemical bond. When, in the context of the present invention, reference is made to "functional group" present on the linker L and the protein P, this refers to the functional groups as such, as well as to the bond species formed by reaction between the groups.

A part of the present invention are traceless double prodrug linker structures and their EPICs resulting in a novel mechanism of cleavage and subsequent release of the protein from the EOC.

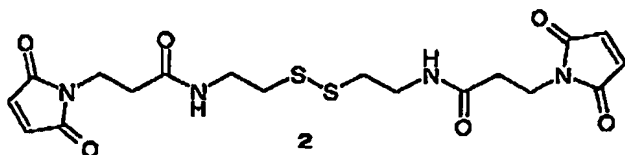
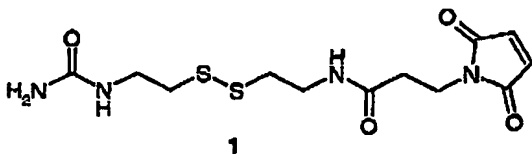
Many widely applied and commercially available protein linker reagents cleave in such a fashion that part of the linker remains conjugated to the protein. In such linker structures the linker is not cleaved at the point of attachment to the protein, but rather at a point further away from the protein, leaving a part of the linker attached to the protein. This is not desirable, as it may lead to the formation of a stable adduct, which may be difficult to remove. The present invention provides a method for the cleavage of such linker structures, resulting in the release of the protein from the EOC.

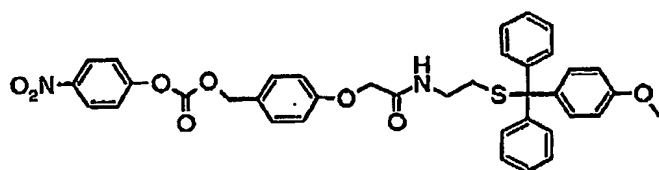
More advantageous are cleavable, traceless linkers that release the protein in an unmodified form under in vivo conditions such as neutral pH without the addition of chemical or biological cleaving agents. Examples are double prodrugs which are based on linker moieties which are cleaved in a two-step process in vivo. WO 99/30727A1, which is incorporated herein by reference, reveals conjugates containing a PEG moiety, a double prodrug linker and protein. The advantage of such systems is that the protein is released in an unmodified form. The linker cleavage process is traceless, the protein end product of the cleavage step do not contain remnants of the linker structure. In a first, rate-determining step one bond is hydrolyzed. This is typically an ester bond, such as in a phenol ester, and hydrolysis may occur by enzymatic attack (lipases) or autohydrolysis or a combination of both. The resulting free phenol is instable and rapidly rearranges for instance through 1,4- or 1,6-arylelimination, and cleavage of a carbamate releases the protein, CO₂ and an instable aromatic moiety.

Furthermore, linker are known to the person skilled in the art that can be cleaved in such a fashion that after cleavage no parts of the linker remain at the EOC.

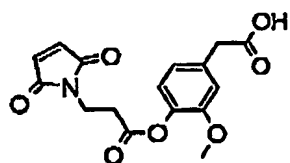
Thus, a preferred embodiment of the present invention are traceless prodrug linkers which contain an ester functionality, in particular a phenol ester functionality, and a carbamate functionality.

Examples for suitable linker reagents are those according to the formulae (1), (2), (5), (6) and (7).

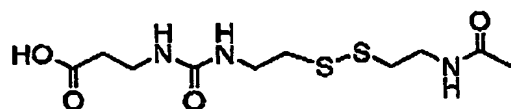




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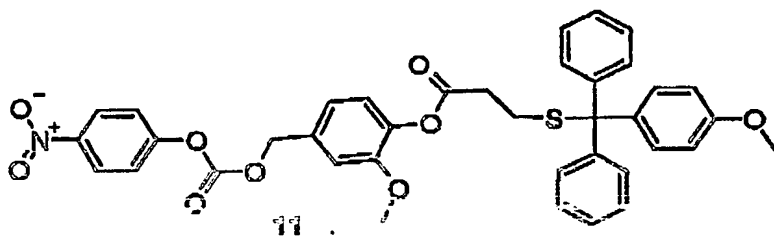


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7

The most preferred linker reagent is the linker reagent according to the formula (11) below (traceless prodrug linker).



11

After the cleavage of the linker, a EPIC results in which the protein is not connected to the EOC via a linker (a chemical bond), but the protein is held within the cavity defined by the EOC. The resulting EPICs are an object of the present invention. It depends on the release kinetics of the protein if the respective EPIC having no bond between the protein and the EOC can be isolated as such.

In the EPICs according to the present invention, the protein can be encapsulated entirely or partially by the EOC. It is preferred to encapsulate the protein entirely, i.e. the cavity is of a size sufficiently large to accept the entire protein therein.

The EPICs according to the present invention can, in principle, accommodate any protein which has a physiological or pharmacological activity. These are known to the person skilled in the art. Important proteins can be found in standard text books which are known to the skilled artisan.

Relevant therapeutic proteins and polypeptides which can be encapsulated according to the present invention are: ACTH, adenosine deaminase, agalsidase, albumin, alfa-1 antitrypsin (AAT), alfa-1 proteinase inhibitor (API), alteplase, anistreplase, anicrod serine protease, antibodies (monoclonal or polyclonal, and fragments or fusions), antithrombin III, antitrypsins, aprotinin, asparaginases, biphalin, bone-morphogenic proteins, calcitonin (salmon), collagenase, DNase, endorphins, enfuvirtide, enkephalins, erythropoietins, factor VIIa, factor VIII, factor VIIIa, factor IX, fibrinolysin, fusion proteins, follicle-stimulating hormones, granulocyte colony stimulating factor (G-CSF), galactosidase, glucagon, glucocerebrosidase, granulocyte macrophage colony stimulating factor (GM-CSF), gonadotropin chorionic (hCG), hemoglobins, hepatitis B vaccines, hirudin, hyaluronidases, iduronidase, immune globulins, influenza vaccines, interleukins (1 alfa, 1 beta, 2, 3, 4, 6, 10, 11, 12), IL-1 receptor antagonist (rhIL-1ra), insulins, interferons (alfa 2a, alfa 2b, alfa 2c, beta 1a, beta 1b, gamma 1a, gamma 1b), keratinocyte growth factor (KGF), lactase, leuprolide, levothyroxine, luteinizing hormone, lyme vaccine, natriuretic peptide, pancrelipase, papain, parathyroid hormone, PDGF, pepsin, platelet activating factor acetylhydrolase (PAF-AH), prolactin, protein C, octreotide, secretin, sermorelin, superoxide dismutase (SOD), somatopins (growth hormone), somatostatin, streptokinase, sucrase, tetanus toxin fragment,

tilactase, thrombins, thymosin, thyroid stimulating hormone, thyrotropin, tumor necrosis factor (TNF), TNF receptor-IgG Fc, tissue plasminogen activator (tPA), TSH, urate oxidase, urokinase, vaccines.

Preferred proteins are antibodies, calcitonin, G-CSF, GM-CSF, erythropoietins, hemoglobins, interleukins, insulins, interferons, SOD, somatropin, TNF, TNF-receptor-IgG Fc.

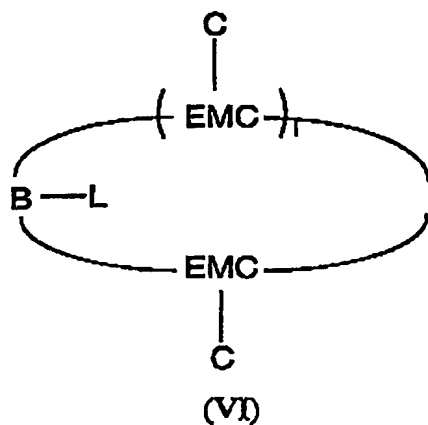
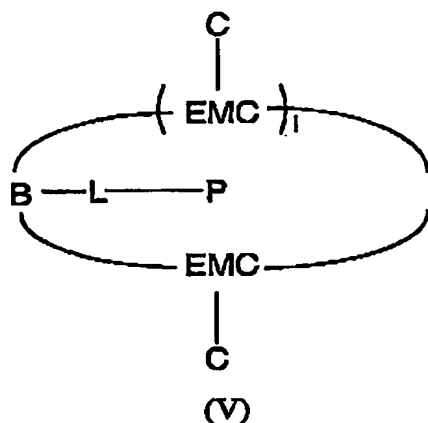
The most preferred proteins are erythropoietins, interferons, insulins, somatropins and hemoglobins.

It is understood that the invention is not restricted to therapeutic proteins. Protection from aggressive environments is also desirable for other proteins such as amylases, proteases, peptidases, xylanases, lipases, lipoxygenases, cellulases, pectinases, phytases, oxidoreductases applied in industrial processes such as food and animal feed applications, as cleaning compounds in laundry detergents, dishwashing detergents, in the manufacture of chemicals such as alcohol, steroids and antibiotics, amino acids, proteins, triglycerides, phospholipids, and for textile, leather and fur applications, especially in the prebleaching of pulp.

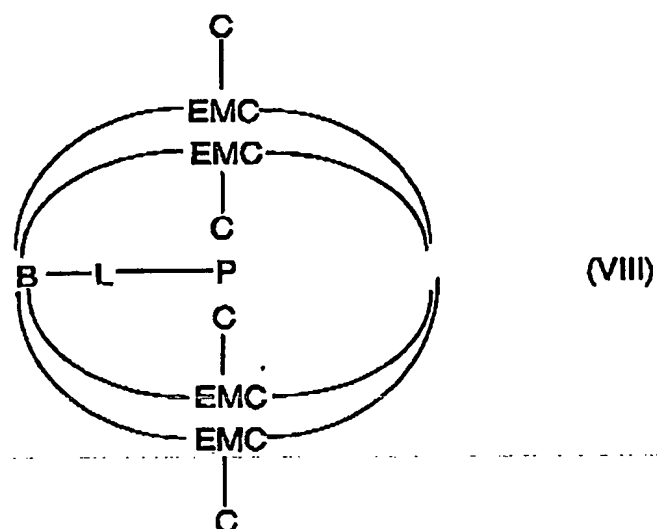
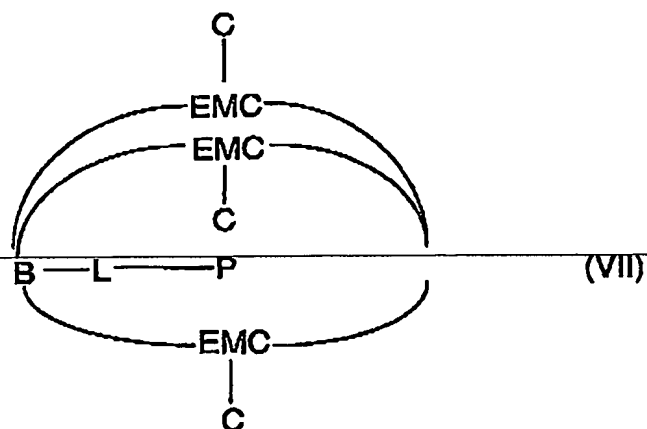
All proteins, in particular those cited beforehand, can be encapsulated in a macrocyclic structure according to the present invention, to result in an MPIC, or in a dendrimer resulting in a DPIC.

The size of the cavity in the EOCs (i.e. the proteophors, macrocyclic structures and dendrimers) according to the present invention needs to be adapted to the proteins diameter. The size should be larger than the diameter of the smallest sphere that can be drawn around a correctly folded protein. From this diameter estimation, the length of the corresponding molecular chain in the EOC host can be calculated. In order to encapsulate insulin

results. When such an EMC encloses a protein, dendrimer-protein inclusion compounds DPIC according to the formula (V) wherein a dendrimer (VI) encapsulates the protein result. Such DPICs are a part of the present invention. In the following formulae (V) and (VI) C denotes a capping group as defined beforehand and the other symbols have the meanings defined for formula (I).



Embodiments of the capped EOCs having three and four EMCs are shown in the formulae (VII) and (VIII) below.



The DPICs according to the general formulae (V) to (VIII) show a 1:1 ratio protein/dendrimer. The DPICs are soluble in water. In general, the cavity of the dendrimer also comprises water, in addition to the protein.

Dendrimers are known to the person skilled in the art. Reference is made to: Dendrimer II Architecture, Nanostructure and Supramolecular Chemistry, Springer Verlag 2000, T. Vögelle. ~~Further information can be found at: <http://www.dendrimers.com>~~

synthesis. Central structural component is the core unit from which hyperbranched dendrimers extend in a radially symmetric fashion.

The dendrimers according to the present invention may contain, in the capping groups, centers branching into two, three, four, or more directions, preferably two.

The length of dendritic chains may be identical or vary between chains of one dendrimer. Preferred chain lengths for individual dendrimers are up to 5000 bonds.

By the choice of appropriate capping groups, it is perfectly possible to protect the encapsulated protein from the attack of e.g. antibodies or the elimination by the kidney or the liver.

The capping groups C have been defined above, which definition also applies here.

In all formulae (V) to (VIII) shown beforehand, an EMC can carry one capping group C (as shown in the formulae) or more than one, i.e. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or even more capping groups C.

In a preferred embodiment of the present invention, some capping groups in dendrimers according to the invention contain branched heterofunctional units carrying at least one thio-succinimido moiety. Thio-succinimido groups are the result of a reaction between a maleimido and a thiol group and can be obtained under mild conditions, proving useful for the synthesis of hyperbranched or dendritic structures. Further units which can be used in the synthesis of the dendrimers according to the present invention comprise tris-(2-maleimidoethyl)amine and hydroxysuccinimide ester (EP-A 0 618 192).

Most useful for the divergent assembly of hyperbranches or dendrimers are heterofunctional reagents carrying both a maleimido as well as a protected thiol group. Such reagents may be employed for repeated stepwise synthesis in a similar fashion as for instance protected amino acids or protected nucleosides.

It was found that the dendrimers according to the present invention can efficiently be formed of multidentate compounds containing only one maleimide group and a number of protected thiols, in a divergent synthesis approach. The monomaleimido-tetrathio-dendrimer compounds according to the present invention are of the general formula $M-A-(S-Pg)_n$, with the following meanings: M: maleimido, A: spacer, S: sulfur, Pg: thiol protecting group, n: 2 to 200.

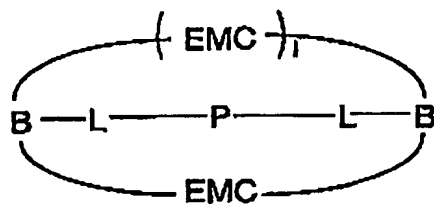
Suitable thiol protecting groups: benzyl, 4-methoxybenzyl, 2,4-dimethoxybenzyl-, 2,4,6-trimethoxybenzyl-(Tmob), 4,4'-dimethoxyphenylmethyl-(diMpm), trityl-, 4-methoxytrityl-(Mmt), 4,4'-dimethoxytrityl- (DMTr), 4,4',4''-trimethoxytrityl- (TMTr), tert.-butyl-MeCONHCH₂- (Acm), PhCH₂CONHCH₂- (PhAcm), MeOCOS- (Scm), BzlOCOS- (SZ) PhN(Me)COS- (Snm), TrtS-, 2-pyridinesulfenyl-, 2-(3-nitropyridinesulfenyl), 4,5,6-trimethoxy-2,3-dihydro-7-benzofuranylmethyl- (Tmbf), 2-(2,4-dinitrophenyl)ethyl- (Dnpe), 9H-xanthen-9-yl- (Xan), 2-methoxy-9H-xanthen-9-yl- (Moxan), Fmoc-S-sulfonate.

Multidentate compounds are water soluble and the conjugation reactions will not compromise the biomolecule's structural integrity or bioactivity.

According to the present invention, the core of the DPIC is formed by the protein to be encapsulated which is connected (conjugated) to the polymer backbone by a suitable linker, in general one of the linkers listed above.

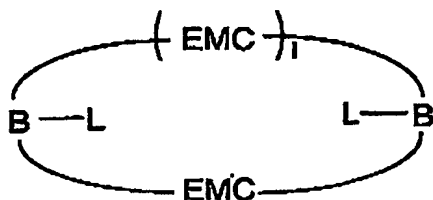
After the cleavage of the linker, a DPIC results in which the protein is not connected to the dendrimer via a linker (a chemical bond), but the protein is held within the cavity defined by the dendrimer. The resulting DPICs are an object of the present invention. It depends on the release kinetics of the protein if the DPIC having no bond between the protein and the dendrimer can be isolated such

encapsulation is realized by a macrocycle-protein inclusion compound MPIC according to the formula (IX)



(IX)

containing a protein P and a macrocyclic structure encapsulating the protein totally or partially according to the general formula (X)



(X)

wherein the symbols in formulae (IX) and (X) have the following meanings:

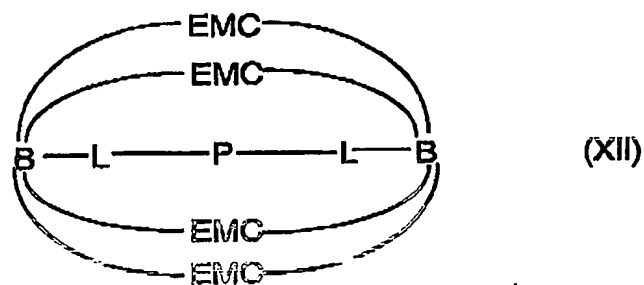
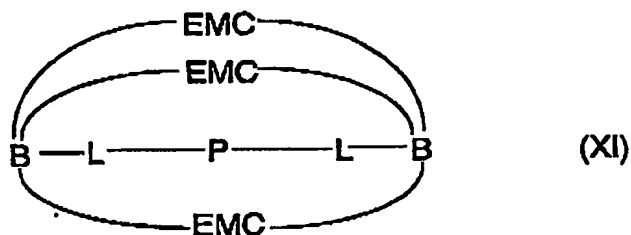
- B:** branching unit (basic unit, core) containing at least one branching center Bc and at least two branching functional groups Bfg connected to or capable of reacting with an encapsulating unit EMC;
- EMC:** encapsulating molecular chain;
- L:** linker containing at least one functional group Lfg which is connected to the protein P or capable of connecting with functional groups present on the protein P under the formation of a chemical bond;
- l:** 1, 2, 3, 4, 5, 6, 7, 8 or 9, preferably 1, 2, 3, 4 or 5, in particular 1, 2 or 3;
- P:** pharmacologically active protein.

The MPICs according to the general formula (IX) show a 1:1 ratio protein/macrocylic structure. The MPICs are soluble in water. In general, the cavity of the macrocylic structure also comprises water, in addition to the protein.

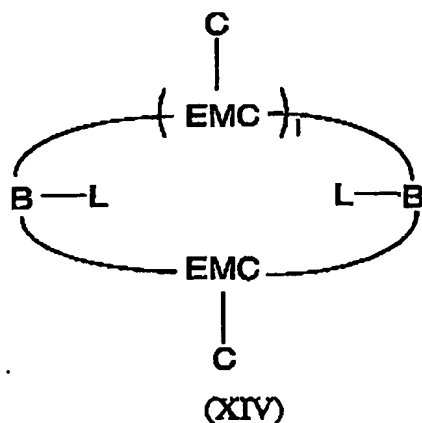
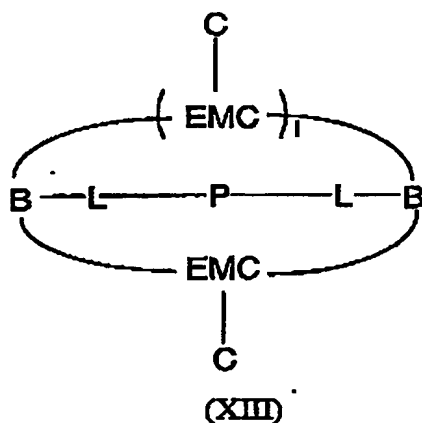
The EMCs contain hydrophilic groups, in an appropriate ratio and amount with respect to hydrophobic groups which may be present in the macrocylic structure, to render the latter water soluble.

B, EMC, L and P are in accordance with the definitions given above.

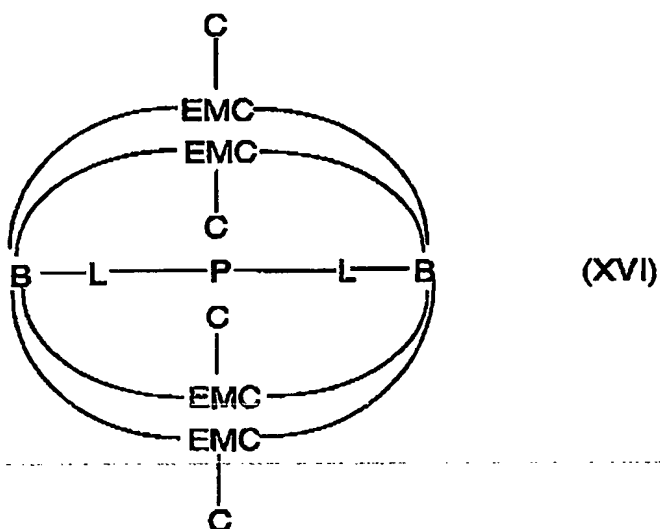
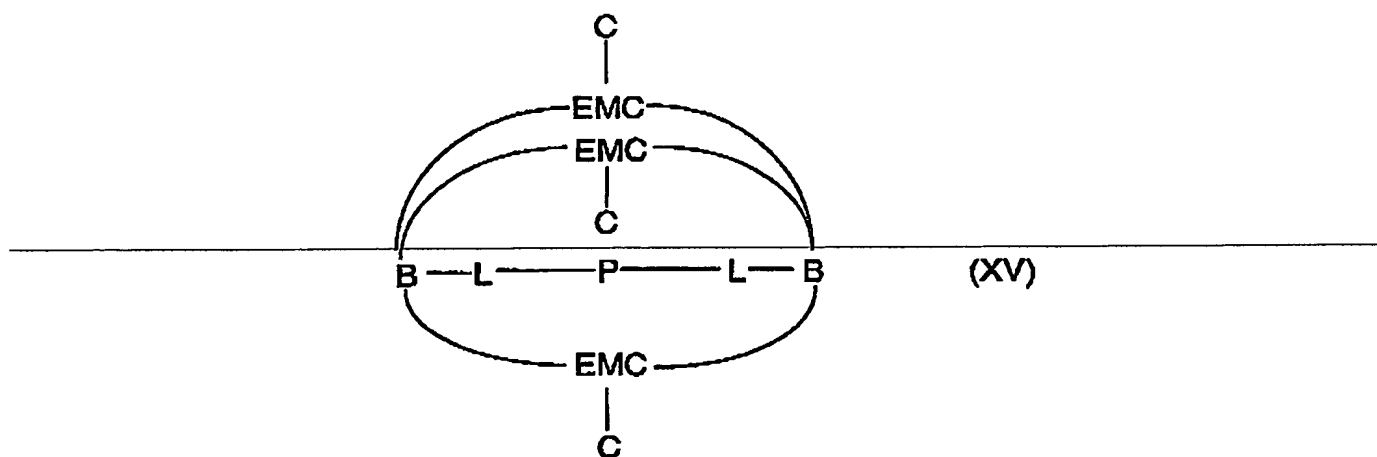
The macrocylic structure and the MPIC according to the present invention comprise at least two EMCs (l=1). The macrocylic structure can however comprise 3 (l=2), 4 (l=3), 5 (l=4), 6 (l=5) or even up to 10 EMCs. Some embodiments are depicted in the formulae (XI) and (XII) below.



The EMCs can carry capping groups C, as defined above, on their backbone. Appropriate capping groups are sterically demanding groups. The macrocyclic structures of the present invention are not rigid, and the EMCs, due to the rotation around the bonds connecting them to the branching units B, may swing to one side, resulting in a staggering and crowding on one side of the macrocyclic structure, and opening of one or more sides of the cavity. Capping groups prevent sterical proximity of the EMCs. If the EMCs come too close to one another, insufficient protection of the encapsulated protein from the attack of enzymes, antibodies or the like may result. Furthermore, the protein may leave the cavity (if the linker is broken) through the gap, resulting in an undesired release kinetics of the protein. In case capping groups are present, the following structures (XIII) and (XIV) result.

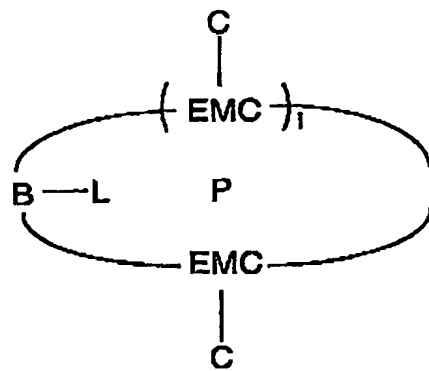


The figures (XV) and (XVI) below show macrocyclic structures having capping groups on the three and four EMCs of the respective macrocyclic structure.

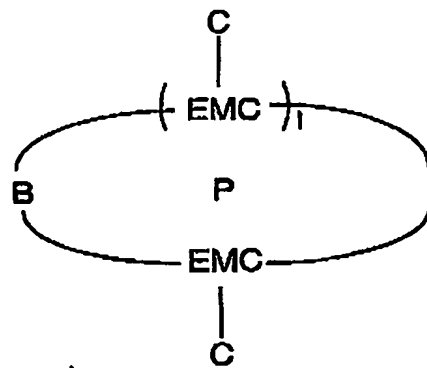


In all formulae (XI) to (XVI) shown beforehand, an EMC can carry one capping group C (as shown in the formulae) or more than one, i.e. 2, 3, 4, 5 or even more capping groups C.

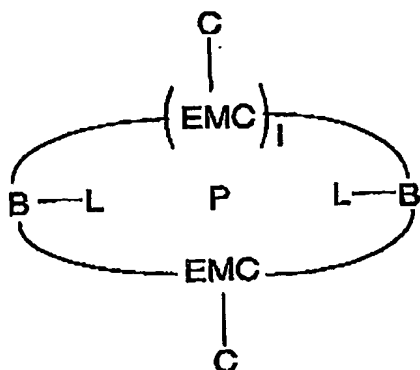
Examples for EPICs, DPICs and MPICs wherein the linker has been cleaved and the protein is not held within the cavity by covalent bonds are depicted in the formulae (XVII) to (XXI) below. Which include examples in which some of the EMCs within the B-C. domain are
represented by a single line and others by a double line to indicate the difference in the number of EMCs.



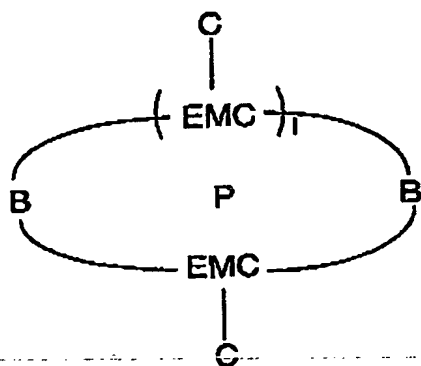
(XVII)



(XVIII)



(XIX)



(XX)

Modified proteins containing a linker are a part of the present invention. Within this embodiment, it is preferred if the linker is a prodrug linker or a traceless linker, more preferably traceless prodrug linker.

Another embodiment of the present invention is a protein in which the protein is not connected to the
linker.
Another embodiment of the present invention is a protein in which the protein is not connected to the
linker.
Another embodiment of the present invention is a protein in which the protein is not connected to the
linker.

The EPICs, MPICs and the DPICs of the present invention are synthesized from the protein and the EOC, macrocyclic structure and dendrimer, respectively, by a combination of solid-phase and solution synthesis methods known to the person skilled in the art.

The host molecule may be equipped with the linker moiety and be attached to the protein in one single reaction step (convergent synthesis). Alternatively, the linker-protein conjugate may be reacted with the branching unit B contained in the EOC, dendrimer or macrocyclic backbone structure. In another, divergent manifestation of the process, protein-linker-branching unit conjugate is reacted with presynthesized EOCs, macrocyclic structures or dendrimers. In an even more divergent approach, the dendrimers, macrocyclic structures or EOCs are assembled in a stepwise fashion in an extension of the central protein-linker-branching unit structure.

Efficient encapsulation is demonstrated by antibody binding studies. Antibodies against therapeutic proteins are high-affinity, high-selectivity probes. Steric shielding of the protein prevents access of the antibodies to the epitopes for molecular recognition. Antibody binding may be conveniently and reliably measured by methods known to the person skilled in the art, preferably immunoprecipitation or, as exemplified here, by label-free surface plasmon resonance scanning. In a study involving various insulin derivatives including insulins conjugated to different PEG reagents, and three monoclonal anti-insulins, complete prevention of antibody recognition was only achieved if insulin was complexed with a macrocyclic structure according to the present invention. In order to eliminate any bias by insulin release, non-cleavable covalent PEG or EOC conjugates were employed respectively.

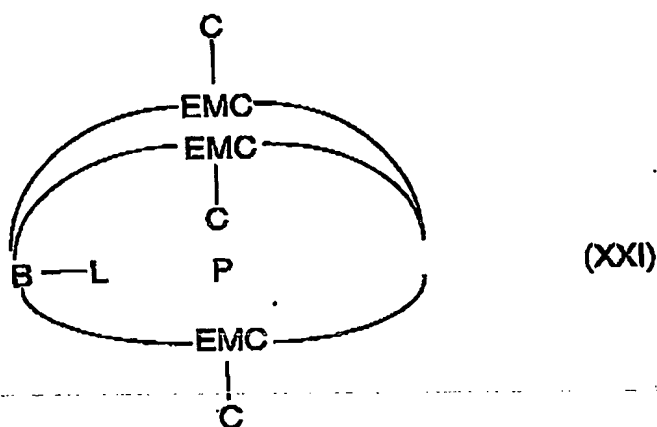
The resulting EOC self-organizes into a biomolecule-containing void by conformationally folding around the protein. This arrangement may be driven by sterical constraints or by chemical reactions or both.

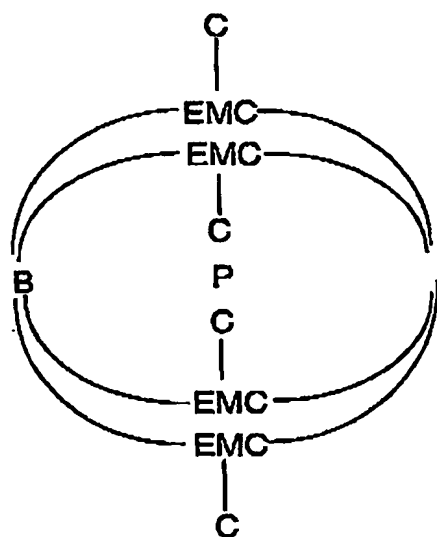
Eventually, the linker is cleaved, and a host-guest complex is obtained. Linker cleavage may be performed in vitro to generate a non-covalent complex. Alternatively, linker cleavage may occur in a prodrug approach in vivo after administration. Dissociation kinetics of the complex

may be governed by linker hydrolysis or protein effusion through the molecular matrix of the EOC, dendrimer, or macrocyclic structure, or a combination of both.

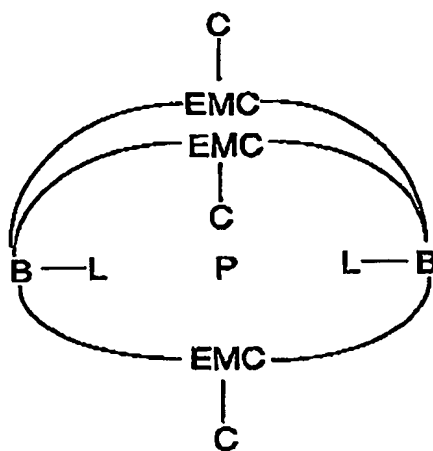
In the presence of the protein, the complex is characterized by a well-defined cavity-forming chemical structure and precise stoichiometry. After protein release, the host molecule may adopt various conformations due to its structural flexibility, for which reason protein release is essentially irreversible.

The formulae (XXI) to (XXIV) below show DPICs and MPICs wherein the bond between the linker and the protein has been cleaved; in two cases, traces of the linker remain at the EOC, dendrimer or macrocyclic structure, and in two cases the linker has totally been removed.

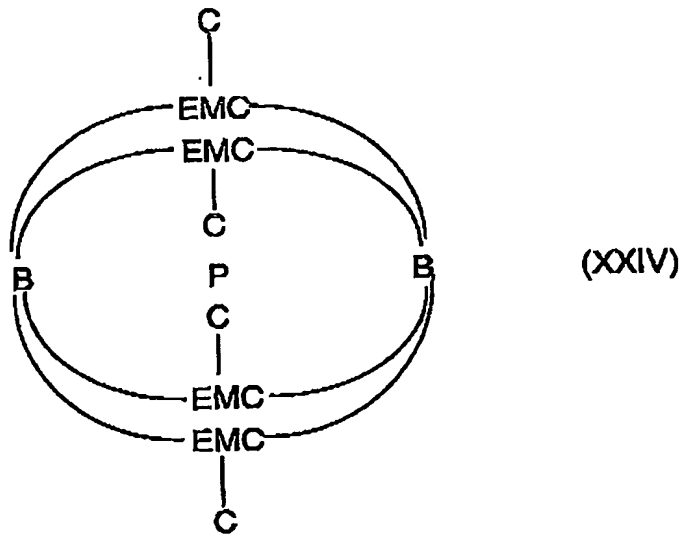




(XXII)



(XXIII)



The present invention also relates to method for selectively delivering a protein to a target, which method comprises

- providing an encapsulated protein;
- bringing the encapsulated protein into contact with a body liquid containing the target.

The encapsulated protein can be formulated into a drug, optionally together with one or more pharmaceutically acceptable carriers. The drug can contain one or more encapsulated protein types.

Drug containing at least one encapsulated protein according to the present invention and optionally one or more pharmaceutically acceptable carriers are also an object of the present invention

The present invention will now be illustrated by the following, non-limiting examples.

Examples

Materials and methods

Materials

Fmoc-amino acids, resins and TBTU were purchased from Novabiochem and are named according to the catalogue. Fmoc-Ado-OH was obtained from Neosystem (France) and Fmoc-PP-OH from Polypure (Norway). All additional chemicals were purchased from Sigma Aldrich. Human insulin was from ICN Biomedicals (USA). Maleimide-PEG5k, Maleimide-PEG20k and Maleimide-PEG2x20k were obtained from Nektar.

Reaction medium

Solid phase synthesis was performed on TentaGel TGR or Sieber amide resin with a loading of 0.2 mmol/g or 0.5 mmol/g, respectively. Syringes equipped with polypropylene frits were used as reaction vessels.

Standard coupling cycle for Fmoc-protected amino acids

For Fmoc protecting-group removal, resin was repeatedly (three times, 4 min each) agitated with 2/2/96 (v/v/v) piperidine/DBU/DMF and repeatedly (six times) washed with DMF. Coupling of Fmoc-protected amino acids to resin was achieved by agitating the resin with 3 equivalents (eq) of Fmoc-amino acid, 3 eq TBTU and 6 eq DIEA in DMF for 60 min. Finally, the resin was repeatedly (five times) washed with DMF.

Standard cleavage protocol for TentaGel TGR resin

Upon completed synthesis, resin was washed with DCM, dried in vacuo and treated with 95/5 (v/v) TFA/TES. After evaporation, compounds were purified by preparative RP-HPLC (Waters 600).

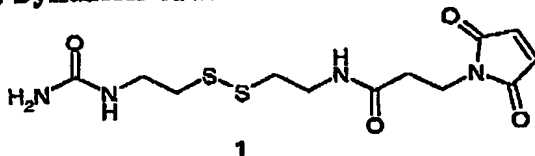
Analysis

Mass spectrometry (MS) was performed on a waters ZQ 4000 ESI instrument and spectra were, if necessary, interpreted by waters software MaxEnt.

Size exclusion chromatography were performed using a Waters 600 systems equipped with either a Superdex 75 or Superdex 200 column (Pharmacia) or manually using a PD10 column (Pharmacia).

I – Synthesis of bifunctional linkers

I-1) Synthesis of linker 1



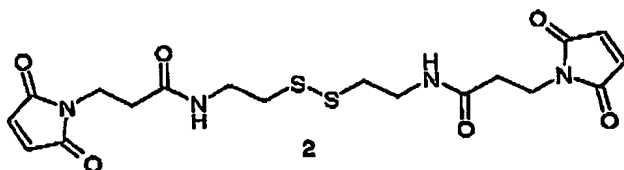
0.7 g TentaGel TGR resin was soaked in THF and incubated for 60 min with a solution of 4 ml of 0.5 M chloroformic acid-4-nitrophenyl ester and 0.5 M DIEA in THF, washed with THF and dried. 4 ml of a suspension of 0.3 M cystamine-dihydrochloride and 0.7 M DIEA in DMSO were added to the resin and agitated for 90 min. Resin was washed with DMSO and

DMF, and 3 eq maleimidopropionic acid and 3 eq DIC in DMF were added and agitated for 60 min.

After washing the resin with DMF and DCM, compound 1 was cleaved from the resin and purified by RP-HPLC.

MS (MW calculated) 1: 346 g/mol (346.4 g/mol)

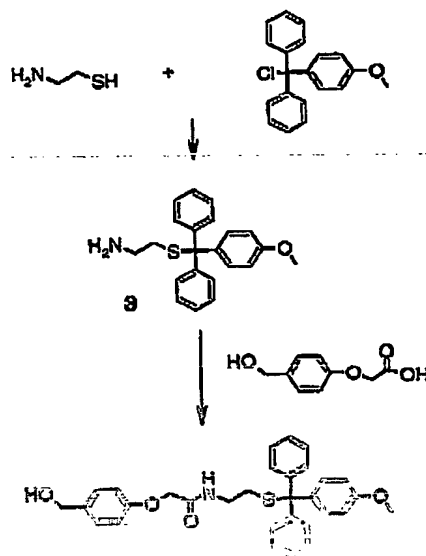
I-2) Synthesis of linker 2



Cystamine-dihydrochloride was suspended in 1/1 (v/v) DMSO/DMF and mixed with 2 eq maleimidopropionic acid, 2 eq DIC and 2 eq DIEA. The suspension was agitated for 2 h at room temperature (RT) and after acidification with formic acid, compound 2 was purified by RP-HPLC.

MS (MW calculated) 2: 454 g/mol (454.5 g/mol)

I-3) Synthesis of linker 5



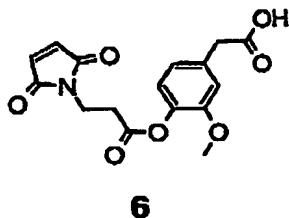
Cysteamine hydrochloride was dissolved in TFA and 0.5 eq Mmt-chloride were added. After 30 min, TFA was removed under nitrogen flow and the residue was taken up in pyridine. After adding a solution of 0.2 M Na_2CO_3 , product was extracted with ether and dried over Na_2SO_4 . Following filtration, solvents were removed using a rotary evaporator and compound 3 was obtained as a highly viscous oil.

3 was reacted with 4-hydroxymethyl-phenoxyacetic acid (HMPA) and 1 eq HOBT/DIC for 30 min. After purification by RP-HPLC, compound 4 was neutralized by DIEA and lyophilized.

4 was reacted with 5 eq chloroformic acid-4-nitrophenyl ester and 10 eq DIEA in dioxane for 2 h. Subsequent purification by RP-HPLC gave product 5.

MS $[\text{MNa}]^+$ (MW+Na calculated) 5: 700 g/mol (701 g/mol)

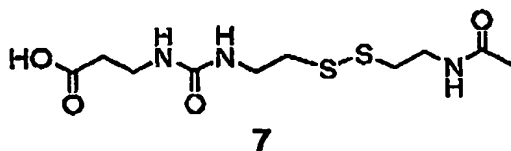
I-4) Synthesis of linker 6



300 mg of 4-hydroxy-3-methoxy-phenylacetic acid and 570 μl DIEA were dissolved in 5 ml DCM and added to 0.5 g 2-chlorotrityl-chloride resin (1.5 mmol/g). The suspension was agitated for 1 h at RT and resin was washed with DCM. Resin was resuspended in a solution of 190 mg 3-maleimidopropionic acid, 333 mg MSNT and 73 μl N-methyl imidazole in 3 ml DCM and agitated for 1h. After washing of the resin with DCM, cleavage was performed by agitation of the resin for 30 min in 10 ml 4/1 (v/v) DCM/TFA. Solvent was removed under nitrogen flow and compound 6 was purified by RP-HPLC.

MS $[\text{MNa}]^+$ (MW+Na calculated): 356 g/mol (356 g/mol)

I-5) Synthesis of linker 7

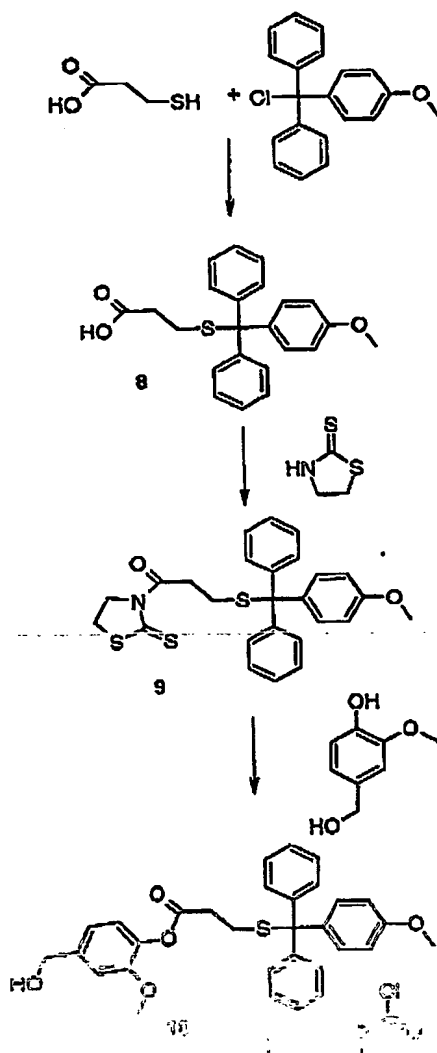


512 mg Fmoc-3-aminopropionic acid and 570 μl DIEA were dissolved in 5 ml DCM and added to 0.5 g 2-chlorotrityl-chloride resin (1.5 mmol/g). The suspension was agitated for 1 h at RT and resin was washed with DCM. After Fmoc-removal with 96/2/2 (v/v/v) DMF/DBU/piperidine and washing of the resin with THF, 4 ml of a solution of 0.5 M chloroformic acid-4-nitrophenyl ester and 0.5 M DIEA in THF were incubated with the resin

for 30 min. Resin was washed with THF and DMF and agitated for 30 min with a solution of 1 M cystamine in DMF and washed with DMF. Subsequently, resin was agitated for 15 min in a solution of 2/1/1 (v/v/v) DMF/acetic anhydride/pyridine. After washing the resin with DMF and DCM, compound 7 was cleaved by agitation for 30 min in 10 ml of 4/1 (v/v) DCM/TFA. After evaporation of solvent under nitrogen flow, product 7 was purified by RP-HPLC.

MS (MW calculated): 309 g/mol (309 g/mol)

I-6) Synthesis of linker 11



Mmt-chloride (1 eq) and mercaptopropionic acid (1.1 eq) were taken up in TFA and incubated for 30 min. TFA was removed under nitrogen flow. The product was dissolved in pyridine, diluted in water, acidified by acetic acid and extracted in ether. The ether phase was separated and dried over Na_2SO_4 . Solvent was removed and product **8** was purified by RP-HPLC.

Compound **8** and DMAP (2.5 eq) were dissolved in dry DCM and 2 eq of EDC HCl were added at 0 °C. The solution was stirred for 14 h at RT and washed with sodium acetate buffer (0.25 M, pH 4.5). The organic phase was dried over Na_2SO_4 , concentrated and compound **9** was purified by silica gel column chromatography using heptane/acetic acid ester (1/1) as mobile phase.

MS (MW calculated) **9**: 479 g/mol (479.7 g/mol)

Compound **9**, 4-hydroxy-3-methoxy benzylalcohol (7 eq) and DMAP (7 eq) were refluxed in DCM for 2 h under nitrogen atmosphere. After neutralization with acetic acid, the solution was concentrated and compound **10** was purified by RP-HPLC:

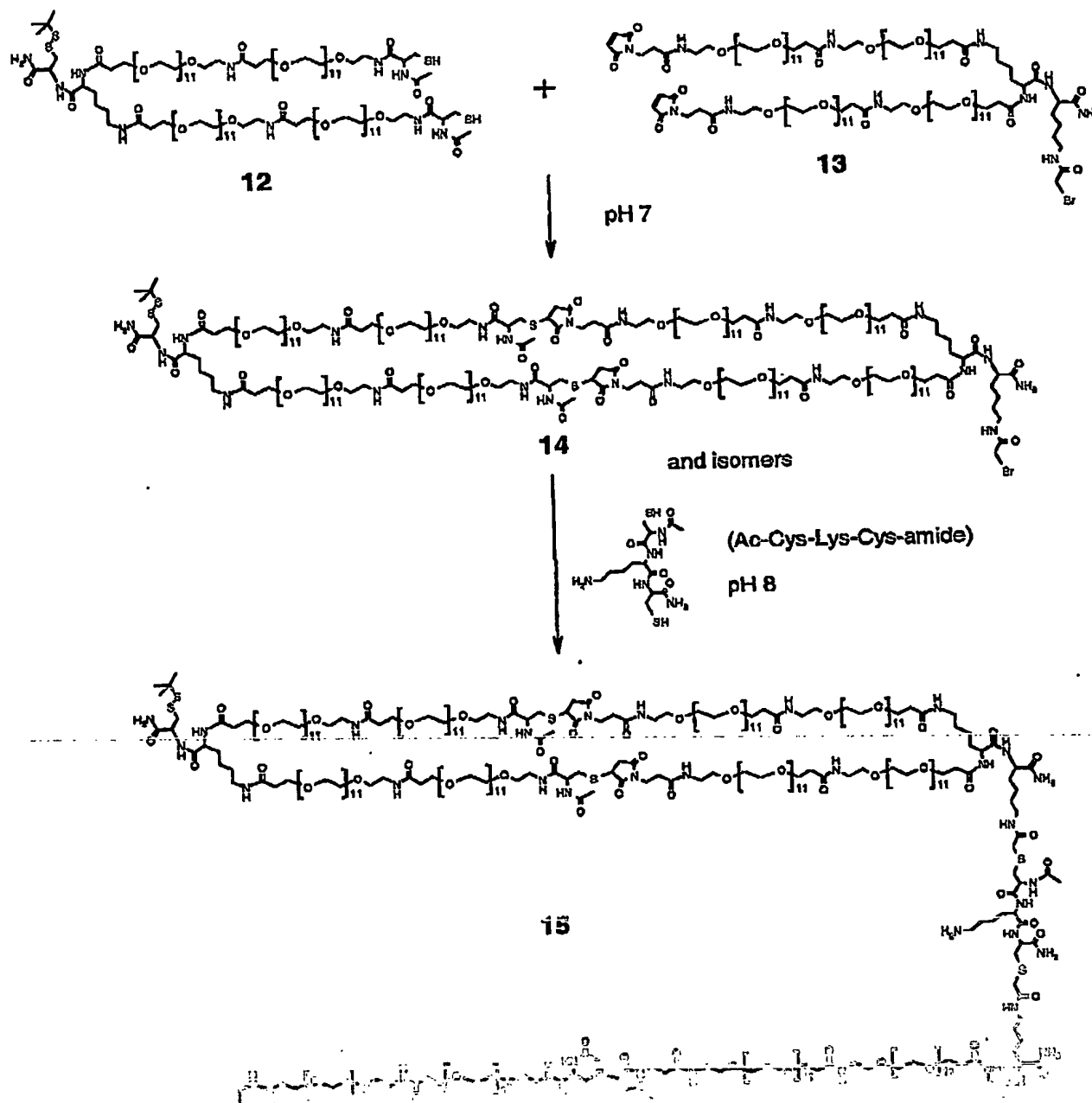
MS $[\text{M}+\text{Na}]^+$ (MW+Na calculated) **10**: 537 g/mol (537.6 g/mol)

Compound **10**, chloroformic acid-4-nitrophenyl ester (10 eq), and DIEA (20 eq) were stirred in dry dioxane for 3 h at 40 °C under nitrogen atmosphere. After addition of acetic acid (25 eq) the mixture was concentrated and compound **11** purified by RP-HPLC:

MS $[\text{M}+\text{Na}]^+$ (MW+Na calculated) **11**: 702 g/mol (702.7 g/mol)

II - Synthesis of bifunctional macrocyclic structures

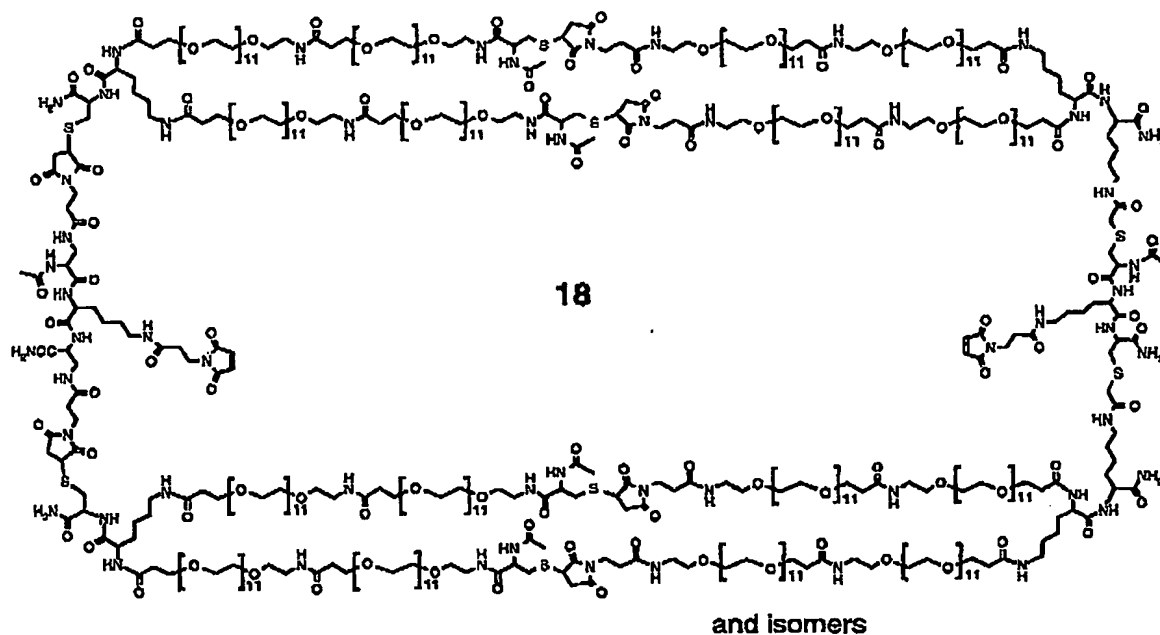
II-1) Synthesis of bis-maleimido-macrocyclic structure 18





and isomers





Employing the standard protocol for solid-phase synthesis, the amino acids Fmoc-Cys(S-tBu)-OH, Fmoc-Lys(Fmoc)-OH, two units of Fmoc-PP-OH and Fmoc-Cys(Trt)-OH were coupled to TGR resin. After final Fmoc-removal, resin was treated with 2/1/1 (v/v/v) DMF/acetic anhydride/pyridine for 15 min. Compound 12 was cleaved from the resin and purified by RP-HPLC.

MS (MW calculated) 12: 3024.8 g/mol (3025.8 g/mol)

Employing the standard protocol for solid-phase synthesis, the amino acids Fmoc-Lys(Mtt)-OH, Fmoc-Lys(Fmoc) and two units of Fmoc-PP-OH were coupled to TGR resin. After final Fmoc removal, the resin was treated for 30 min with 5 eq maleimidopropionic acid and 5 eq DIC in DMF. Removal of the Mtt-protecting group was achieved by repeated washing of the resin with 99/1 (v/v) DCM/TFA until the solution remained colorless. Resin was washed with DCM, 1% DIEA in DMF (briefly) and DMF. Subsequently, resin was incubated for 25 min with a solution of 0.5 M bromoacetic acid and 0.5 M DIC in DMF. After cleavage, compound 13 was purified by RP-HPLC.

MS (MW calculated) 13: 3095.2 g/mol (3095.5 g/mol)

12 and 13 were mixed at an equimolar ratio and concentration was adjusted to 35 μ M by addition of 0.1 M sodium TFA. The pH of the solution was adjusted to 7.5 with 0.1 M NaOH and the solution was filtered. The solution was then used for the experiment.

was adjusted to 8.0 with phosphate buffer. After 90 min the reaction was quenched by acidification with acetic acid, and product **15** was purified by RP-HPLC.

MS (MW calculated) **15**: 12472 g/mol (12474.2 g/mol)

S-tBu protecting groups in **15** were removed by reduction with 100 mM DTT in phosphate buffer (pH 8.0). After 3 h the pH of the solution was adjusted to 2.0, and compound **16** was purified by RP-HPLC.

MS (MW calculated) **16**: 12295 g/mol (12297.8 g/mol)

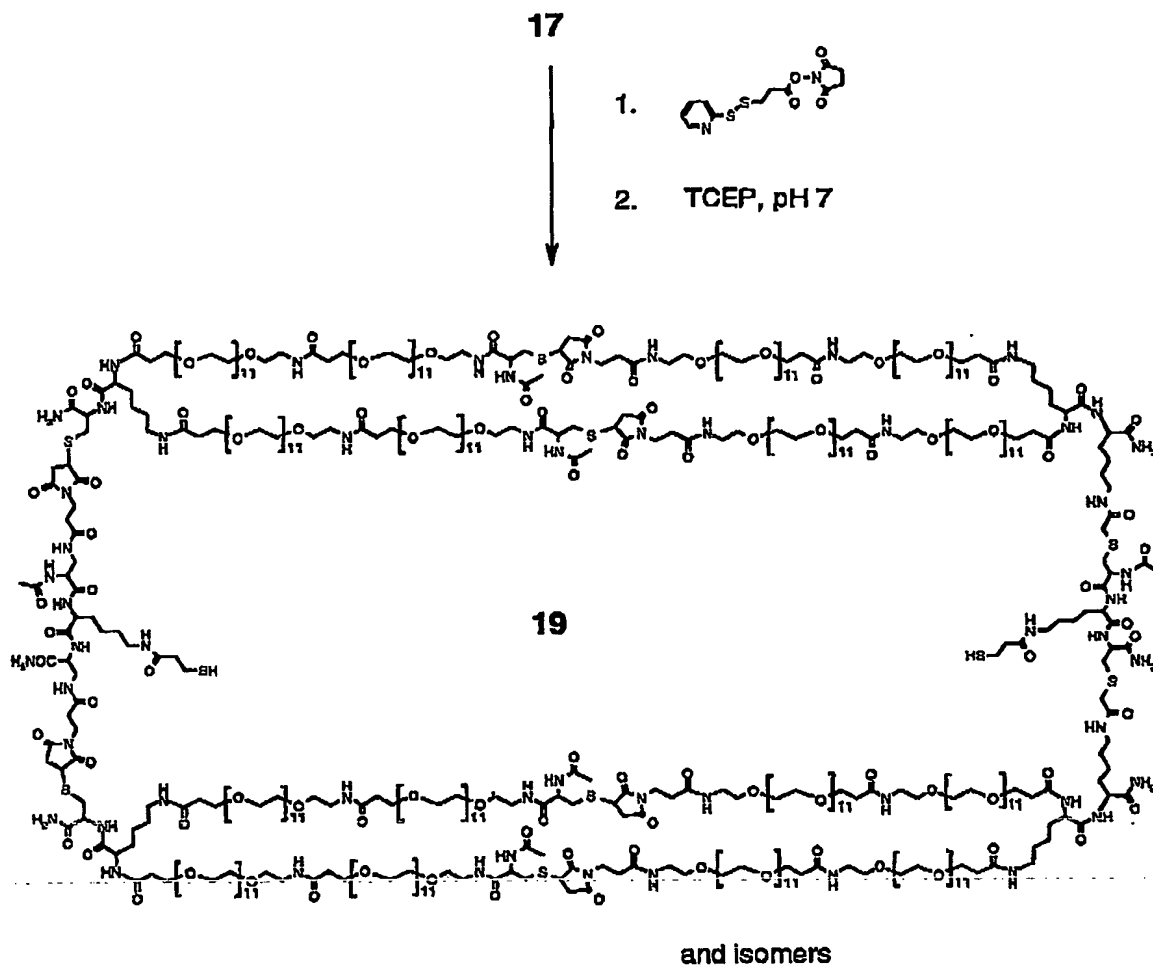
16 was mixed with 1 eq Ac-Dpr(Mal)-Lys-Dpr(Mal)-NH₂ (obtained according to the standard protocol for solid phase synthesis using Fmoc-Dpr(ivDde)-OH) and concentrations were adjusted by addition of water to 15 µM. Subsequently, pH was adjusted to 7.5 with 0.1 M phosphate buffer (pH 7.5) and stirred for 20 min. The pH of the solution was brought to 2.0 by formic acid, and compound **17** was purified by RP-HPLC and lyophilized.

MS (MW calculated) **17**: 12955 g/mol (12959.5 g/mol)

17 was dissolved in DMF and agitated for 30 min with a solution of 20 eq maleimidopropionic acid, 30 eq DIEA and 20 eq DIC in DMF. Subsequently, the solution was acidified with formic acid, diluted with water and **18** was purified by RP-HPLC and lyophilized.

MS (MW calculated) **18**: 13257 g/mol (13261.7 g/mol)

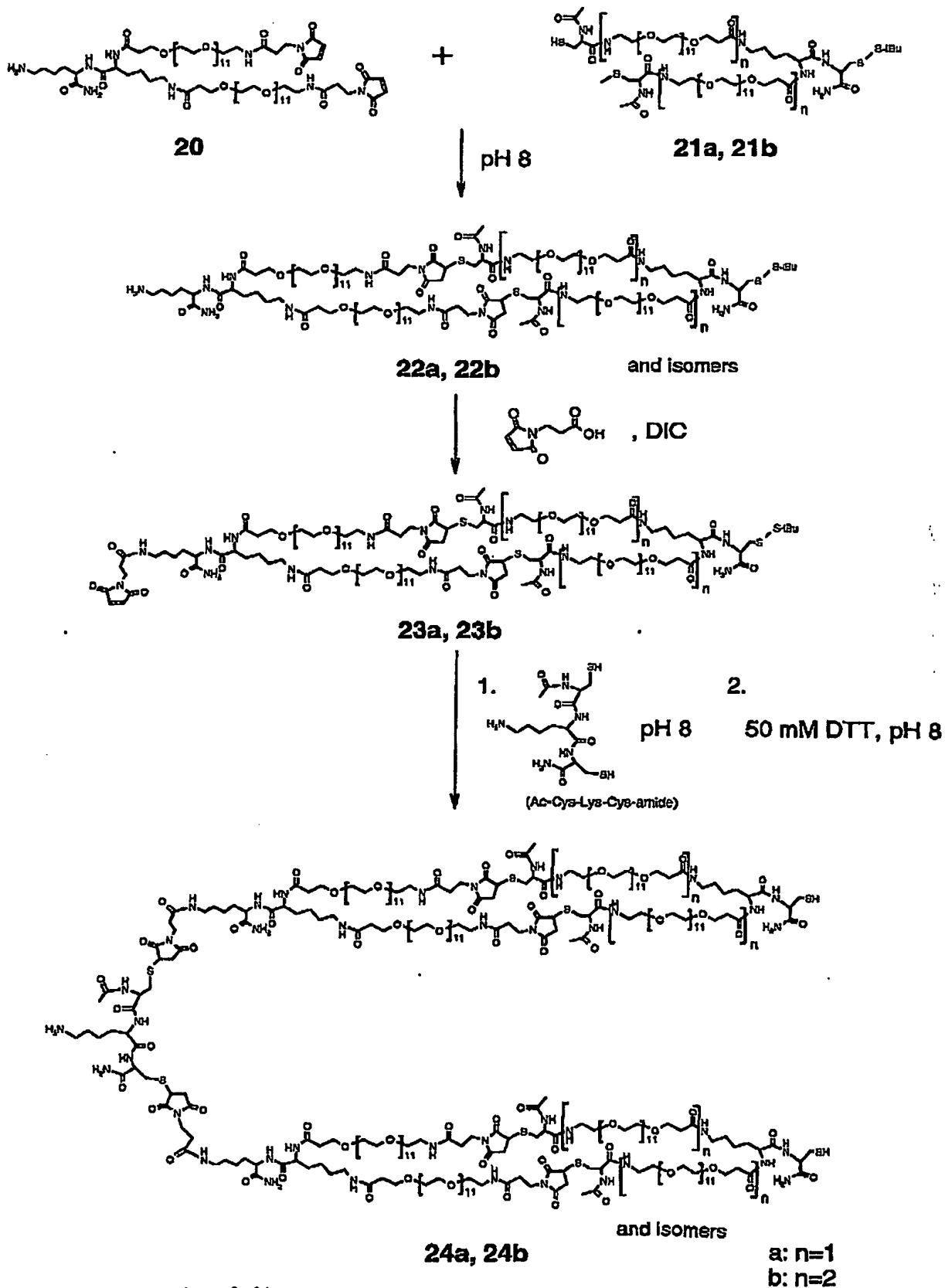
II-2) Synthesis of bis-thiol-macrocyclic structure 19



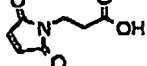
17 was adjusted to a concentration of 300 μM in 0.1 M phosphate buffer (pH 7.0) and 15 eq SPDP in DMSO were added. The resulting suspension was agitated for 20 min at RT. 10 mM TCEP were added, and the cocktail was agitated for another 20 min at RT. Product **19** was purified by RP-HPLC.

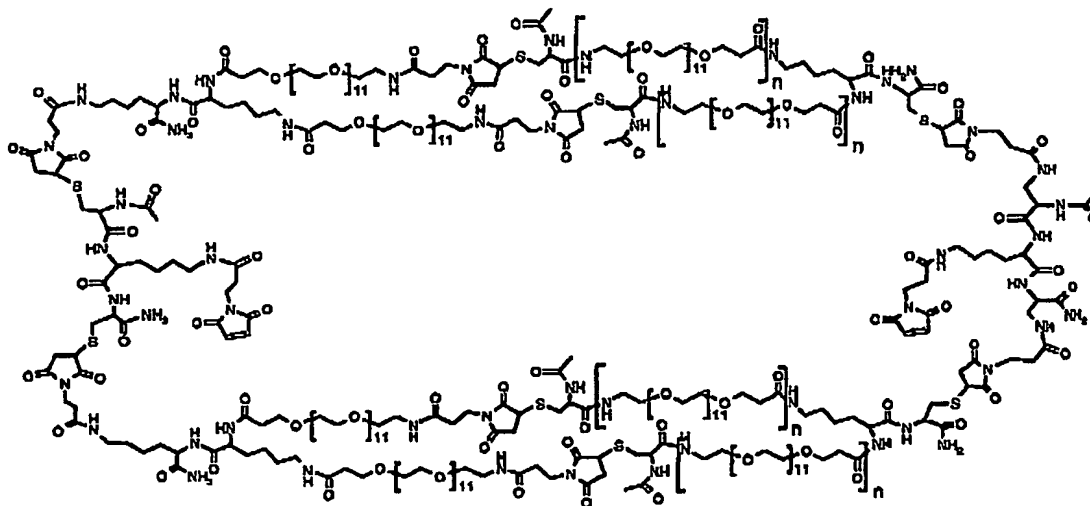
MS (MW calculated) **19**: 13132 g/mol (13133,7 g/mol)

II-3) Synthesis of bis-maleimido-macrocylic structures 25a and 25b



1. Ac-Dpr(Mal)-Lys-Dpr(Mal)-amide, pH 8

2.  , DIC



25a, 25b

and isomers

a: n=1
b: n=2

Fmoc-Lys(Mtt)-OH, Fmoc-Lys(Fmoc)-OH and Fmoc-PP-OH were coupled according to the standard protocol for solid phase synthesis. After Fmoc removal, resin was incubated with 6 eq maleimidopropionic acid and 6 eq DIC for 30 min. Following cleavage from resin, compound 20 was purified by RP-HPLC.

MS (MW calculated) 20: 1774 g/mol (1775 g/mol)

Compounds 21a and 21b were obtained by solid phase synthesis according to protocol II-1) for product 12.

MS (MW calculated) 21a: 1825 g/mol (1826 g/mol)
MS (MW calculated) 21b: 1825 g/mol (1826 g/mol)

MS (MW calculated) 22a: 3600 g/mol (3601 g/mol)

MS (MW calculated) 22b: 4800 g/mol (4801 g/mol)

Compounds 22a or 22b, respectively, were dissolved in DMF and pH was adjusted to 8.0 with DIEA. 6 eq Maleimidopropionic acid and 6 eq DIC were added and mixtures were incubated for 30 min to yield compounds 23a and 23b, respectively. Purification was performed by RP-HPLC.

MS (MW calculated) 23a: 3752 g/mol (3753 g/mol)

MS (MW calculated) 23b: 4950 g/mol (4952 g/mol)

2 eq of compound 23a or 23b respectively were mixed with 1 eq Ac-Cys-Lys-Cys-amide (obtained according to the standard protocol for solid phase synthesis). After adjusting the pH to 8.0 by addition of 0.5 M phosphate buffer (pH 8.0) the solution was agitated for 10 min. The reaction was quenched by addition of 10 eq DTT. After lyophilization the residue was taken up in 1:1 (v/v) acetonitrile/50 mM phosphate buffer (pH 8.0). Removal of the S-tBu protecting group by reduction with 50 mM DTT for 2 h yielded products 24a and 24b, respectively. Purification was by RP-HPLC.

MS (MW calculated) 24a: 7719 g/mol (7722 g/mol)

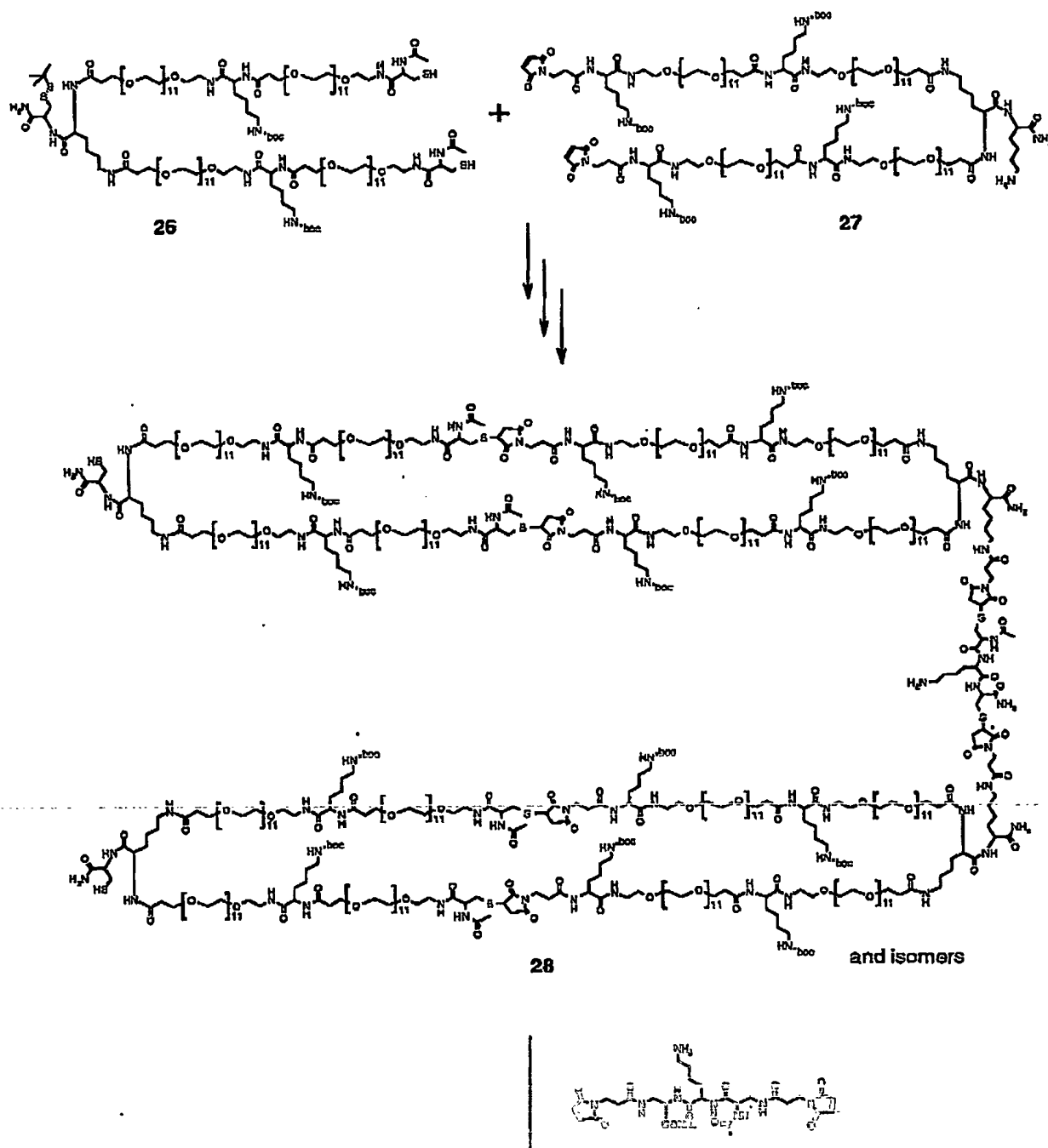
MS (MW calculated) 24b: 10120 g/mol (10121 g/mol)

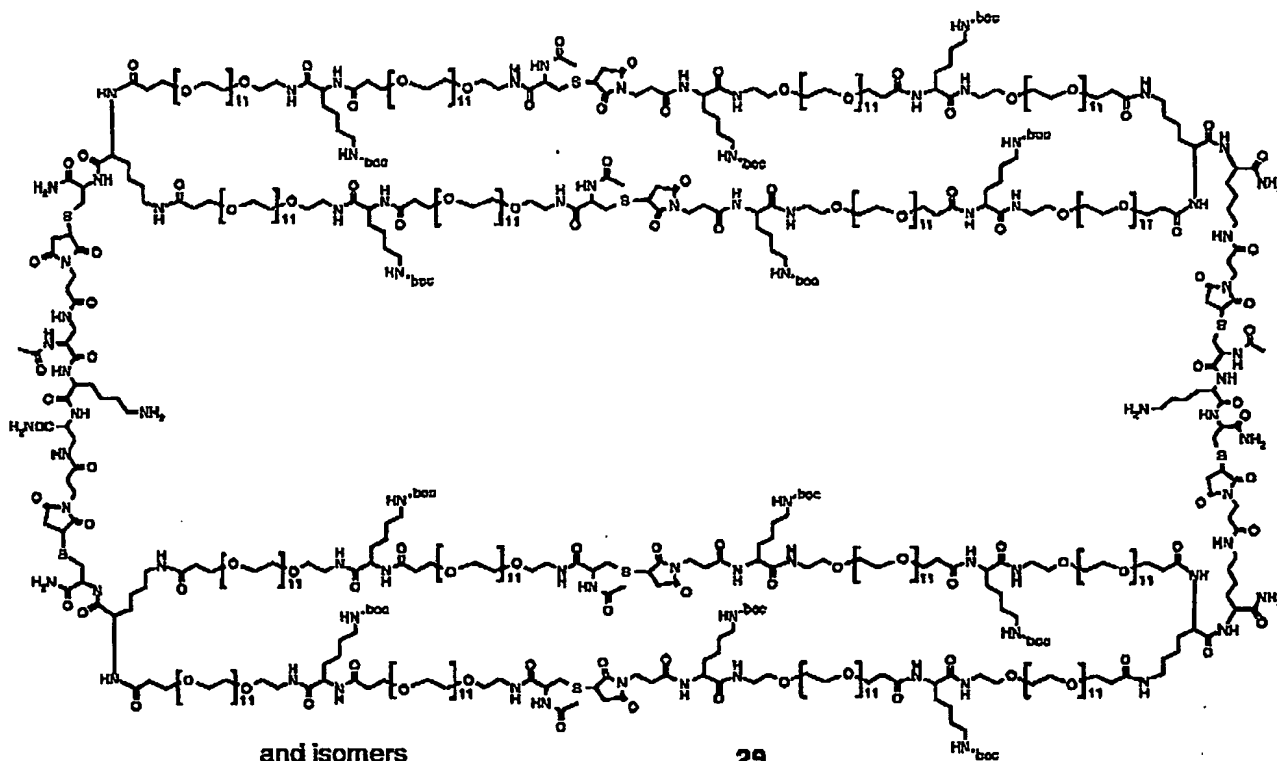
Compounds 24a or 24b, respectively, were subjected to two additional reaction steps to yield 25a or 25b, respectively, according to protocol II-1 and in analogy to the reaction of compounds 16 and 17 to product 18.

MS (MW calculated) 25a: 8686 g/mol (8686 g/mol)

MS (MW calculated) 25b: 11085 g/mol (11085 g/mol)

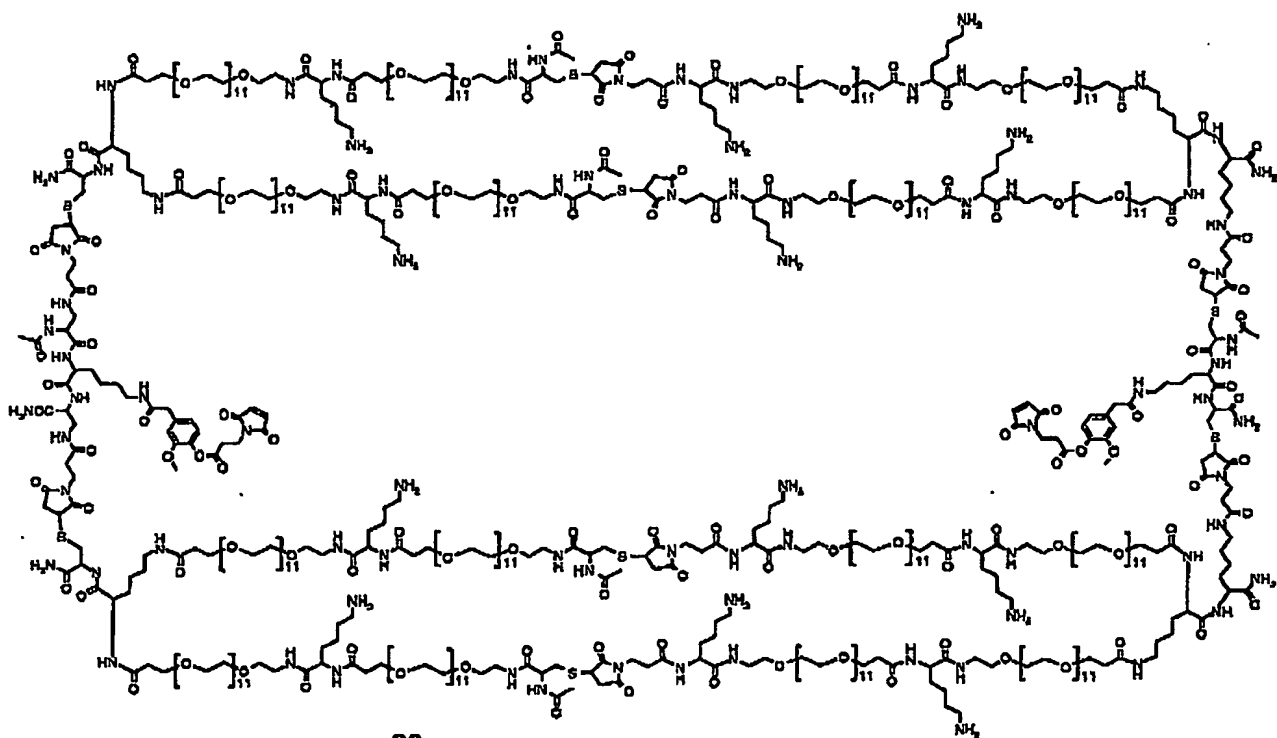
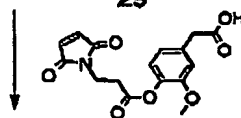
II-4) Synthesis of bis-maleimido-core-macrocylic structure 31



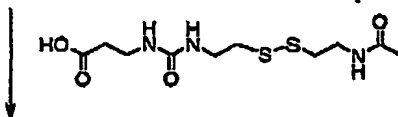


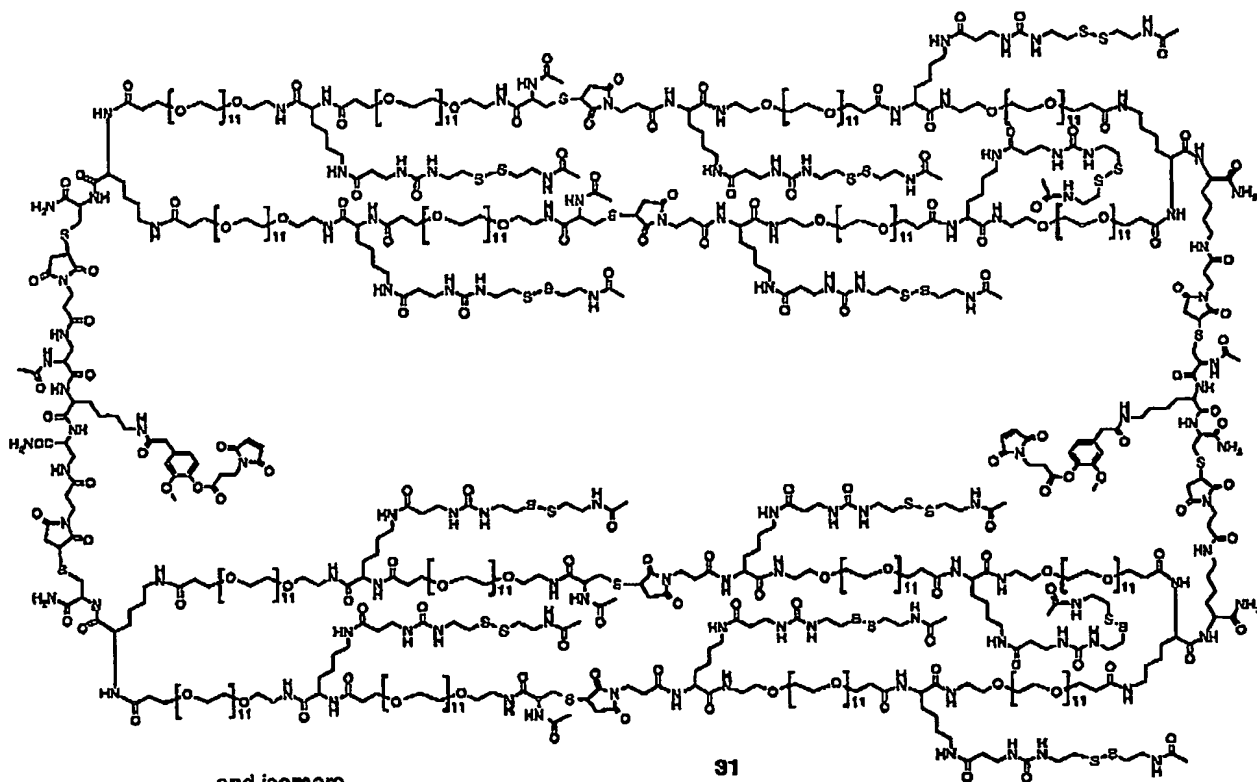
and isomers

29



30





and isomers

Compound 26 was obtained according to the standard protocol for solid phase synthesis. The amino acids Fmoc-Cys(StBu)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-PP-OH, Fmoc-Lys(Boc)-OH, Fmoc-PP-OH and Fmoc-Cys(Mmt) were coupled to Sieber amide resin. After final Fmoc-removal, resin was treated with a solution of 2/1/1 (v/v/v) DMF/acetic acid anhydride/pyridine for 15 min, washed with dichloromethane and dried in vacuo. Cleavage was performed by repeated treatment (15 times) of the resin for 2 min with a solution of 97/1/2 (v/v/v) dichloromethane/TFA/TES. Collected supernatant was mixed and buffered with 1 eq pyridine (vs. TFA). After concentrating the mixture, compound 26 was purified by RP-HPLC.

MS (MW calculated) 26: 3482 g/mol (3482 g/mol)

Compound 27 was obtained according to the standard protocol for solid phase synthesis. Starting from Sieber amide resin, the amino acid sequence Fmoc-Lys(Mmt)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-PP-OH, Fmoc-Lys(Boc)-OH, Fmoc-PP-OH and Fmoc-Lys(Boc)-OH was assembled. After final Fmoc-removal, the resin was reacted for 30 min with 5 eq

Compound 28 was obtained from educts 26 and 27 in analogy to the synthesis of 24 from educts 20 and 21 according to protocol II-3.

Cyclization of 28 to 29 was performed in analogy to cyclization of compound 16 according to protocol II-1.

For analysis, a sample was treated with TFA to effect removal of the Boc-protecting groups.

MS (MW calculated without Boc-groups) 29: 14717 g/mol (14720 g/mol)

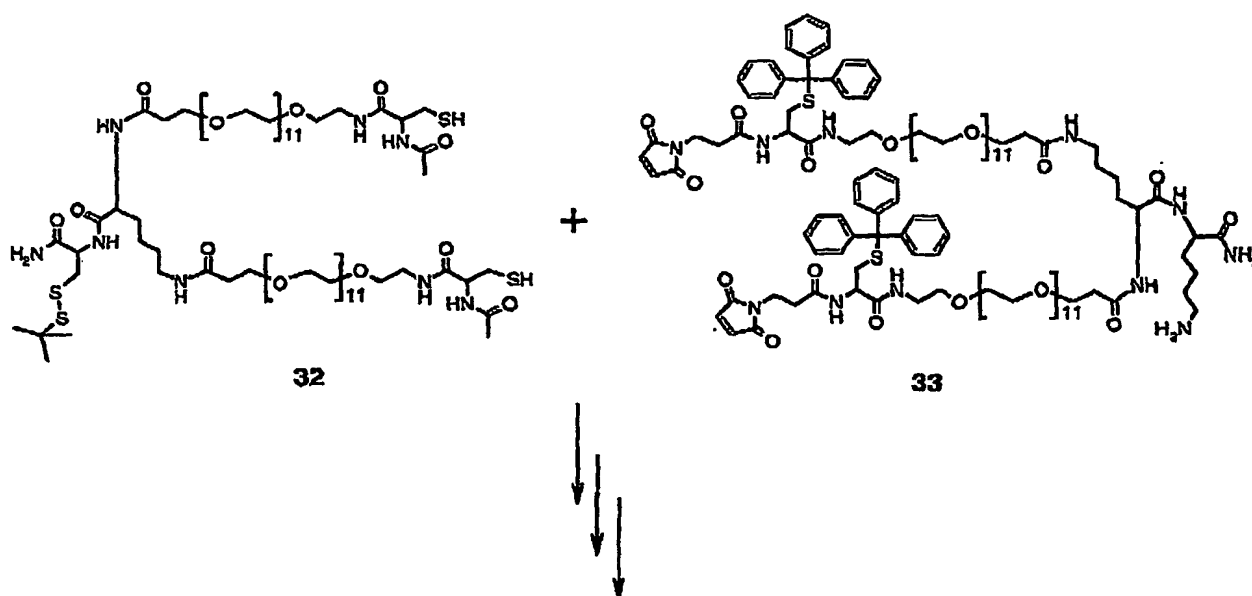
29 were reacted with 10 eq 6 and 10 eq DIC in DMF for 30min. Product was purified by RP-HPLC and lyophilized. Boc-protecting groups were removed by incubation for 30 min in 1/1 (v/v) TFA/DCM. Solvent was evaporated under nitrogen flow. Extraction of the residue with water and subsequent lyophilization yielded 30.

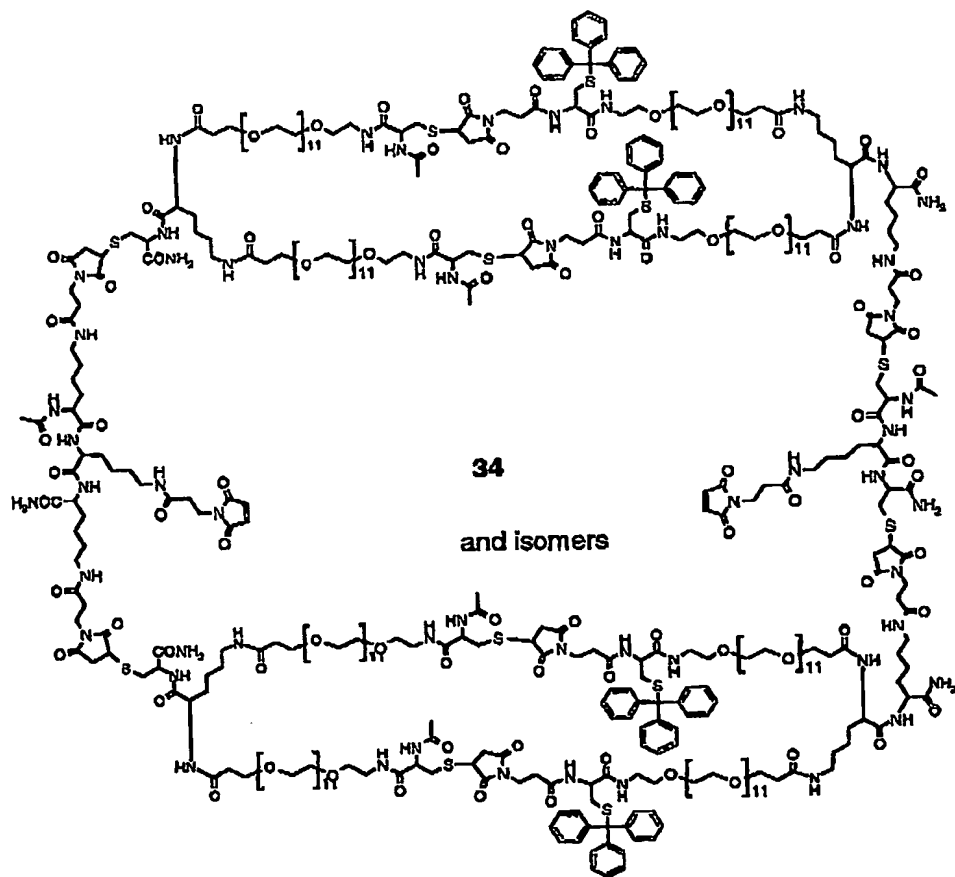
MS (MW calculated): 15352 g/mol (15360 g/mol)

Product 31 was obtained by reacting 30 with 30 eq 7 and 30 eq DIC in DMF for 45 min. Product was purified by RP-HPLC and lyophilized.

MS (MW calculated): 18851 g/mol (18847 g/mol)

II-5) Synthesis of bis-maleimido-core-macrocyclic structure 34





Compound 32 was obtained according to the standard protocol for solid phase synthesis. The amino acids Fmoc-Cys(StBu)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-PP-OH and Fmoc-Cys(Mmt)-OH were coupled to Sieber amide resin. After final Fmoc-removal, the resin was incubated with a solution of 2/1/1 (v/v/v) DMF/acetic acid anhydride/pyridine for 15 min, washed with dichloromethane, and dried in vacuo. Cleavage from the resin was afforded by treatment with 97/1/2 (v/v/v) Dichloromethane/TFA/TES for 30 min. After concentration, product 32 was purified by RP-HPLC

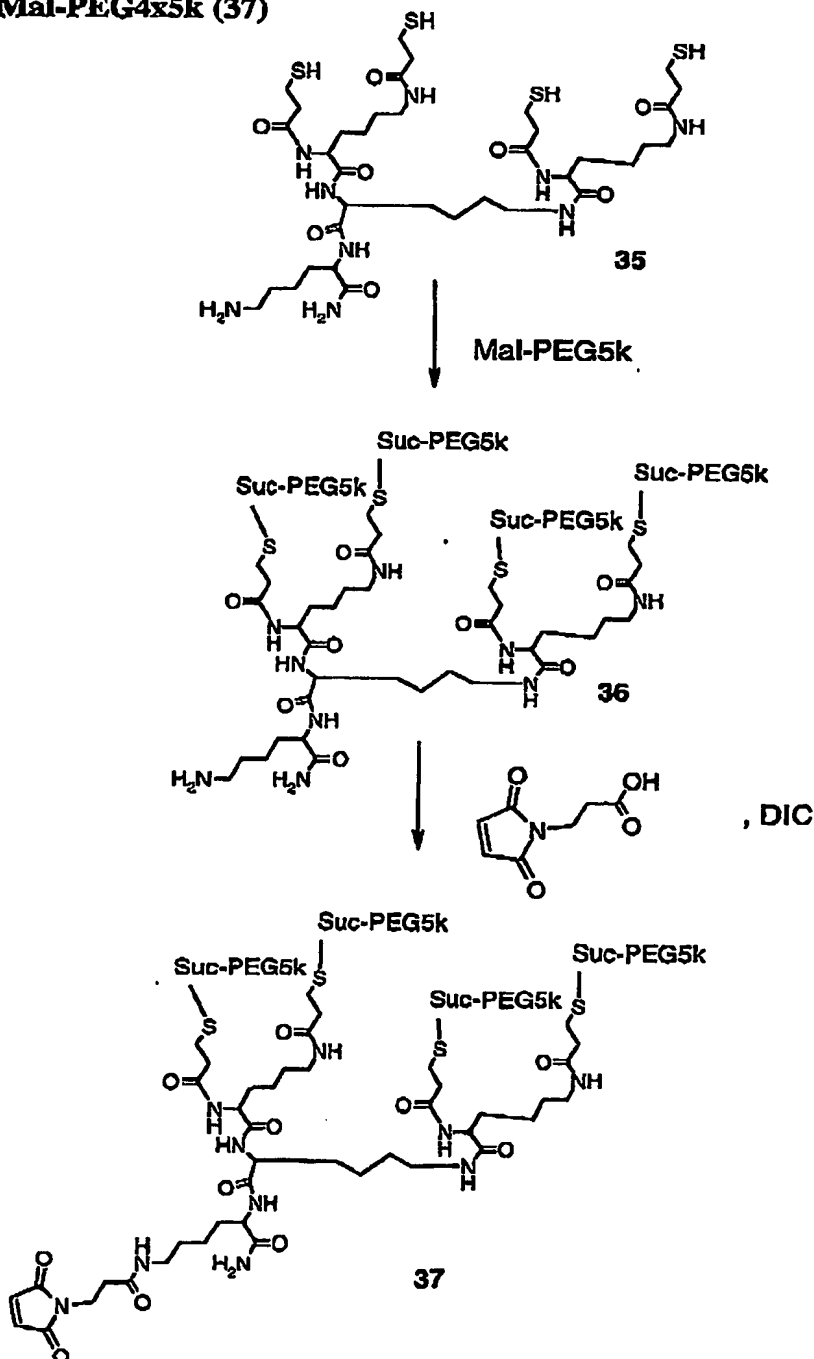
Compound 33 was obtained according to the standard protocol for solid phase synthesis. The amino acids Fmoc-Lys(Mtt)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-PP-OH and Fmoc-Cys(Trt)-OH were coupled with Sieber amide resin. After final Fmoc-removal, the resin was incubated with 6 eq maleimidopropionic acid and 6 eq DIC for 30 min, washed with dichloromethane, and dried in vacuo. Cleavage from the resin was afforded by treatment with 99/1/ (v/v) dichloromethane/TFA for 15 min and repeated washing with DCM. Pooled eluates were lyophilized with 0.1% acetic acid (vacuum, TFA) and solvent was removed in vacuo. Product was purified by RP-HPLC.

Synthesis of **34** was performed in analogy to the cyclization of compound **25** according to protocol II-3, except for the use of Ac-Lys (Mal)-Lys-Lys(Mal)-NH₂ instead of Ac-Dpr(Mal)-Lys-Dpr(Mal)-NH₂. For analysis, a sample of **34** was treated with 48/50/2 (v/v/v) TFA/DCM/TES for 10 min to effect removal of the Trt-protecting groups.

MS (MW calculated without Trt groups) **34**: 9183 g/mol (9183 g/mol)

III – Synthesis of proteophore-capping reagents

III-1) Synthesis of Mal-PEG4x5k (**37**)



Compound 35 was obtained according to the standard protocol for solid phase synthesis. The amino acids Fmoc-Lys(Boc)-OH, Fmoc-Lys(Fmoc)-OH and Fmoc-Lys(Fmoc)-OH were coupled to TGR resin, and Mmt-3-mercaptopropionic (8) acid was used as the terminal building block. After resin cleavage, product was purified by RP-HPLC.

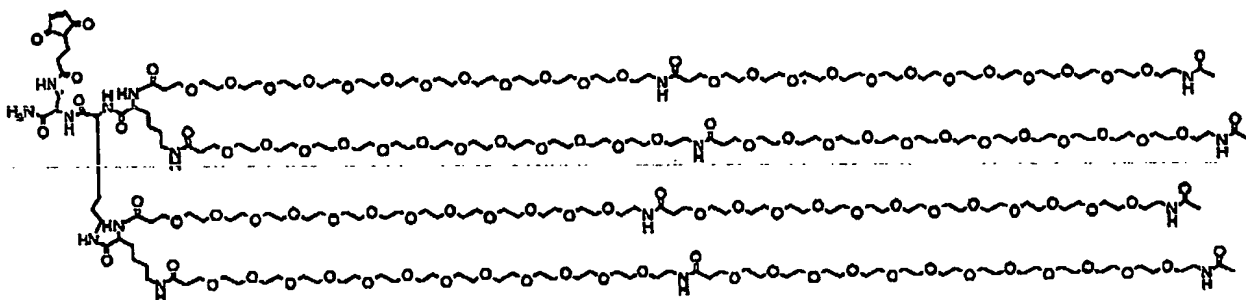
MS (MW calculated) 35: 881.5 g/mol (882 g/mol)

Compound 35 and 4.1 eq Maleimide-PEG5k were dissolved in 0.1 M sodium phosphate buffer (pH 7.0) and stirred for 30 min. Excess Maleimide-PEG5k was reacted with mercaptoethanol and product 36 was purified by RP-HPLC and lyophilized.

Compound 36 was taken up in DMF, 6 eq maleimidopropionic acid and 6 eq DIC in DMF were added and the mixture was agitated for 30 min. The product was purified by RP-HPLC and characterized by size exclusion chromatography (Superdex 200 column, flow rate: 0.75 ml/min)

SEC (retention time) 36: 16 min

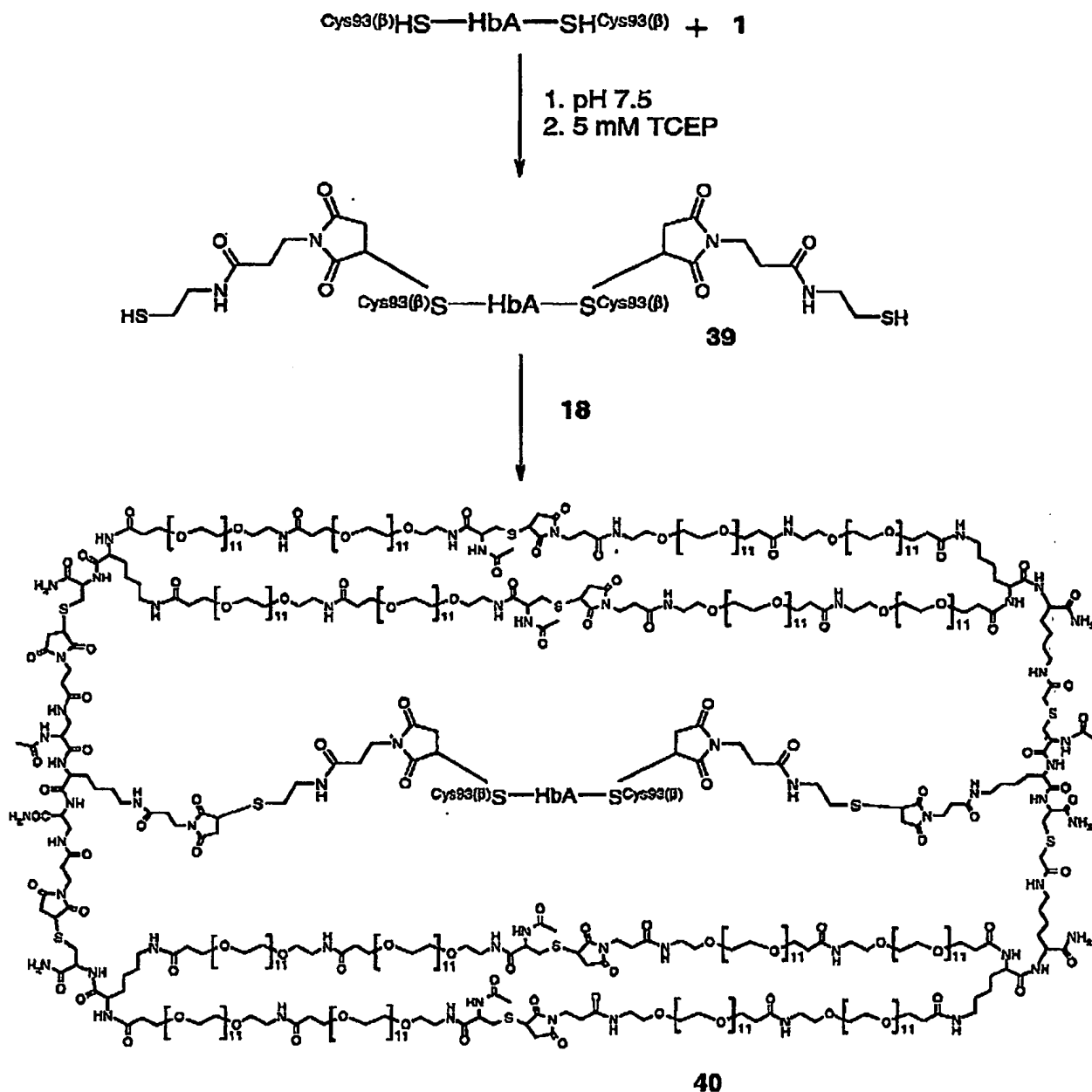
III-2) Synthesis of Mal-PP4x1.25k (38)



38

Compound 38 was obtained according to the standard protocol for solid phase synthesis. The amino acids Fmoc-Dpr(ivDde)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-Lys(Fmoc)-OH and two units of Fmoc-PP-OH were coupled to TGR resin. After final Fmoc-removal, resin was incubated with a solution of 2/1/1 (v/v/v) DMF/acetic acid anhydride/pyridine for 15 min. Removal of the ivDde-protecting group was afforded by repeatedly agitating (three times) the resin for 5 min with 98/2 (v/v) DMF/hydrazine. The resin was washed with DMF and treated for 24 hrs with a solution of 6 eq maleimidopropionic acid and 6 eq DIC in DMF.

IV - Synthesis of a Hemoglobin MPIC



IV-1) Modification of Hb Cys93(β) with linker 1

Human hemoglobin (Hb) was adjusted to a concentration of 10 mg/ml in 0.1 M sodium phosphate buffer (pH 7.5), 5 eq 1 were added and the mixture was agitated for 30 min at RT. Excess 1 was removed by size exclusion chromatography (PD10 column).

MS alpha-subunit, unmodified (MW calculated): 15121 g/mol (15127 g/mol)
MS beta-subunit (MW calculated): 16208 g/mol (16214 g/mol)

Cleavage of the disulfide bond of the conjugated linker was effected by reduction of the modified Hb for 30 min in 5 mM TCEP (pH 7.5). Product **39** was purified by SEC (Superdex 200).

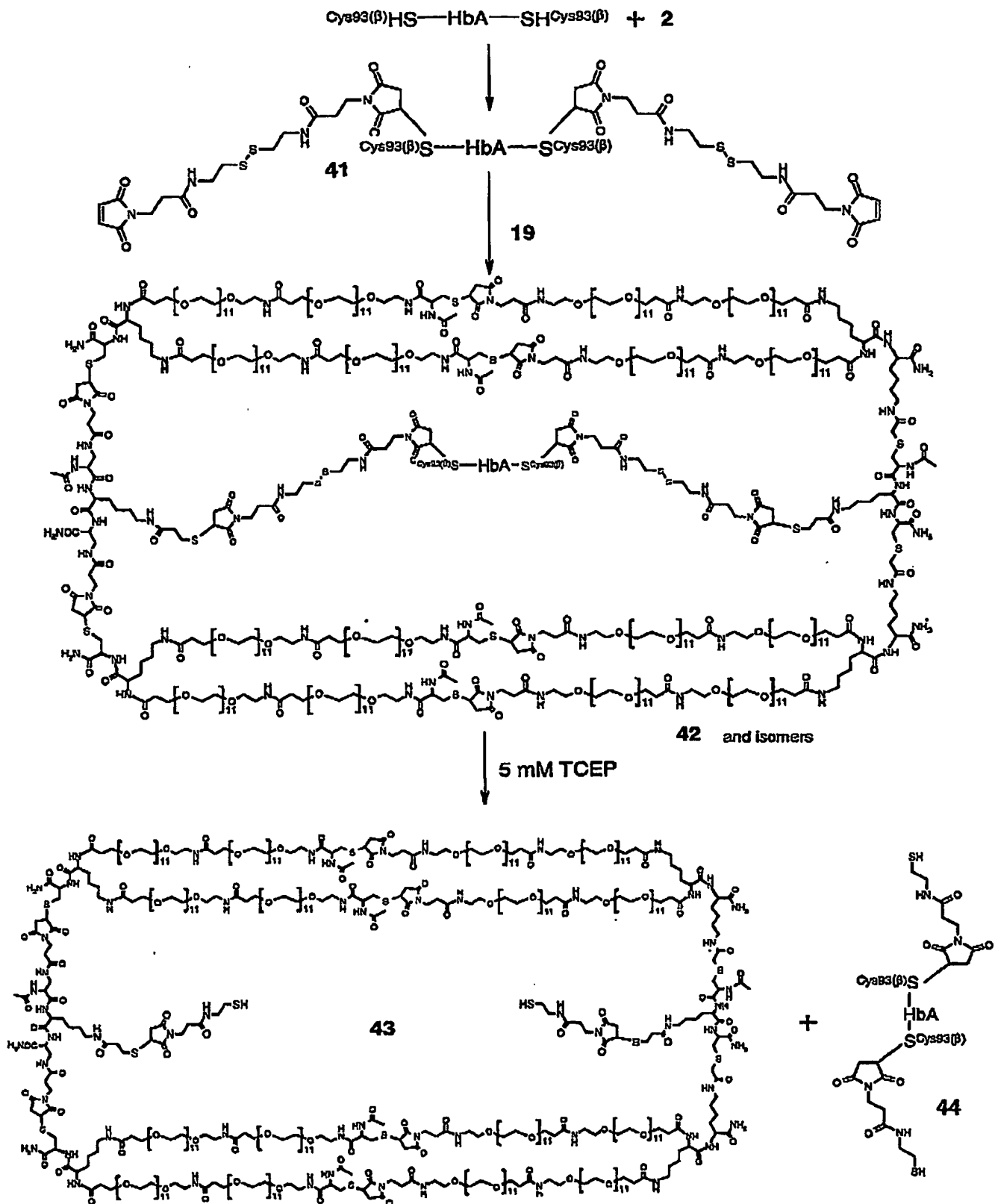
MS alpha subunit, unmodified (MW calculated) **39**: 15121 g/mol (15127 g/mol)
MS beta subunit (MW calculated) **39**: 16090 g/mol (16096 g/mol)

IV-2) Conjugation to bis-maleimido-macrocyclic structure 18

Compound **39** was adjusted to a concentration of 20 μ M in 0.1 M phosphate buffer (pH 7.5) and 2 eq of **18** were added. After incubation for 20 min at RT, the resulting Hb MPIC **40** was purified by SEC (Superdex 200 column). Figure 1 displays size exclusion chromatograms of native Hb, **39**, the reaction mixture of **39** and Hb, and purified product **40**.

MS alpha subunit, unmodified (MW calculated) **40**: 15122 g/mol (15127 g/mol)
MS crosslinked beta subunits (MW calculated) **40**: 45432 g/mol (45454 g/mol)

V - Synthesis of a cleavable Hemoglobin MPIC



V-1) Modification of Hb Cys93(β) with linker 2

Hb was adjusted to a concentration of 10 mg/ml in 0.1 M sodium phosphate buffer (pH 7.5). After addition of 10 eq 2 the solution was agitated for 30 min at RT. Hb-conjugate 41 was purified by SEC (Superdex 200).

MS alpha subunit, unmodified (MW calculated) 41: 15123 g/mol (15127 g/mol)
MS beta subunit (MW calculated) 41: 16318 g/mol (16322 g/mol)

V-2) Conjugation to bis-thiol-macrocylic structure 19

The concentration of 41 was adjusted to 20 μ M in 0.1 M phosphate buffer (pH 7.5). After addition of 2 eq 19 the solution was incubated for 45 min at RT. The Hemoglobin MPIC 42 was purified by SEC (Superdex 200).

MS alpha subunit (MW calculated) 42: 15125 g/mol (15127 g/mol)
MS beta subunit (MW calculated) 42: 45776 g/mol (45780 g/mol)

In order to prove the reversibility of the conjugation, product 42 was treated with 5 mM TCEP in 0.1 M phosphate buffer (pH 7.5). The quantitative release of Hb from the MPIC was assessed by LC/MS.

MS beta subunit (MW calculated) 42: 16091 g/mol (16096 g/mol)

Figure 2 displays size exclusion chromatograms of 41, 42 and of Hb (44) released from 42.

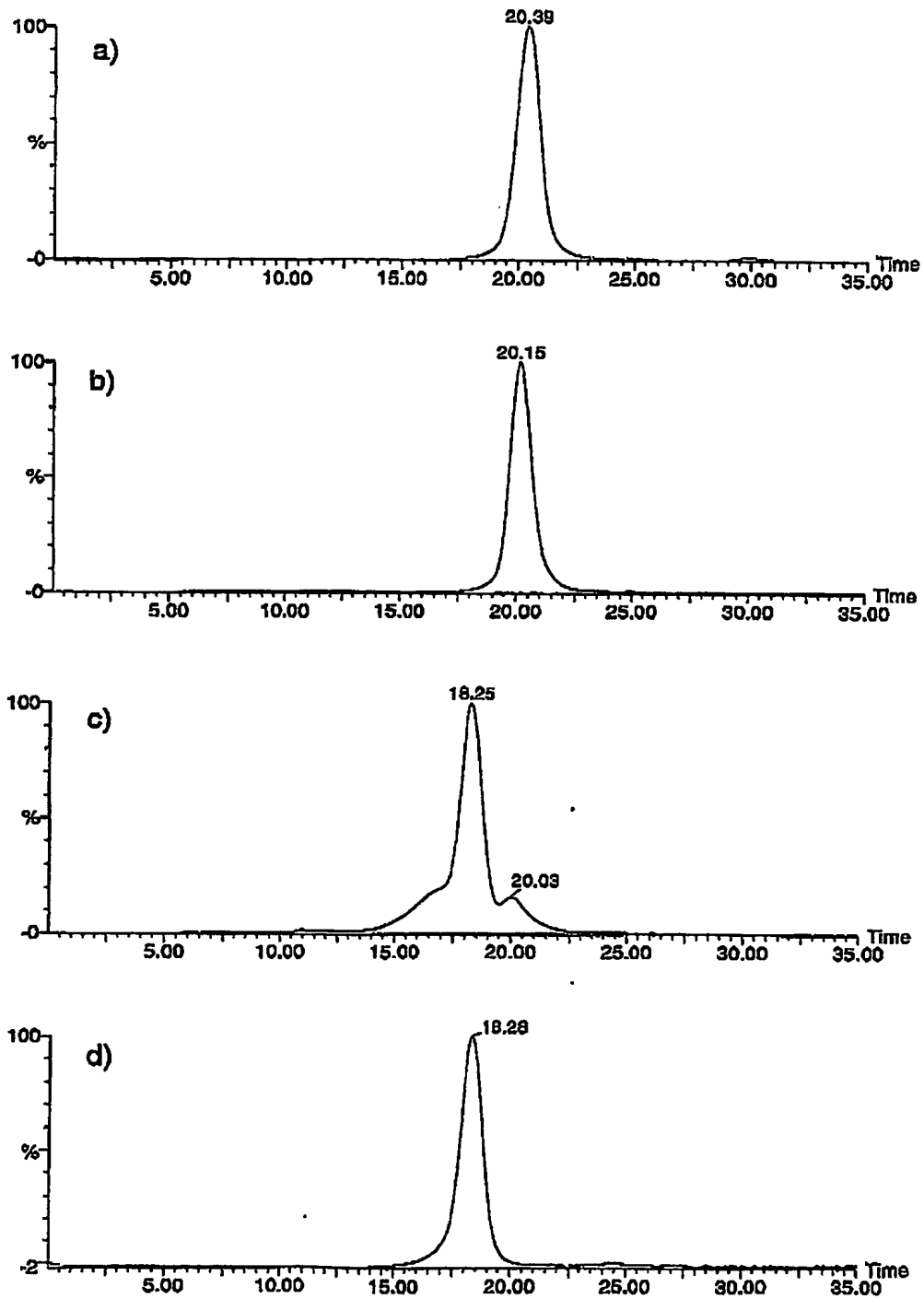


Fig. 1 : Size exclusion chromatograms of a) native Hb, b) 39, c) reaction mixture 40, and d) purified covalent Hemoglobin MPIC 40. UV signals were recorded at 280 nm.

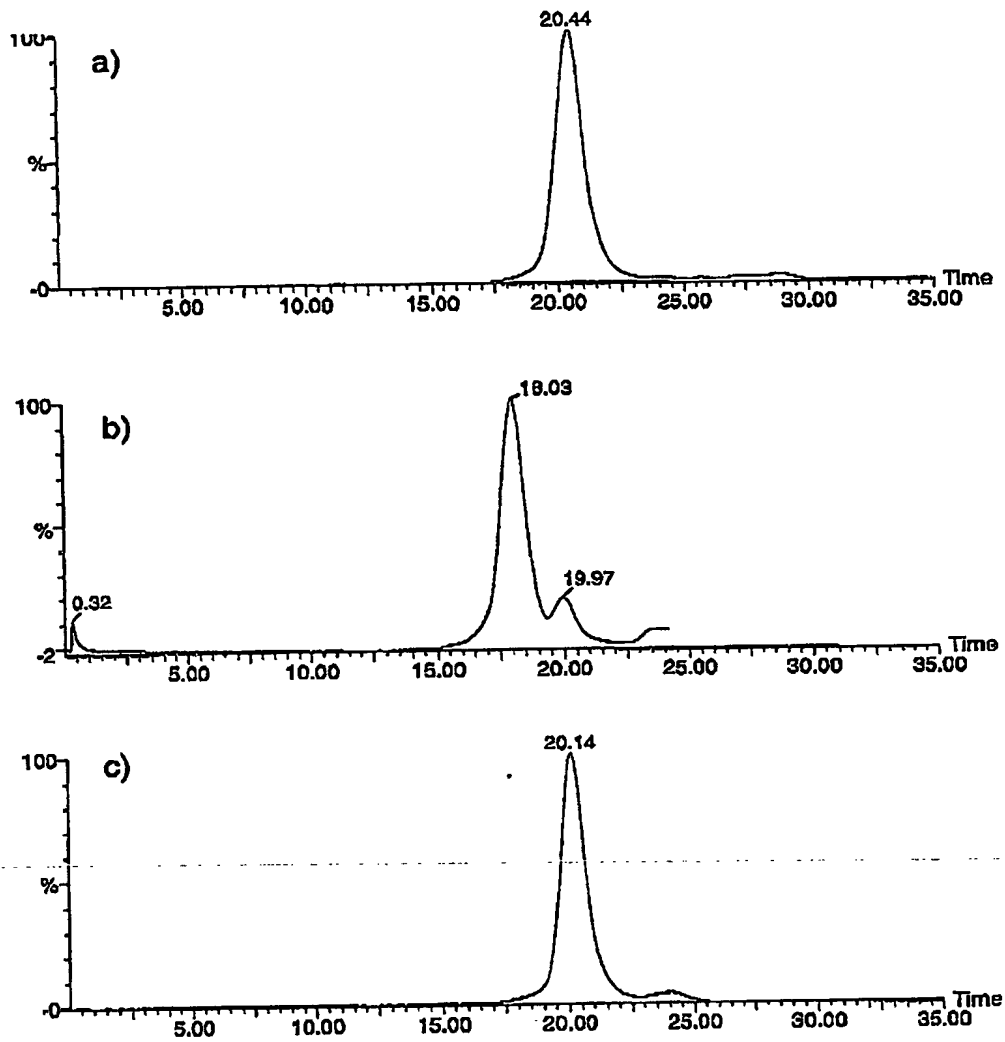
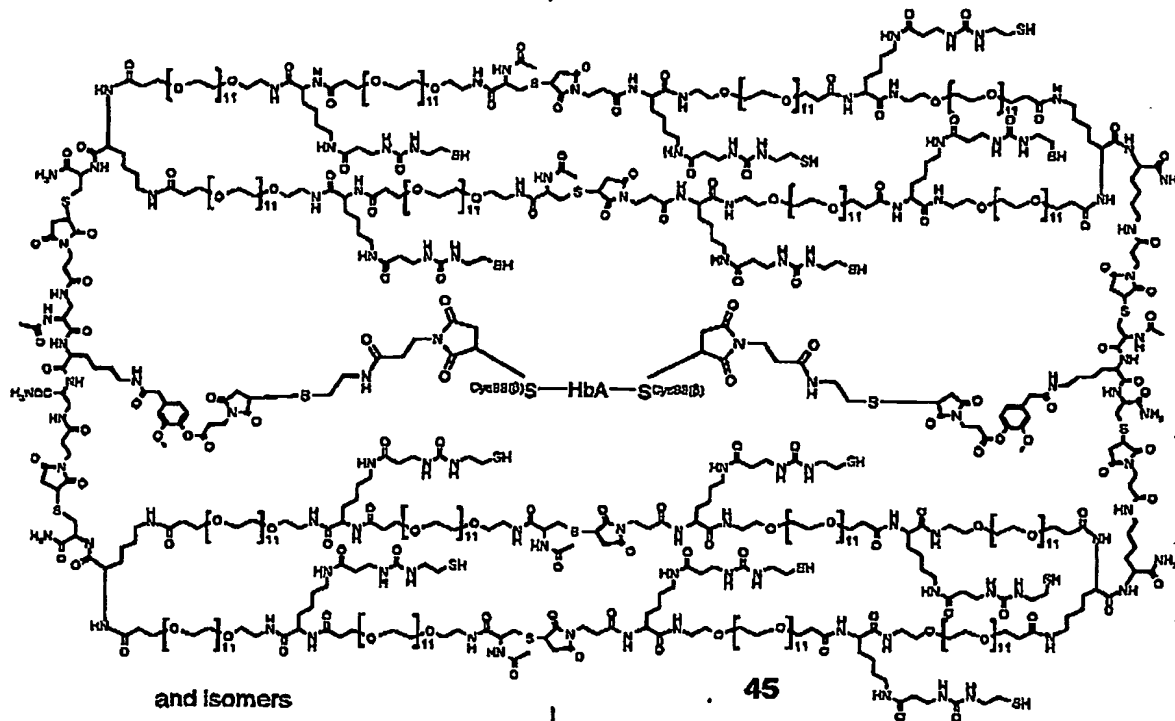


Fig. 2 Size exclusion chromatograms of a) 41, b) 42 and c) Hb released from 42. UV signals were recorded at 280 nm.

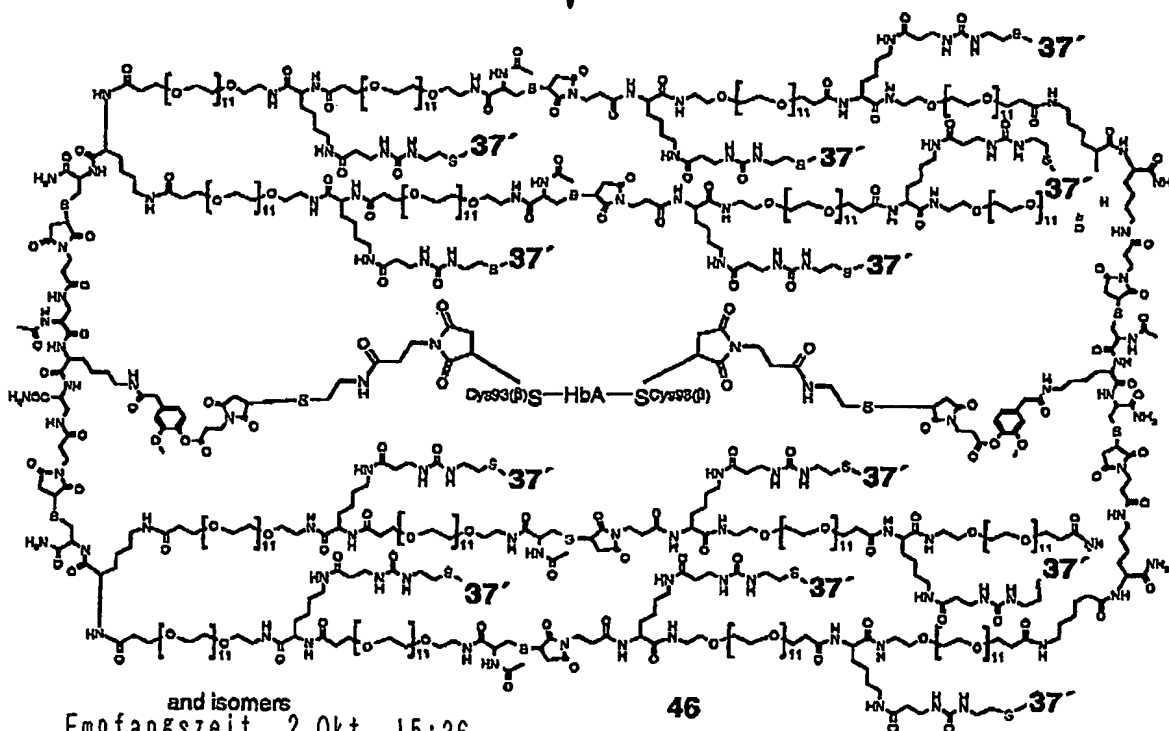
VI - Synthesis of a Hemoglobin MPIC with capping reagents

39 + 31

1. pH 7.5
2. 10 mM TCEP



37



Empfangszeit 2.Okt. 15:36

Structural element **37'** represents the succinimidyl-containing product of the Michael addition of the neighboring proteophore thiol to the maleimido group of **37**.

VI-1) Conjugation of 39 to bis-maleimide-macrocyclic structure 31

The conjugation of **39** to **31** was performed according to protocol IV-2 in analogy to the preparation of **40**. Disulfide-moieties associated with the proteophore were reduced by incubation in 10 mM TCEP (pH 7.5) for 30 min. Product **45** was purified by SEC (Superdex 200 column).

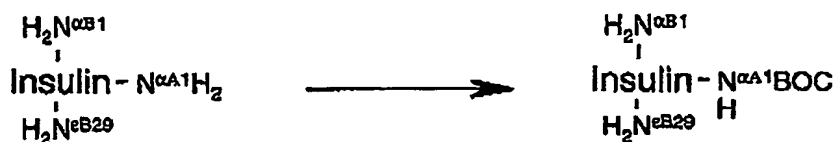
VI-2) Modification of Hemoglobin MPIC with Mal-PEG4x5k (37)

Compound **45** was reacted with 30 eq **37** for 1 h at RT in 50 mM sodium phosphate buffer (pH 7.0) and purified by SEC (Superdex 200, flow rate: 0.75 ml/min).

SEC (retention time) **46**: 11.2 min

VII - Synthesis of an insulin MPIC

VII-1) Synthesis of N^{αA1}-Boc-insulin 47



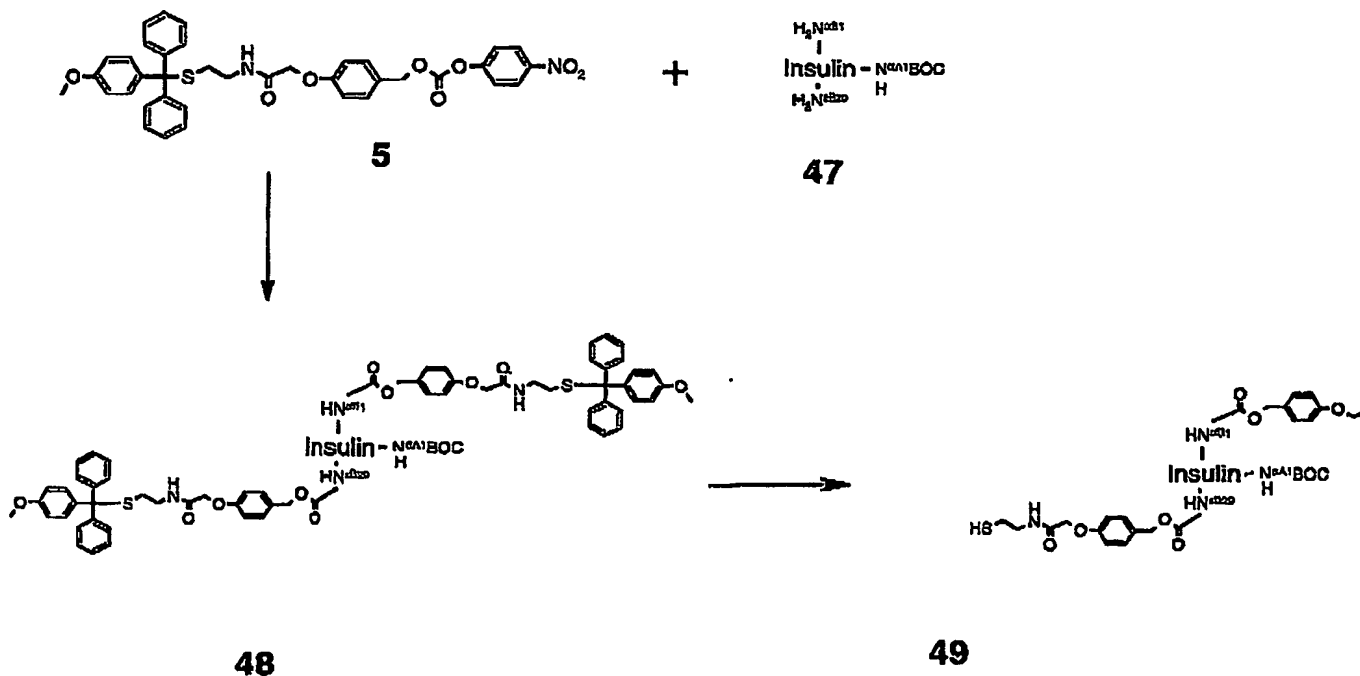
rh Insulin

47

Insulin was dissolved in DMSO and reacted with 1.1 eq (t-BOC)₂O for 60 min to yield N^{αA1}-Boc-insulin 47. Purification was achieved by RP-HPLC. Enzymatic digestion of 47 with endo-GluC and subsequent characterisation of the resulting fragments by LCMS confirmed regioselective modification of the amino terminus of the A chain of insulin.

MS (MW calculated) 47: 5907 g/mol (5907 g/mol)

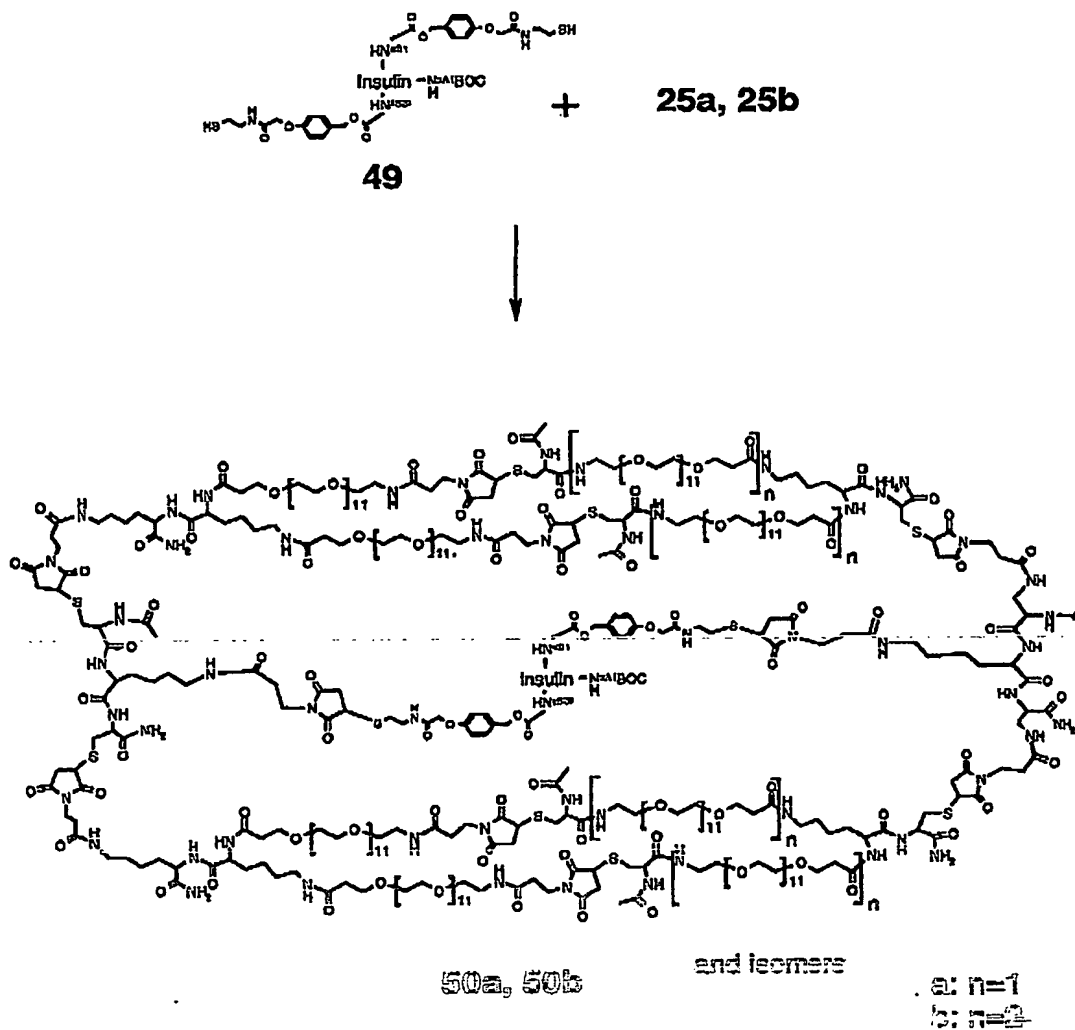
VII-2) Synthesis of N^{αA1}-Boc, N^{αB1}, N^{αB29}-bis-(thiolinker)-insulin 49



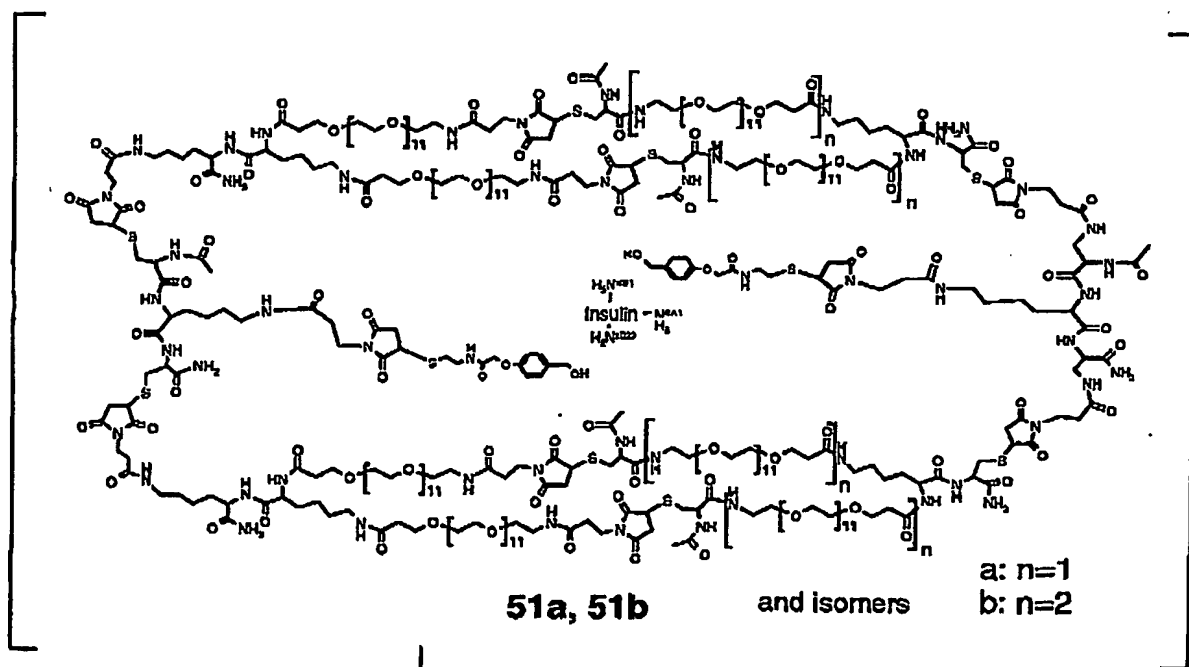
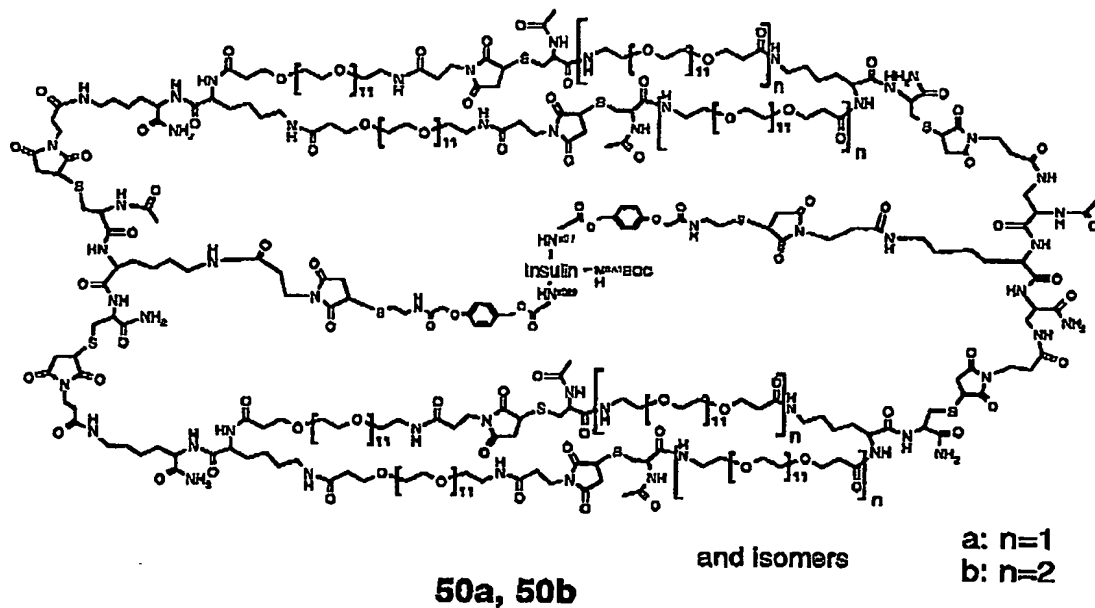
47 was dissolved in DMSO and incubated with 10 eq of activated linker 5 for 5 h at pH 8-9. The pH was adjusted by addition of DIEA, if necessary. Subsequent RP-HPLC purification afforded $N^{\alpha A1}$ -Boc, $N^{\alpha B1}$, $N^{\epsilon B29}$ -bis-(Mmt-thiolinker)-insulin 48. After lyophilization, the Mmt-protecting group was removed by incubation for 30 min with 1/99 (v/v) TFA/DCM and product $N^{\alpha A1}$ -Boc, $N^{\alpha B1}$, $N^{\epsilon B29}$ -bis-(thiolinker)-insulin 49 was purified by RP-HPLC.

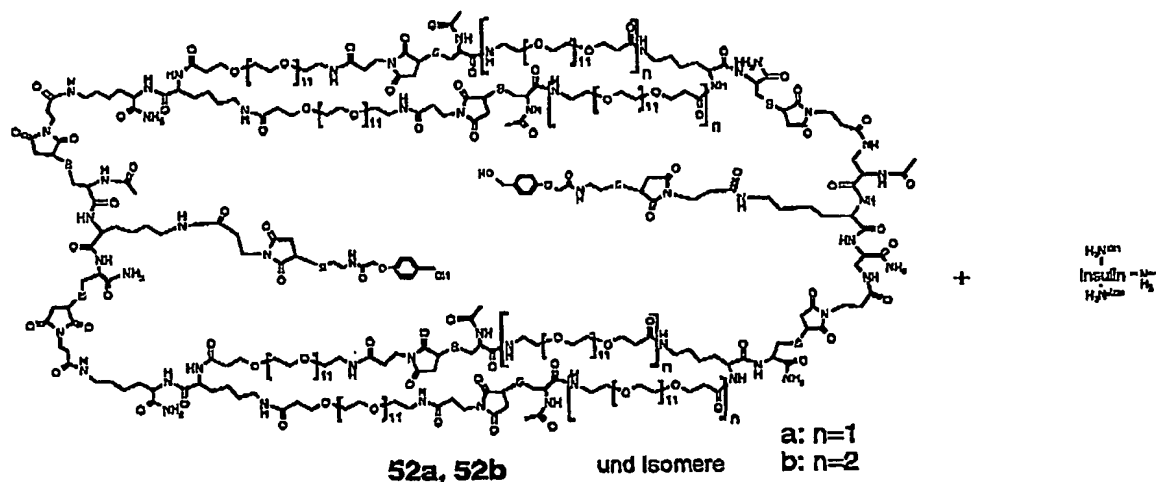
MS (MW calculated) 49: 6450 g/mol (6442 g/mol)

VII-3) Synthesis of macrocyclic structure-insulin conjugates 50a and 50b



VII-4) Release of insulin from insulin MPIC 50a/50b

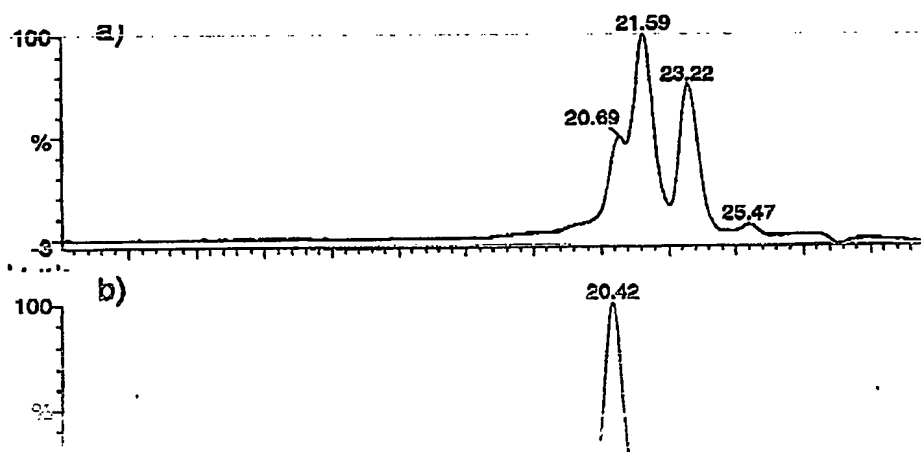




Insulin was released from conjugates 50a or 50b, respectively, by cleavage of the linker moieties by incubation with 1:1 (v/v) DCM/TFA for 15 min. These conditions also effected removal of the Boc-protection group on the α -Aminogruppe of the insulin A-chain.

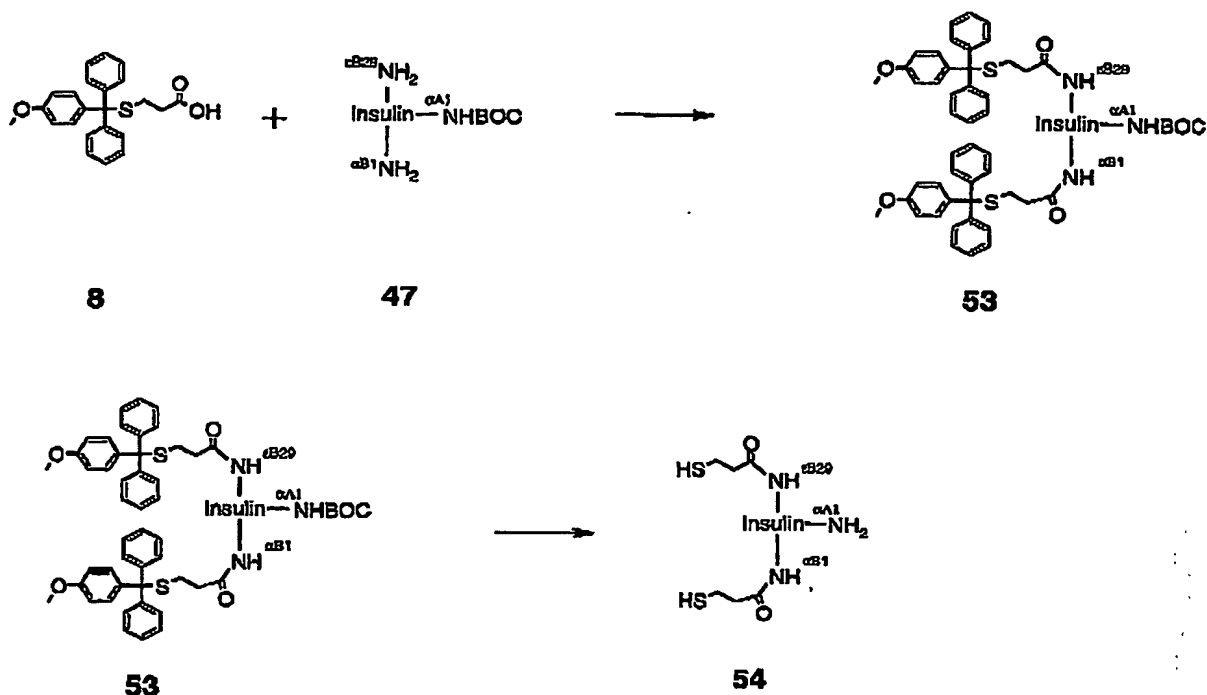
Educts 50a and 50b, linker cleavage mixtures containing released insulin and remaining macrocyclic structures 52a and 52b were characterized by SEC (Superdex 200, flow rate: 0.75 ml/min).

Figure 3 displays size exclusion chromatograms of 50a and products of the linker cleavage procedure. Three peaks were detected and identified by LCMS analysis. The peak at retention time 23.2 min contains insulin, the peak at 21.6 min is EOC 52a. The peak at retention time 20.7 min corresponds to EOC-insulin-monoconjugate.



VIII - Synthesis of insulin MPIC 57, 58 and 59

VIII-1) Synthesis of N^{αB1},N^{αB29}-bis-(mercaptopropionyl)-insulin 54



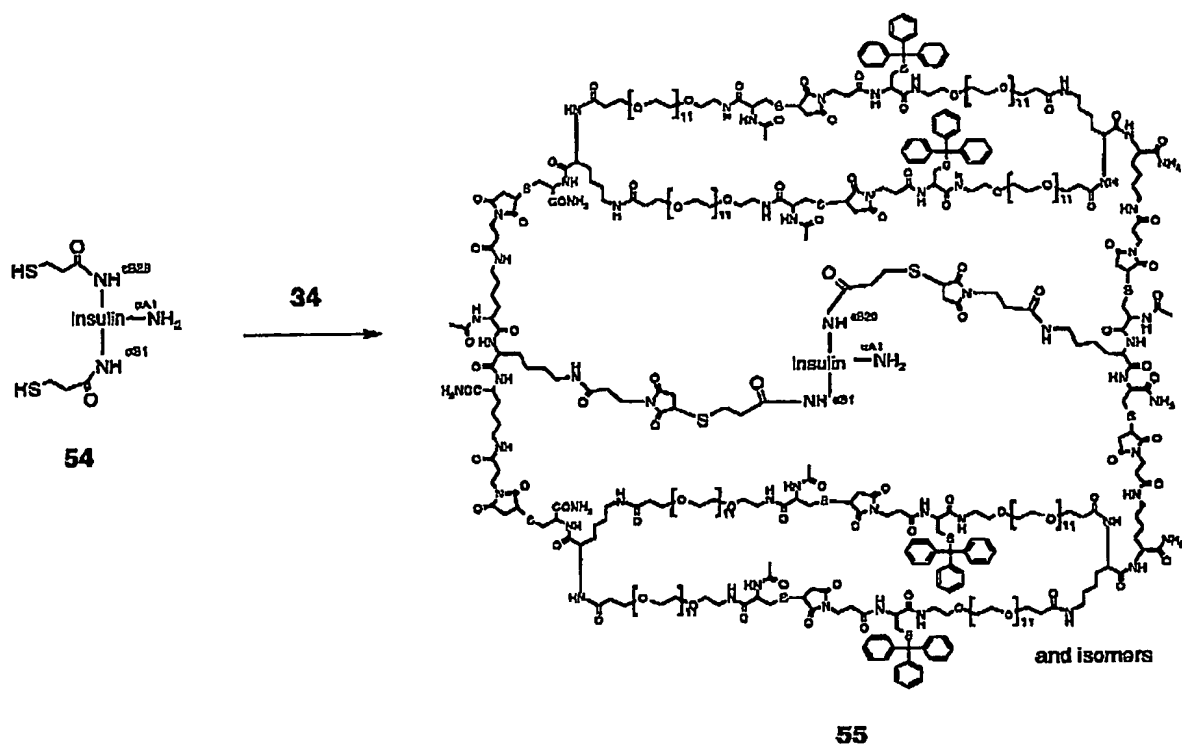
240 μ l of a solution containing 0.64 M DIC, 0.56 M 8 and 0.29 M N-hydroxysuccinimide in DMF were preincubated for 30 min. 15 mg 47 were dissolved in 1 ml 1/1 (v/v) DMSO/water and mixed with 40 μ l DIEA. The mixtures were combined and incubated for 1 h. Compound 53 was purified by RP-HPLC.

MS (MW calculated) 53: 6630 g/mol (6627 g/mol)

After lyophilization, Mmt- and Boc-protection groups were removed by incubation with 2/48/50 (v/v/v) TES/TFA/DCM for 30 min. Product 54 was purified by RP-HPLC.

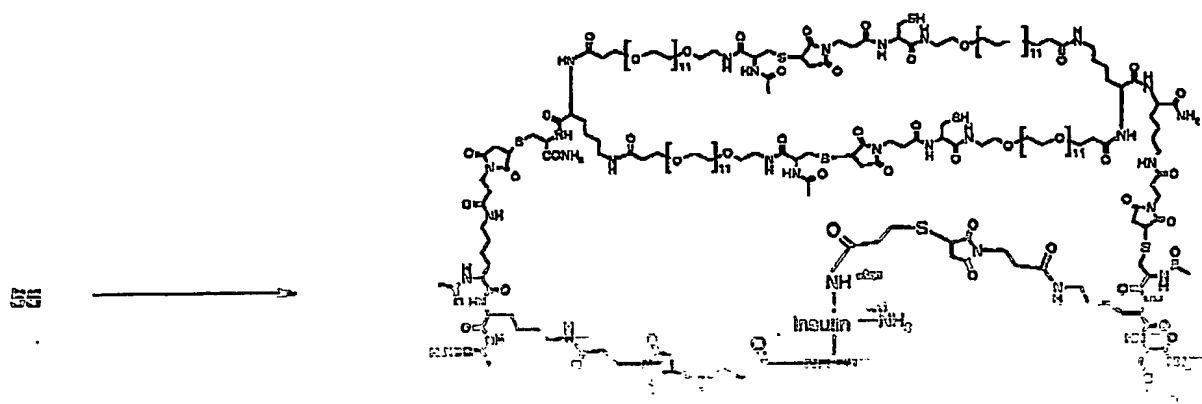
MS (MW calculated) 54: 5981 g/mol (5983 g/mol)

VIII-2) Synthesis of Insulin MPIC 56



150 μ l of a 3 mM solution of compound 54 in 1/1 (v/v) water/acetonitrile were mixed with 270 μ l of a 1.5 mM solution of macrocyclic structure 34 in 1/1 (v/v) water/acetonitrile and diluted with 20/80 (v/v) acetonitrile/water to a total volume of 22 ml. Subsequently, pH was adjusted to 7.5 by addition of 0.5 M phosphate buffer pH 8.0.

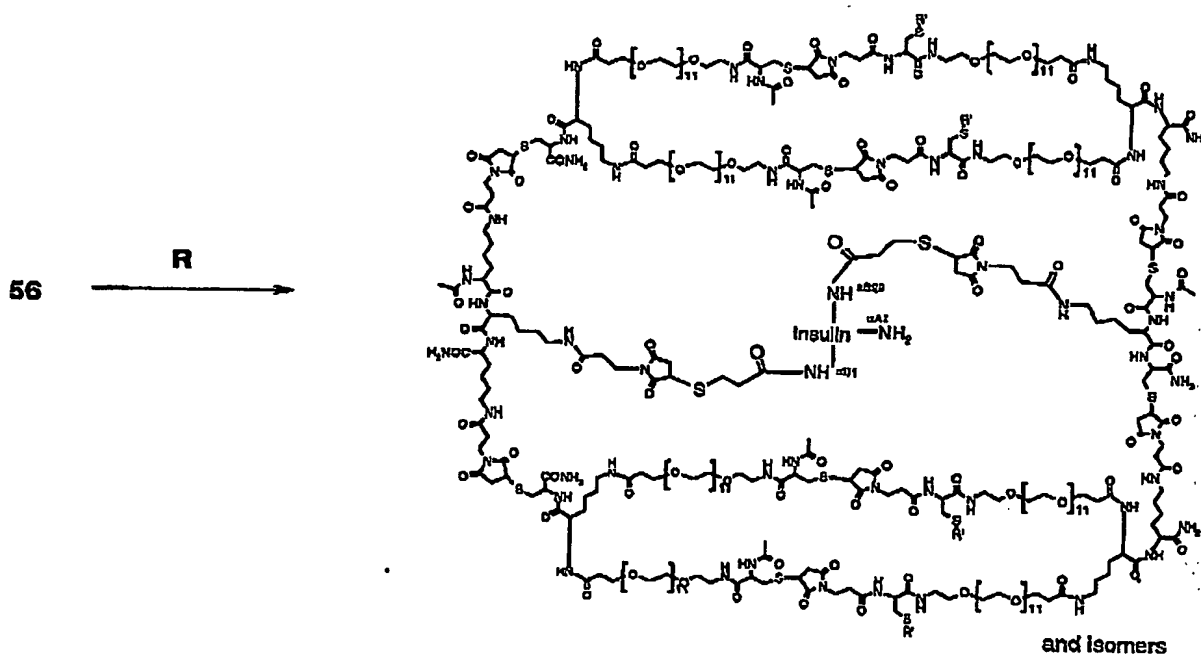
After 15 min incubation, compound 55 was purified by RP-HPLC and lyophilized.



Removal of Trt-protecting groups from compound **55** was effected by incubation for 30 min in 2/58/40 (v/v/v) TES/TFA/DCM. A two-step purification procedure employing RP-HPLC and SEC (Superdex 75, flow rate: 0.75 ml/min) yielded product **56**.

MS (MW calculated) **56**: 15167 g/mol (15166 g/mol)

VIII-3) Synthesis of modified Insulin MPIC **57**, **58** and **59**



57: R = N-Ethyl-Maleimide

58: R = **38**

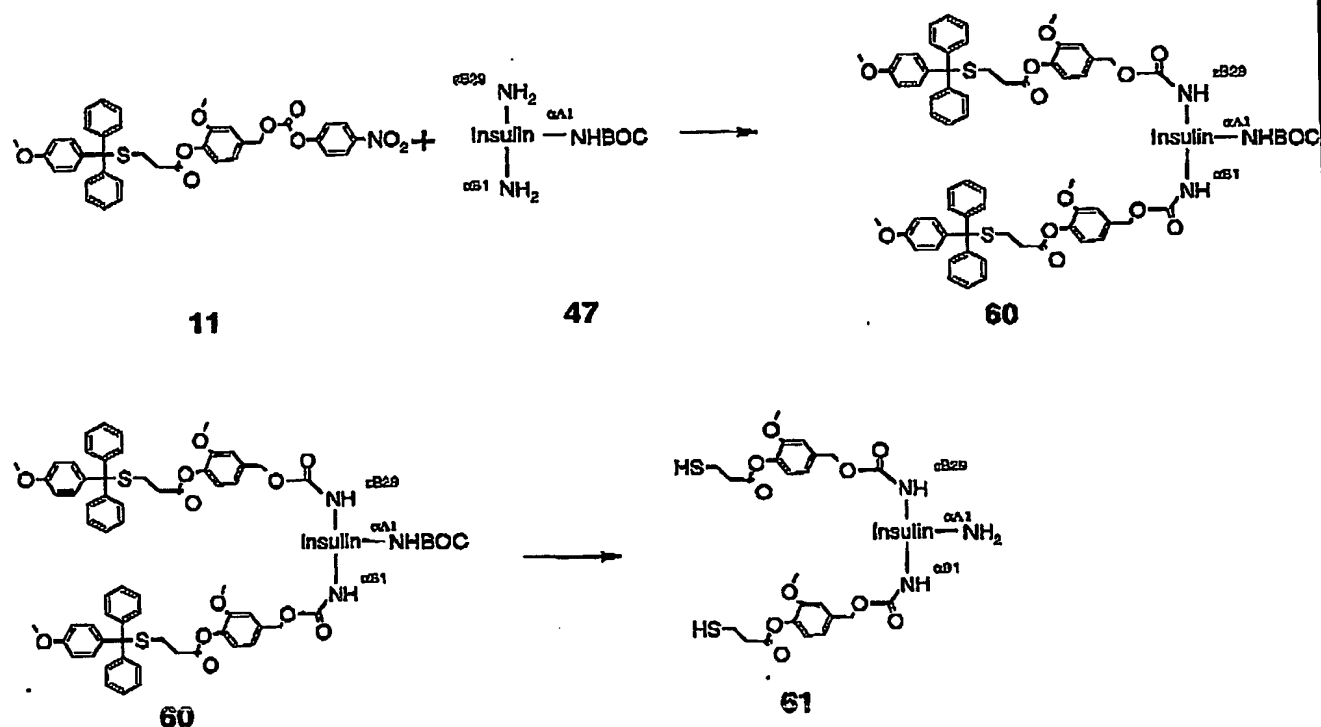
59: R = **37**

Structural element **R** represents the succinimidyl-containing product of the Michael addition of the neighboring proteophore thiol to the maleimido group of **37**, **38** or N-ethyl maleimide, respectively. Aliquots (60 nmol) of compound **56** were reacted with 10 eq N-ethyl-maleimide, or **37** or **38**, respectively, for 15 min in 500 μ l 1/4 (v/v) acetonitrile/100 mM phosphate buffer (pH 8.0). Subsequent purification by SEC (Superdex 200, flow rate: 0.75 ml/min) yielded products **57**, **58** or **59**, respectively.

Compound	Retention time
57	20.8 min
58	16.8 min
59	13.5 min

IX - Synthesis of Insulin MPIC 64 and 65

IX-1) Synthesis of N^{αB1}, N^{αB29}-bis-(thiolinker)-insulin 61

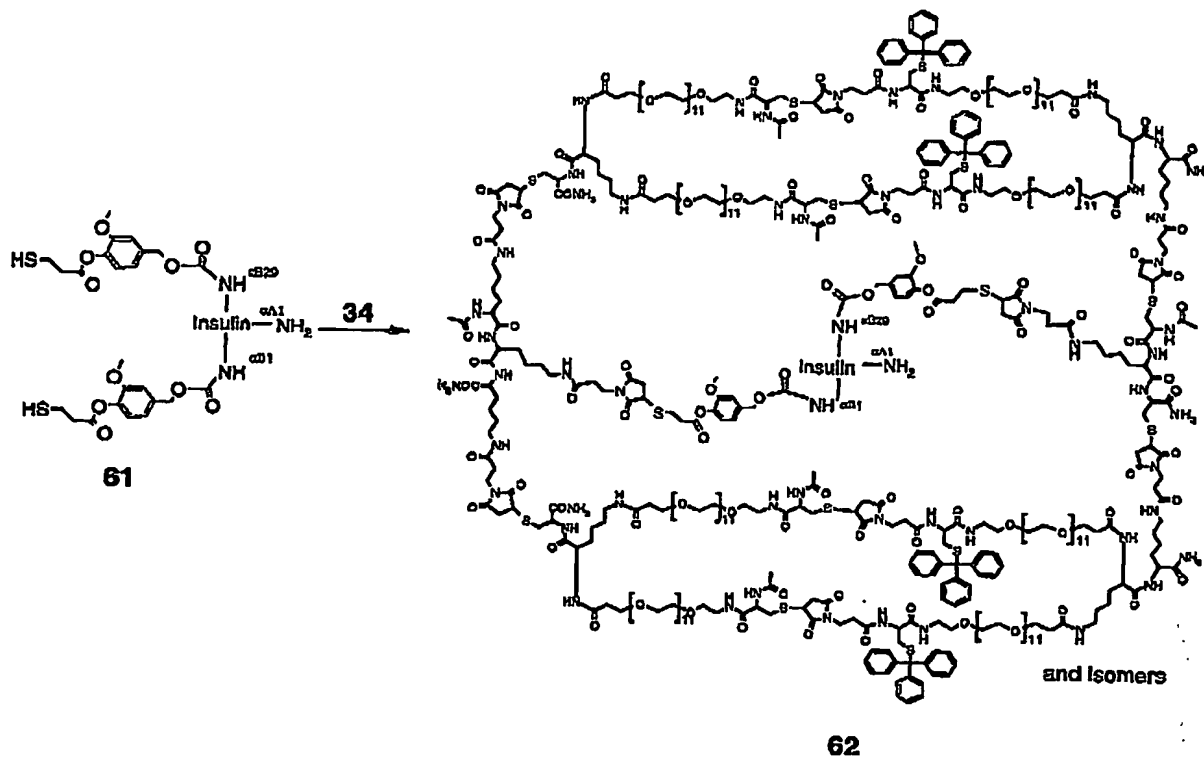


47 was mixed with 4 eq 11 in DMSO. The solution was adjusted to pH 8.0 by addition of DIEA and stirred for 2.5 h at RT under nitrogen atmosphere, RP-HPLC-purification and lyophilization gave compound 60.

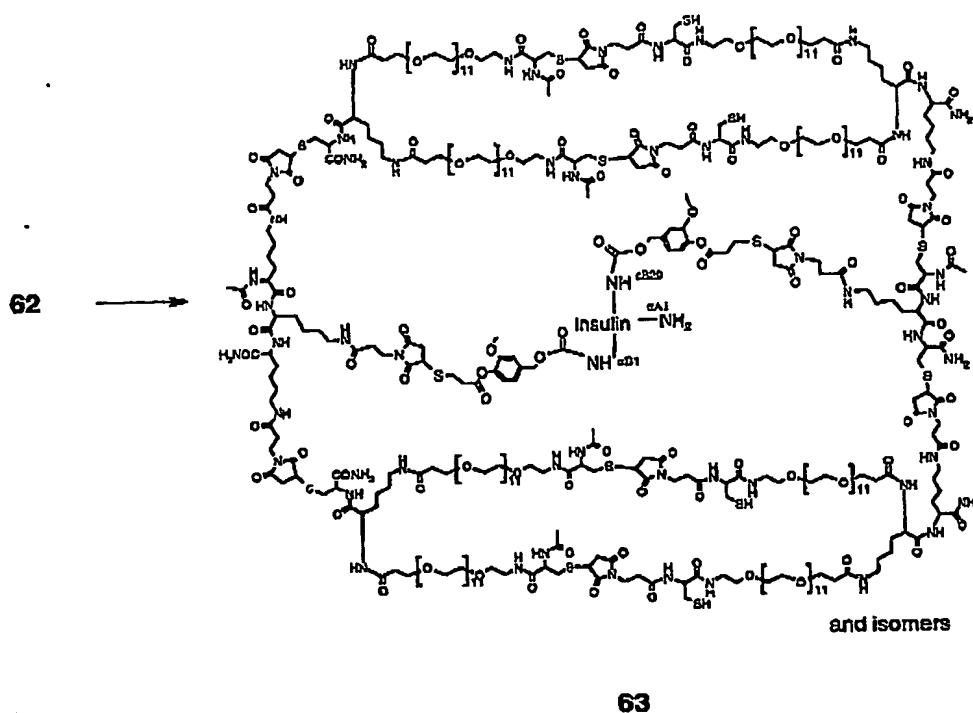
MS (MW calculated) 60: 6991 g/mol (6991 g/mol)

Removal of Trt-protection of 60 was afforded by stirring the compound in TFA/TES 95/5 (v/v) for 10 min at RT. The solution was dried under nitrogen flow. The residue was incubated for 30 min at 55°C in 1/1 TFA/EDC (v/v) and solvent was removed by a nitrogen stream. The residue was purified by RP-HPLC.

IX-2) Synthesis of Insulin MPIC 62



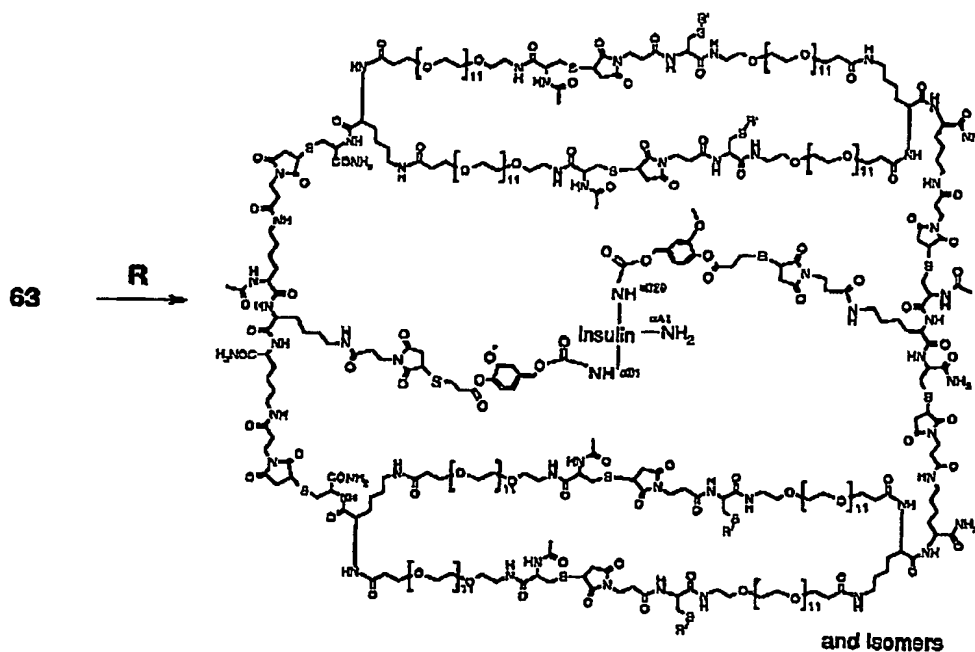
1.8 ml of a solution of compound 61 (200 μ M) in 1/1 (v/v) water/acetonitrile were mixed with 180 μ l of a 1.5 mM solution of macrocyclic structure 34 in 1/1 (v/v) water/acetonitrile and diluted with 80/20 (v/v) water/acetonitrile to a total volume of 24 ml. Subsequently, the pH was adjusted to 7.5 by addition of 0.5 M phosphate buffer (pH 8.0). After incubating for 15 min, 62 was purified by RP-HPLC and lyophilized.



Detritylation of compound 62 was afforded by incubation for 30 min with 2/58/40 (v/v/v) TES/TFA/DCM. A two-step purification procedure employing RP-HPLC and SEC (Superdex 200, flow rate: 0.75 ml/min) yielded product 63.

MS (MW calculated) 63: 15518 g/mol (15526 g/mol)

IX-3) Synthesis of modified Insulin MPIC 64 and 65



64: R = N-Ethyl-Maleinimide

65: R = 37

Structural element **R** represents the succinimidyl-containing product of the Michael addition of the neighboring proteophore thiol to the maleimido group of compound 37 or N-ethyl-maleimide, respectively.

Aliquots (9 nmol) of compound 63 were incubated with 15 eq of N-ethyl-maleimide or 37, respectively, for 3 min in 70 μ l 1/4 (v/v) acetonitrile/phosphate buffer 100mM (pH 7.5). Subsequent purification by SEC (Superdex 200, flow rate: 0.75 ml/min) gave products 64 or 65, respectively.

Compound	Retention time
64	20.2 min
65	13.3 min

IX-4) Release of insulin from Insulin MPIC 64 and 65

Release of insulin from Insulin MPIC 64 or 65, respectively, was effected by linker hydrolysis in aqueous buffer. A ca. 10 μ M solution of conjugate was incubated in 10 mM HEPES buffer (pH 7.4), 150 mM NaCl, 3 mM EDTA and 0.005 % Tween. Samples were taken at time intervals and analyzed by HPLC and UV detection. The peak correlating with the retention time of native insulin was integrated and plotted against reaction time, and curve-fitting software was applied to estimate the corresponding halftime of release. Under these

conditions, insulin was released from 64 with a halftime of approximately 150 min and from compound 65 with a halftime of ca 230 min.

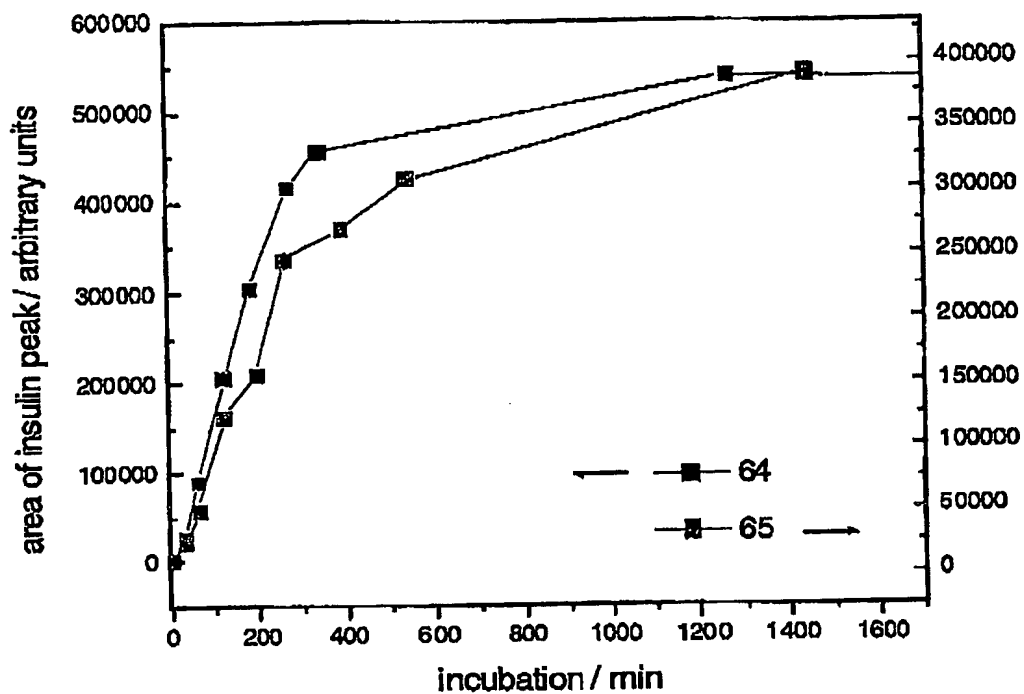
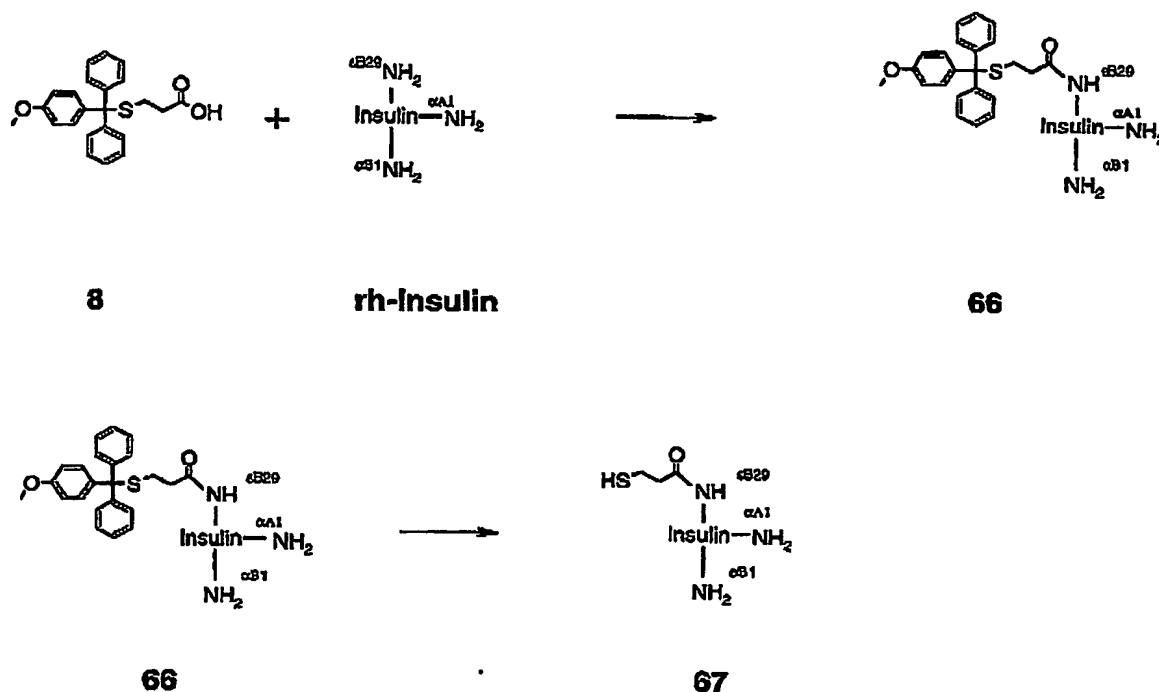


Fig. 4 : In vitro release of insulin from Insulin MPIC 64 or 65, respectively. Kinetics were determined by HPLC using UV detection at 215 nm.

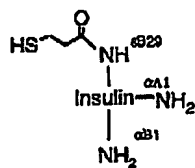
X – Synthesis of reference insulin conjugates 68-75

X-1) Synthesis of N^{B29}-mercaptopropionyl-insulin 67

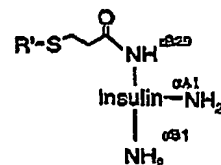


150 μ l of a solution of 0.64 M DIC, 0.56 M **8** and 0.29 M N-hydroxy succinimide in DMF were prepared and incubated for 30 min. 18 mg insulin were dissolved in 1.6 ml 20/30/50 (v/v/v) DMSO/DMF/water and 50 μ l DIEA were added, followed by addition of 50 μ l of the preincubated cocktail containing compound **8**. After incubation for 30 min another 100 μ l of preactivated **8** were added. After incubation for 30 min product **66** was purified by RP-HPLC. Acetonitrile was removed in vacuo and cleavage of the Mmt-protecting group was afforded by addition of 98/2 (v/v) TFA/TES until an intensive yellow colour was observed. Product **67** was obtained by RP-HPLC purification.

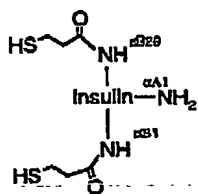
MS (MW calculated) **67**: 5893 g/mol (5896 g/mol)



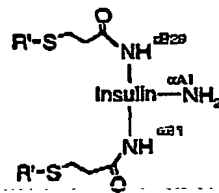
R



- 68: R = N-ethyl-maleimide**
69: R = Maleimide-PEG5k
70: R = 37
71: R = Maleimide-PEG20k
72: R = Maleimide-PEG2x20k



R



- 73: R = N-ethyl-maleimide**
74: R = Maleimide-PEG5k
75: R = Maleimide-PEG20k

~~Structural element R represents the succinimidyl-containing product of the Michael addition
of a thiol to the maleimide group of compound E7EN, ethyl-maleimide-1-maleimide-E6G3b,
maleimide-E6G3c or maleimide-E6H3d.~~

71 μ l of a solution of **67** (464 μ M) in 1/4 (v/v) acetonitrile/water were mixed with 7 μ l of a solution of 10 mM Maleimide-PEG5k (1 eq) in 1/4 (v/v) acetonitrile/water and 10 μ l of 0.5 M phosphate buffer (pH 8.0) and incubated for 15 min. Compound **69** was purified by SEC (Superdex 200).

Compounds **70** and **71** were obtained according to the procedure used for the conjugation of **69** to **37** or Maleimide-PEG20k, respectively.

43 μ l of a solution of **67** (464 μ M) in 1/4 (v/v) acetonitrile/water were mixed with 22 μ l of a solution of 2 mM Maleimide-PEG2x20k (2 eq) in 1/4 (v/v) acetonitrile/water and 10 μ l of 0.5 M phosphate buffer (pH 8.0) and incubated for 15 min. Compound **72** was purified by SEC (Superdex 200).

11.4 μ l of a solution of **54** (2.9 mM) in 1/4 (v/v) acetonitrile/water were mixed with 50 μ l 1/1 (v/v) acetonitrile/water, 7 μ l of a 100 mM solution of N-ethyl maleimide (10eq) and 10 μ l of 0.5 M phosphate buffer (pH 8.0) and incubated for 3 min. Compound **73** was purified by SEC (Superdex 200).

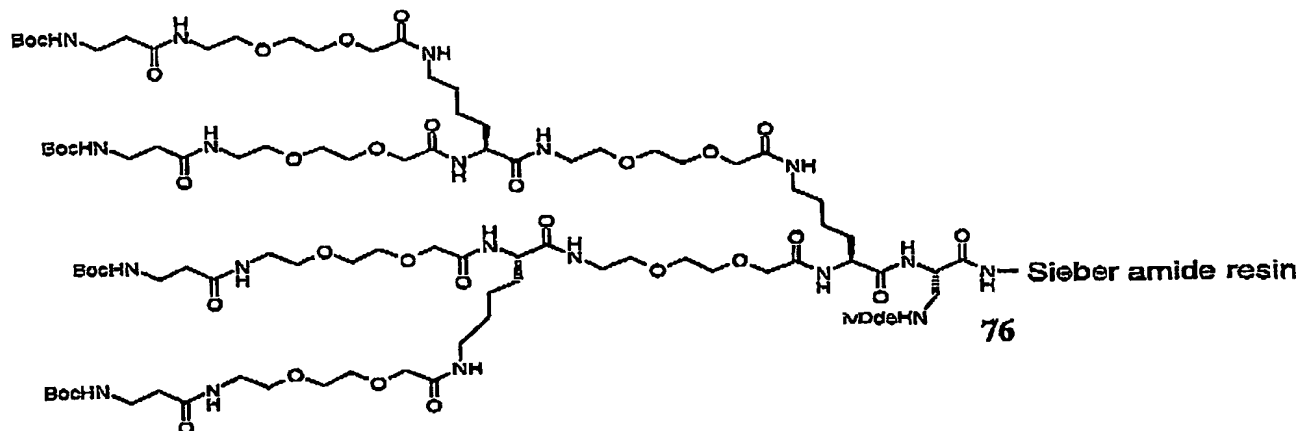
11.4 μ l of a solution of **54** (2.9 mM) in 1/4 (v/v) acetonitrile/water were mixed with 50 μ l 1/1 (v/v) acetonitrile/water, 15 μ l of a 10 mM solution of Maleimide-PEG5k (2eq) in 1/4 (v/v) acetonitrile/water and 10 μ l of 0.5 M phosphate buffer (pH 8.0) and incubated for 15 min. Compound **74** was purified by SEC (Superdex 200, flow rate: 0.75 ml/min). Compound **75** was obtained accordingly through reaction of **74** with Maleimide-PEG20k.

Preparative SEC of compounds **68-75** yielded fractions of 1.4-2.2 ml. Concentrations were determined by measuring UV extinction at 275 nm, assuming an average coefficient of extinction of $\epsilon_{275}=7500$.

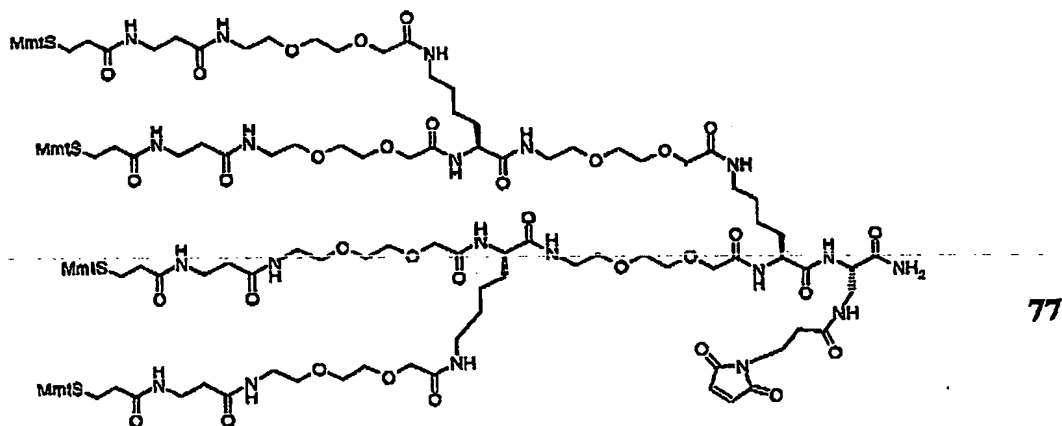
Compound	E_{275} / mOD	Concentration / μ Mol	Retention time /min
68	97	12.9	23.0
69	102	13.6	18.8
70	33	4.4	16.1
71	60	8.0	15.9
72	38	5.0	14.3
73	100	13.3	23.3
74	75	10.0	17.8
75	59	7.9	14.1

XI) Synthesis of Insulin DPICs

XI-1) Synthesis of dendron building block 77

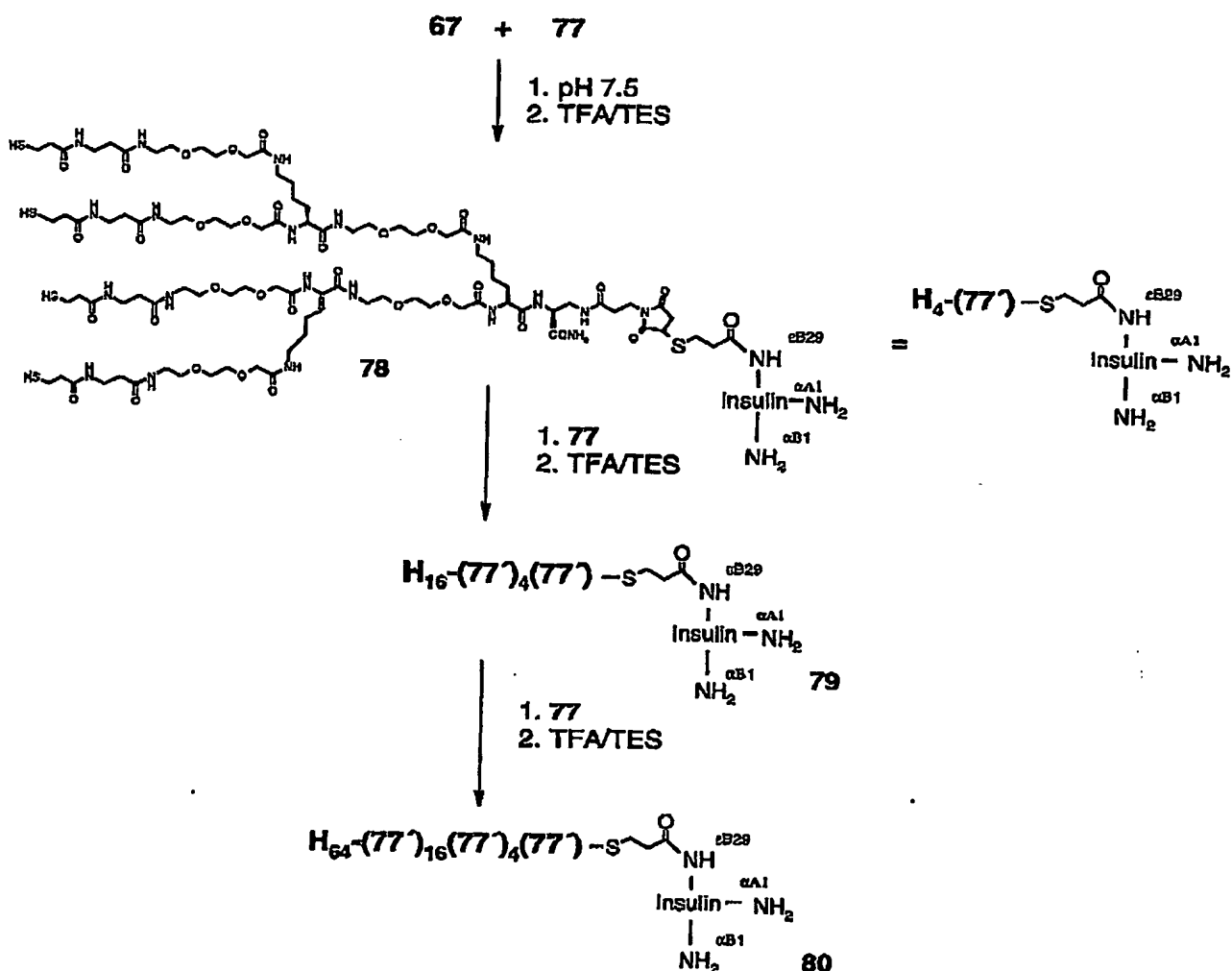


1. DMF/Hydrazin
2. Maleimidopropionic acid/DIC
3. TFA/TES/H₂O
4. 8 / DIC



76 was obtained according to the standard solid-phase synthesis protocol. The amino acid sequence Fmoc-Dpr(ivDde)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-Ado-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-Ado-OH and Boc- β Ala-OH were coupled to Sieber amide resin.

XI-2) Synthesis of Insulin dendrimers 78, 79 and 80



Structural element 77' represents the succinimidyl-containing product of the Michael addition of a thiol group to the maleimido group of compound 77 after Mmt protecting group removal. Thiol groups are either associated with the insulin-linker conjugate or with a dendron-insulin conjugate.

XI-2-1) Synthesis of 1st generation insulin- $N^{\alpha B29}$ -Mpa-dendrimer (78)

2.9 mg 67 were dissolved in 100 μ l 4/1 acetonitrile/water and mixed with 180 μ l of a solution of 77 (3.4 mM) in 4/1 acetonitrile/water. 20 μ l of 0.5 M phosphate buffer (pH 7.5) were added and the solution was treated for 30 min with in an ultrasonic bath. The cocktail was mixed with 300 μ l TFA and 10 μ l TES agitated for 30 s. After dilution with 1/1 acetonitrile/water, compound 78 was purified by RP-HPLC. Yield: 43 %.

MS (MW calculated) 78: 8035 g/mol (8042 g/mol).

XI-2-2) Synthesis of 2nd generation insulin-N^{εB29}-Mpa-dendrimer (79)

350 µl of a solution of 78 (420 µM) in 4/1 acetonitrile/water were mixed with 150 µl of a 5.2 mM solution of dendron 77 in 4/1 acetonitrile/water. 20 µl of 0.5 M phosphate buffer (pH 7.5) were added and the solution was treated for 30 min in an ultrasonic bath. The cocktail was mixed with 500 µl TFA and 20 µl TES and agitated for 30 s. After dilution with 1/1 acetonitrile/water, compound 79 was purified by RP-HPLC.

Yield: 40%

MS (MW calculated) 79: 16621 g/mol (16622 g/mol).

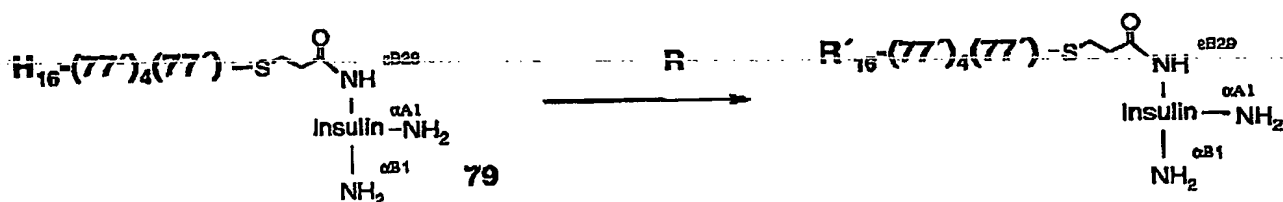
XI-2-3) Synthesis of 3rd generation insulin-N^{εB29}-Mpa-dendrimer (80)

46 µl of a solution of 79 (235 µM) in 4/1 acetonitrile/water were mixed with 60 µl of a 5.2 mM solution of dendron 77 in 4/1 acetonitrile/water. 20 µl of 0.5 M phosphate buffer (pH 7.5) were added and the solution was treated for 30 min in an ultrasonic bath. The cocktail was mixed with 100 µl TFA and 5 µl TES and agitated for 30 s. After dilution with 1/1 water/acetonitrile compound 80 was purified by HPLC.

Yield: 38%

MS (MW calculated) 80: 50967 g/mol (50967 g/mol)

X-3) Synthesis of end-capped dendrimers 81, 82, 83 and 84 based on 2nd generation insulin-N^{εB29}-Mpa-dendrimer



- 81: R = N-Ethylmaleimide
- 82: R = 38
- 83: R = Maleimide-PEG5k
- 84: R = 37

XI-3-1) Synthesis of 81

70 µl of a solution of 79 (230 µM) in 1/1 acetonitrile/water were mixed with 10 µl of a 24 mM solution of N-ethyl-maleimide in acetonitrile. pH was adjusted to 7.5 with 0.5 M phosphate buffer (pH 7.5) and the solution was incubated for 30 min at RT. Purification by SEC (Superdex 200, flow rate: 0.75 ml/min) gave compound 81.

SEC (retention time) 81: 20.28 min

MS (MW calculated) 81: 18629 g/mol (18624 g/mol)

XI-3-2) Synthesis of 82

70 µl of a solution of 79 (230 µM) in 1/1 acetonitrile/water were mixed with 3 mg 38. The pH was adjusted to 7.5 with 0.5 M phosphate buffer (pH 7.5) and the solution was incubated for 30 min at RT. Purification by SEC (Superdex 200, flow rate: 0.75 ml/min) yielded compound 82.

SEC (retention time) 82: 14.5 min

XI-3-3) Synthesis of 83

70 µl of a solution of 79 (230 µM) in 1/1 acetonitrile/water were mixed with 2.6 mg Maleimide-PEG5k. The pH was adjusted to 7.5 with 0.5 M phosphate buffer (pH 7.5) and the solution was incubated for 30 min at RT. Purification by SEC (Superdex 200, flow rate: 0.75 ml/min) gave compound 83.

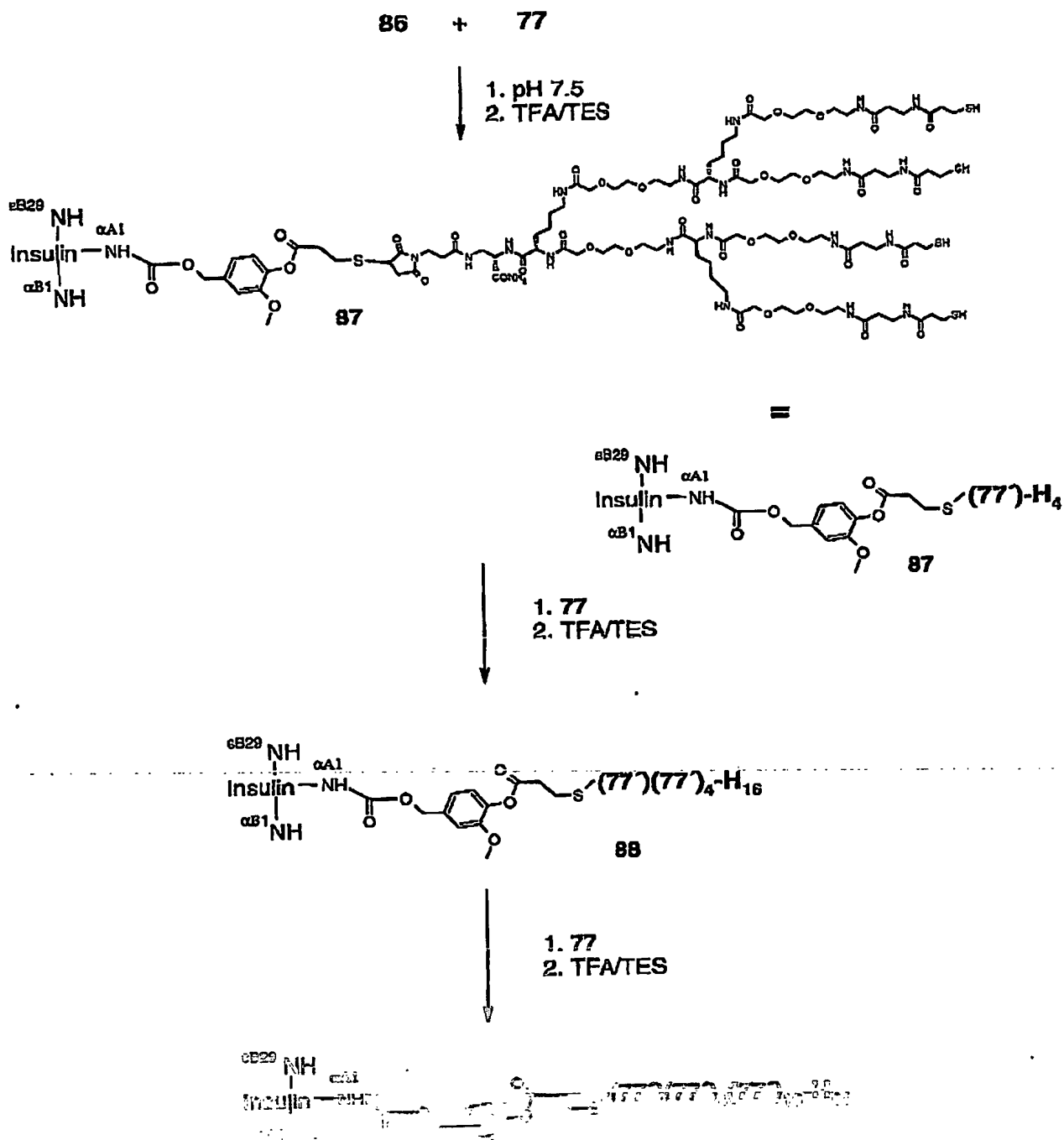
SEC (retention time) 83: 12.9 min

XI-3-4) Synthesis of 84

115 µl of a solution of 79 (65 µM) in 1/1 acetonitrile/water were mixed with 5 mg 37. The pH was adjusted to 7.5 with 0.5 M phosphate buffer (pH 7.5) and the solution was incubated for 30 min at RT. Purification by SEC (Superdex 200, flow rate: 0.75 ml/min) gave compound 84.

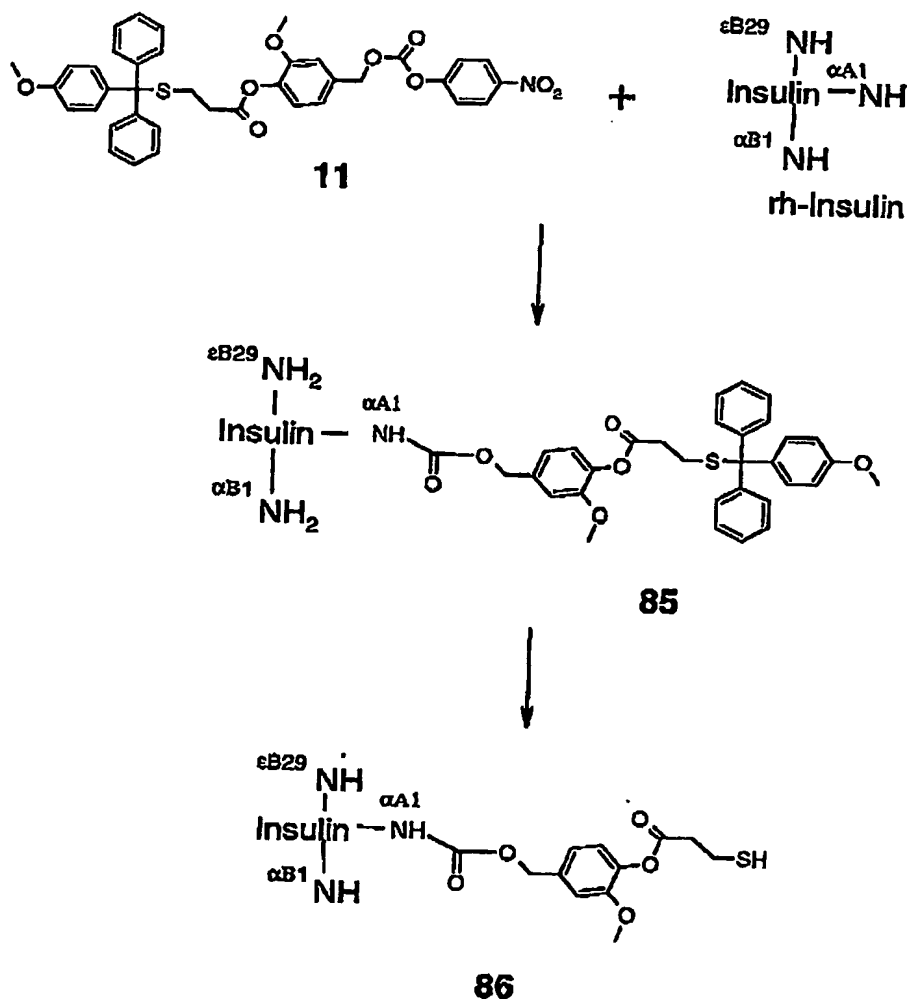
SEC (retention time) 84: 10.9 min

XI-4) Synthesis of cleavable Insulin dendrimeres 87, 88, and 89



Thiol groups are either associated with the insulin-linker conjugate or with a dendron-insulin conjugate.

XI-4-1) Synthesis of N^{αA1}-thiolinker-insulin 86



Insulin in DMSO was mixed with a solution of 0.9 eq **11** in DMSO. The resulting solution was adjusted to pH 8.0 with DIEA and stirred for 1.5 h at RT. RP-HPLC purification gave Mmt-protected intermediate **85**.

MS (MW calculated) **85**: 6350 g/mol (6350 g/mol)

Regioselectivity of the monoconjugation was determined by reduction of **85** with DTT (10 mM) in 0.5 M phosphate buffer pH 8.0 for 1 h at RT and subsequent analysis of the insulin A- and B-chains by LC-MS.

After lyophilization, compound 85 was mixed with 95:5 (v/v) TFA/triethylsilane and stirred for 5 min. Volatiles were removed under nitrogen flow and 86 was purified by RP-HPLC and lyophilized.

MS (MW calculated) 86: 6077 g/mol (6077 g/mol)

XI-4-2) Synthesis of 1st generation N^{αA1}-thiolinker-insulin-dendrimer 87

Synthesis of compound 87 was performed in analogy to the synthesis of compound 78.

Yield: 27 %

MS (MW calculated) 87: 8218 g/mol (8223 g/mol).

XI-4-3) Synthesis of 2nd generation N^{αA1}-thiolinker-insulin-dendrimer 88

Synthesis of compound 88 was performed in analogy to the synthesis of compound 79.

Yield: 17 %

MS (MW calculated) 88: 16809 g/mol (16809 g/mol).

XI-4-4) Synthesis of 3rd generation N^{αA1}-thiolinker-insulin-dendrimer 89

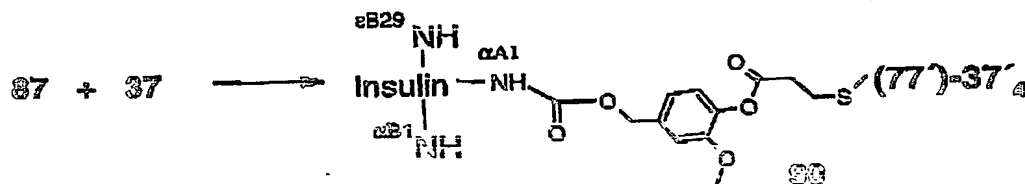
Synthesis of compound 89 was performed in analogy to the synthesis of compound 80.

Yield: 44%

MS (MW calculated) 89: 51117 g/mol (51155 g/mol).

XI-5) Synthesis of end-capped dendrimers 90 and 91 based on 1st or 2nd generation N^{αA1}-thiolinker-insulin-dendrimers

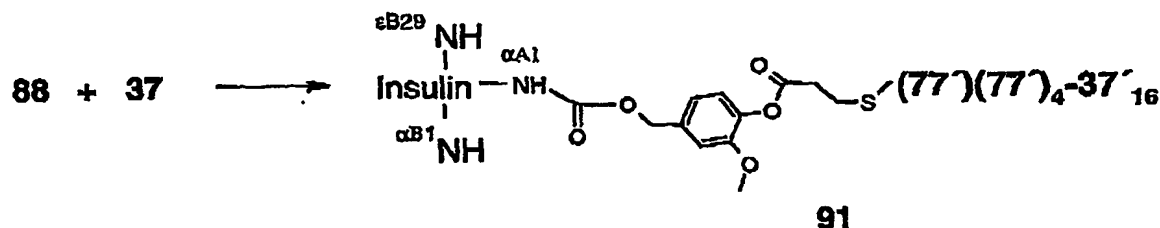
XI-5-1) Synthesis of 90



50 μ l of a solution of **87** (450 μ M) in 1/1 acetonitrile/water were mixed with 3.5 mg of **37**. The pH was adjusted to 7.5 with 0.5 M phosphate buffer (pH 7.5) and the solution was incubated for 30 min at RT. Purification by SEC (Superdex 200, flow rate 0.75 ml/min) gave compound **90**.

SEC (retention time) **90**: 13.0 min

XI-5-2) Synthesis of **91**



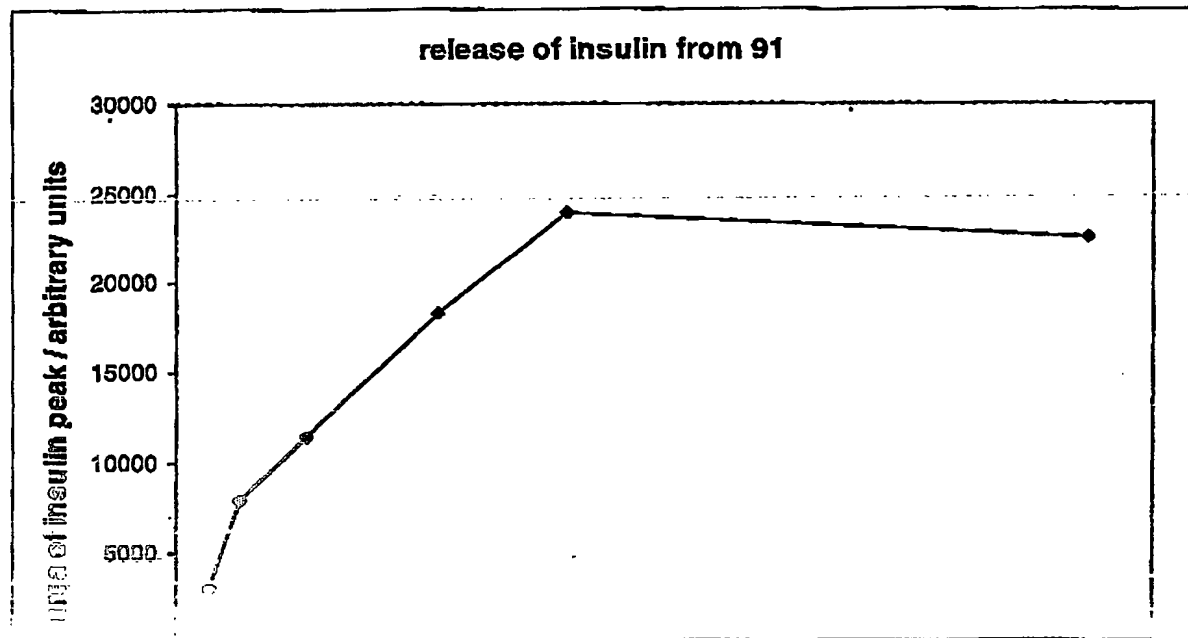
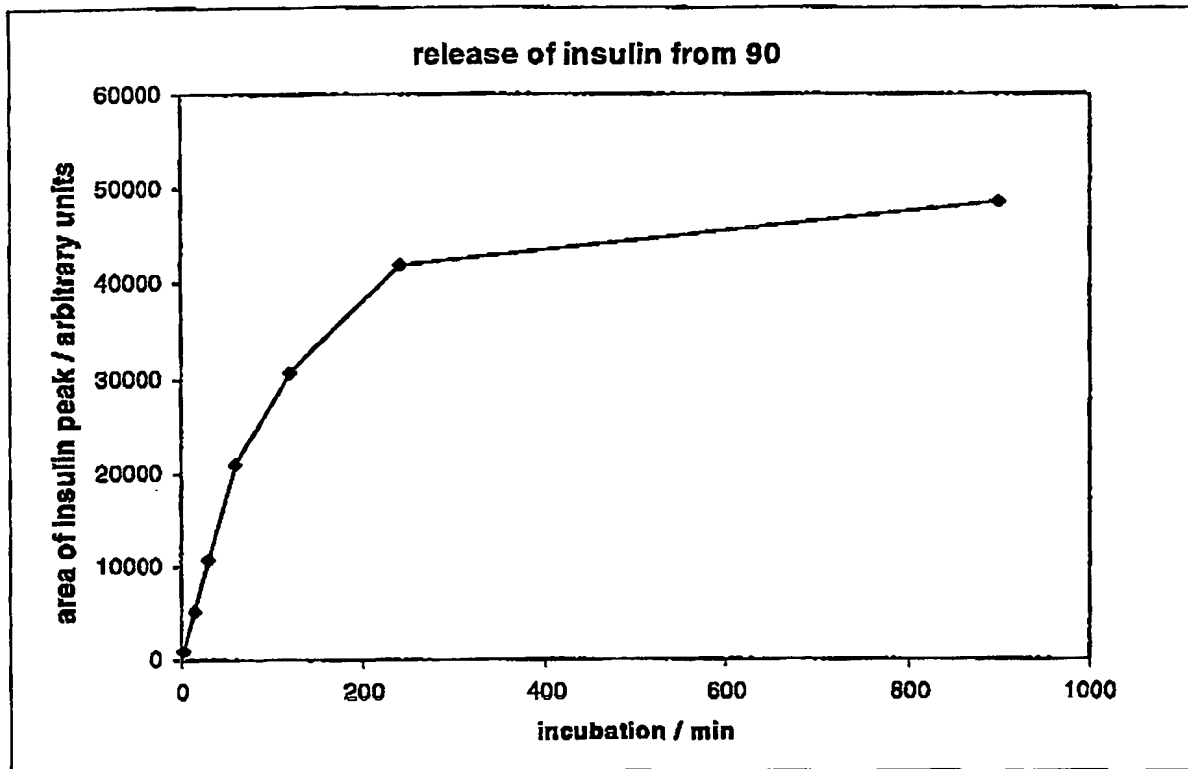
Structural element **37'** represents the succinimidyl-containing product of the Michael addition of a thiol group to the maleimido group of compound **37**. Thiol groups are associated with a generation 2 dendron-insulin conjugate.

75 μ l of a solution of **88** (206 μ M) in 1/1 acetonitrile/water were mixed with 9 mg of **37**. The pH of the solution was adjusted to 7.5 with 0.5 M phosphate buffer (pH 7.5) and the solution was incubated for 30 min at RT. Purification by SEC (Superdex 200, flow rate 0.75 ml/min) yielded compound **91**.

SEC (retention time) **91**: 10.9 min

XI-6) Release of insulin from Insulin DPIC **90** and **91**

SEC-eluates of **90** and **91** were incubated with 10 mM HEPES buffer (pH 7.4), 150 mM NaCl, 3 mM EDTA and 0.005 % Tween at 37 °C, and the amount of free insulin was quantified by HPLC using UV detection at 215 nm (Figure 5).



XII – Analysis of efficiency of encapsulation

Analysis was performed on a BIAcore 2000 surface plasmon resonance instrument using CM5 sensor chips (Biacore).

XII-1) Preparation of sensor chips

A sensor chip was mounted to the instrument and RAMFc (rabbit anti mouse Fc antibody, BIAcore) was immobilized according to Karlsson et al. (J. Immun. Meth, 200, 1997, 121-133 using EDC/NHS activation. Capping of activated, unreacted surface carboxy groups was effected by ethanolamine.

XII-2) Interaction analysis between proteophor-encapsulated insulin and anti-insulin antibodies

Insulin and insulin conjugates 68-75, 57-59 and 81-84 were subject to analysis.

For sample application, dissociation and regeneration, standard flow buffer was used containing 10 mM HEPES (pH 7.4), 150 mM NaCl, 3 mM EDTA, 0,005 % Tween 20.

Three murine anti insulin monoclonal antibodies (Advanced ImmunoChemical Inc., clones C7C9, 8E2, or 7F8, respectively,) were loaded onto prespecified sensor chip areas by injection of 15 µl of a solution of 30µg/ml. A fourth sensor area was used for reference purposes. After equilibration for 2.5 min, 150 µl of a 100 nM solution of insulin or insulin conjugate were injected and flowed across all four sensor areas. A dissociation phase of 3 min was followed by removal of the anti-insulins by injection of 60 µl glycine buffer (pH 2.0) and regeneration of the sensor chip surface. Encapsulation efficiency was measured by recording the refractive index units (RU) of each of the four sensor areas before dissociation.

Highly efficient encapsulation of insulin was achieved by conjugating the protein to proteophore structures **54**, **58** or **59**, respectively, as evidenced by suppression of antibody binding.

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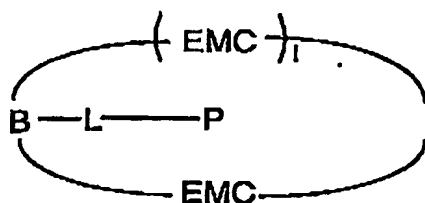
10. The tenth part of the document is a list of names and addresses, which appears to be a directory or a list of contacts. The names are written in a cursive script, and the addresses are listed below them.

DMF	N,N-dimethylformamide
DMSO	dimethylsulfoxide
Dpr	diaminopropionic acid
DTT	dithiothreitol
EDC-HCl	1-Ethyl-3-(3'-dimethylaminopropyl) carbodiimide hydrochloride
Endo-GluC	endoproteinase-GluC
eq	stoichiometric equivalent
Fmoc	9-fluorenylmethoxycarbonyl
Fmoc-PP-OH	Fmoc-aminoethyl-undecaethyleneoxide-propionic acid
Fmoc-Ado-OH	Fmoc-8-amino-3,6-dioxaoctanoic acid
Hb	hemoglobin (human)
HEPES	N-(2-hydroxyethyl) piperazine-N'-(2-ethanesulfonic acid)
HOBT	N-hydroxybenzotriazole
LCMS	mass spectrometry-coupled liquid chromatography
Mal	maleimidopropionyl
Mmt	4-methoxytrityl
Mpa	mercaptopropionyl
MS	mass spectrum
MSNT	1-(mesitylen-2-sulfonyl)-3-nitro-1H-1,2,4-triazole
Mtt	4-methyltrityl
MW	molecular mass
NHS	N-hydroxy succinimide
RP-HPLC	reversed-phase high pressure liquid chromatography
RT	room temperature
RU	response units
SEC	size exclusion chromatography
SPDP	succinimidyl 3-(2-pyridyldithio) propionate
S-tBu	t-butylthio
Suc	succinimidopropionyl
TBTU	O-(benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium tetrafluoroborate
TCEP	tricarboxyethylphosphine
TES	triethylsilane
TFA	trifluoroacetic acid
THF	tetrahydrofurane
Trt	trityl, triphenylmethyl
UV	ultraviolet

1. A protein encapsulated by an encapsulating organic compound EOC wherein the protein and the encapsulating organic compound are present in 1:1 stoichiometry.
2. An encapsulated protein according to claim 1, wherein the EOC is water soluble and contains at least 2 molecule chains EMC of an appropriate length which chains can arrange such that a cavity is formed which can accommodate the protein.
3. An encapsulated protein according to claim 1 or 2, wherein the EMCs within the chain contain linear, branched or cyclical alkyl chains.
4. An encapsulated protein according to any of claims 1 to 3, wherein further groups are present in the EMCs selected from S, N, O, (-S-S)-, oxyethylen, oxypropylene and oxybutylene, amide -C(O)NH- or C(O)NR-, -S-succinimido, amino (-NR-), carboxylic ester (-C(O)O-), sulfonamide (-S(O)₂-NR-, carbamate (-O-C(O)-NR-), carbonate (-O-C(O)-O-), sulfone (-S(O)₂-), ether (-O-), oxime (-CR=N-O-), hydrazone (-CR=N-NR-), urea (-NR-C(O)-NR-), thiourea (-NR-C(S)-NR-), carbohydrate, glyceryl, phosphate (-O-P(O)(OR)O-), phosphonate (-P(O)(OR)O-), saturated and nonsaturated (hetero)cyclic compounds, with R being H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms.
5. An encapsulated protein according to any of claims 1 to 4, wherein the EMCs contain sterically demanding capping groups C.
6. An encapsulated protein according to claim 5, wherein the capping groups within the chain are linear, branched or cyclical alkyl chains.

C(O)-O-), sulfone (-S(O)₂-), ether (-O-), oxime (-CR=N-O-), hydrazone (-CR=N-NR-), urea (-NR-C(O)-NR-), thiourea (-NR-C(S)-NR-), carbohydrate, glyceryl, phosphate (-O-P(O)(OR)O-), phosphonate (-P(O)(OR)O-), saturated and nonsaturated (hetero)cyclic compounds, with R being H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms.

8. An encapsulated protein according to any of claims 5 to 7, wherein the capping groups are highly branched molecules containing branching centers with a branching degree of preferably 2, 3, 4, 5 or 6.
9. An encapsulated protein according to any of claims 5 to 8, wherein the encapsulating organic compound has a dendritic structure.
10. An encapsulated protein according to any of claims 1 to 9, wherein the encapsulation is realized by an EOC-protein inclusion compound EPIC according to the formula (I) in which the EMCs are connected to each other by one branching unit B under formation of a cavity which is vertically and horizontally open



(I)

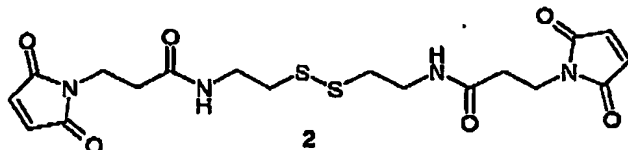
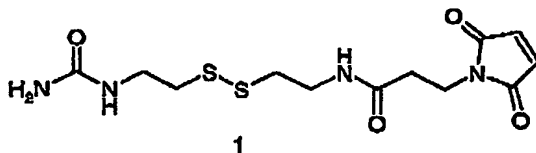
and wherein the symbols have the following meanings:

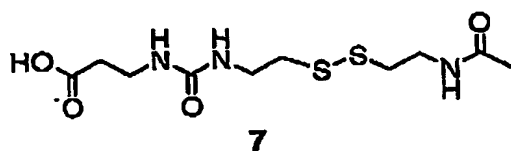
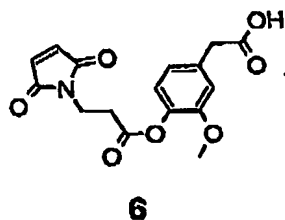
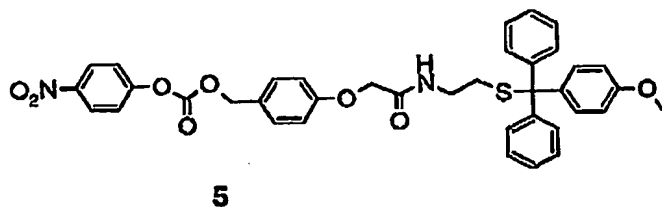
- B:** branching unit containing at least one branching center Bc and at least two branching functional groups Bfg connected to or capable of reacting with an encapsulating molecular chain EMC;
- EMC:** encapsulating molecular chain;
- L:** linker containing at least one functional group Lfg which is connected to the protein P or capable of connecting with functional groups present on the protein P under the formation of a chemical bond;

- I: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 preferably 3, 4, 5, 6, 7 or 8, in particular 2, 3, 4, 5, or 6;
- P: pharmacologically active protein.
11. An encapsulated protein according to claim 10, wherein the branching center Bc contains groups selected from $>C<$, $>CH-$, $>CR-$, $>N-$, $>P-$ which can directly be linked to the branching functional groups Bfg, or can be linked to at least one organic chain.
12. An encapsulated protein according to claim 11, wherein B contains linear, branched or cyclical alkyl chains and optionally further groups selected from S, N, O, $(-S-S)-$, oxyethylen, oxypropylene and oxybutylene, amide $-C(O)NH-$ or $C(O)NR-$, $-S$ -succinimido, amino $(-NR-)$, carboxylic ester $(-C(O)O-)$, sulfonamide $(-S(O)_2-NR-)$, carbamate $(-O-C(O)-NR-)$, carbonate $(-O-C(O)-O-)$, sulfone $(-S(O)_2-)$, ether $(-O-)$, oxime $(-CR=N-O-)$, hydrazone $(-CR=N-NR-)$, urea $(-NR-C(O)-NR-)$, thiourea $(-NR-C(S)-NR-)$, carbohydrate, glyceryl, phosphate $(-O-P(O)(OR)O-)$, phosphonate $(-P(O)(OR)O-)$, saturated and nonsaturated (hetero)cyclic compounds, with R being H, linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms.
13. An encapsulated protein according to claim 11 or 12, wherein B contains branching functional groups Bfg selected from amino $(-NRH)$, carboxylic acid $(-C(O)OH)$ and derivatives, sulfonic acid $(-S(O)_2-OH)$ and derivatives, carbonate $(-O-C(O)-O-)$ and derivatives, hydroxyl $(-OH)$, aldehyde $(-CHO)$, ketone $(-CRO)$, hydrazine (H_2N-NR-) , isocyanate $(-NCO)$, isothiocyanate $(-NCS)$, phosphoric acid $(-O-P(O)(OR)OH)$ and derivatives, phosphonic acid $(-P(O)(OR)OH)$ and derivatives, haloacetyl, alkyl halides, maleimide, acryloyl, arylating agents like aryl fluorides, hydroxylamine, disulfides like pyridyl disulfide, vinyl sulfone, vinyl ketone, diazoalkanes, diazoacetyl compounds, epoxide, oxirane, aziridine, with R being H or a linear, branched or cyclical alkyl group which may contain further functional groups or hetero atoms.

reagent, prodrug linkers containing a bond that can be selectively cleaved under in-vivo conditions, traceless linkers that upon cleavage release protein in such a fashion that the protein is not associated with a remaining linker cleavage product, and traceless prodrug linkers.

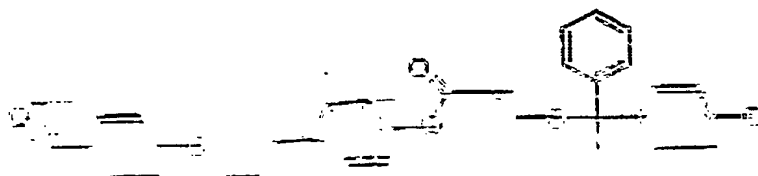
15. An encapsulated protein according to claim 14, wherein the linker is a cleavable linker or a prodrug linker which can be cleaved by TCEP, TFA, DTT, enzymes or buffer.
16. An encapsulated protein according to any of claims 10 to 15, wherein the linker contains functional groups which lend themselves for the reaction between the linker L and the protein under the formation of a chemical bond, and which groups are selected from amino (-NRH), carboxylic acid (-C(O)OH) and derivatives, sulfonic acid (-S(O)₂-OH) and derivatives, carbonate (-O-C(O)-O-) and derivatives, hydroxyl (-OH), aldehyde (-CHO), ketone (-CRO), isocyanate (-NCO), isothiocyanate (-NCS), haloacetyl, alkyl halides, maleimide, acryloyl, arylating agents, aryl fluorides, disulfides, pyridyl disulfide, vinyl sulfone, vinyl ketone, diazoalkanes, diazoacetyl compounds, epoxide, oxirane, aziridine, with R being H or a linear, branched or cyclical alkyl groups which may contain further functional groups or hetero atoms or aryl groups.
17. A encapsulated protein according to any of claims 10 to 16, wherein the linker reagent is selected from those according to the formulae (1), (2), (5), (6) and (7).



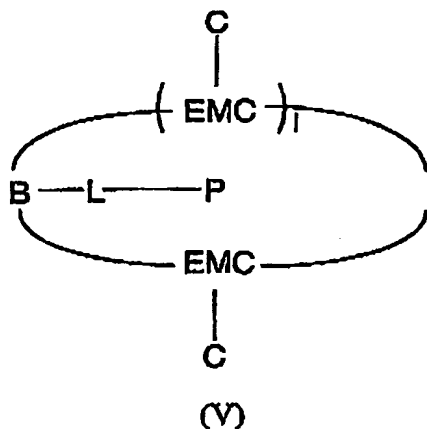


18. An encapsulated protein according to any of claims 10 to 17, wherein the linker is a ~~traceless prodrug linker and contains an ester bond which can be hydrolysed,~~ preferably of a phenol ester, and a carbamate.

19. An encapsulated protein according to claim 18, wherein the linker reagent is in accordance with formula (11)

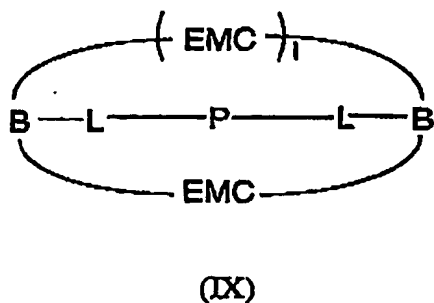


20. An encapsulated protein according to any of claims 10 to 19, wherein the EMCs are in accordance with formula (V) and contain capping groups which are arranged such that a dendrimer-protein inclusion compound DPIC (V) results

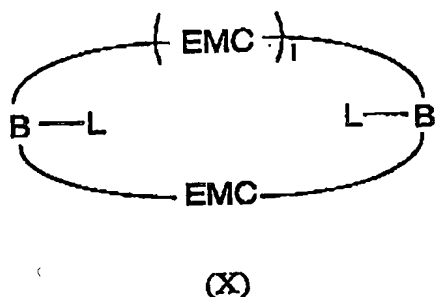


wherein C is a capping group as defined in claims 5 to 8 and the further symbols have the meanings given in claim 10.

21. An encapsulated protein according to claim 20, wherein the capping groups C contain branched heterofunctional units carrying at least one thio-succinimido moiety resulting from a reaction between a maleimido and a thiol group.
22. An encapsulated protein according to any of claims 1 to 9, wherein the encapsulation is realized by a macrocycle-protein inclusion compound MPIC according to the formula (IX)



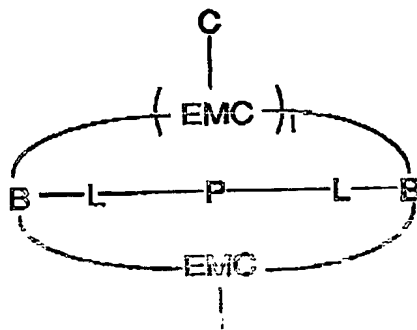
containing a protein P and a macrocyclic structure encapsulating the protein totally or partially according to the general formula (X)



wherein the symbols in formulae (IX) and (X) have the following meanings:

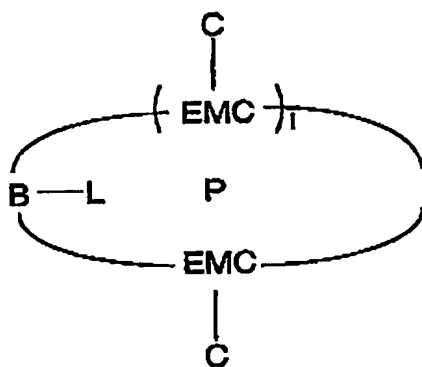
- B: branching unit containing at least one branching center Bc and at least two branching functional groups Bfg connected to or capable of reacting with an encapsulating molecular chain EMC;
- EMC: encapsulating molecular chain;
- L: linker containing at least one functional group Lfg which is connected to the protein B or capable of connecting with functional groups present on the protein P under the formation of a chemical bond;
- l: 1, 2, 3, 4, 5, 6, 7, 8 or 9, preferably 1, 2, 3, 4 or 5, in particular 1, 2 or 3;
- P: pharmacologically active protein.

23. An encapsulated protein according to claim 22, wherein the macrocyclic structure comprises capping groups C resulting in a structure according to the formula

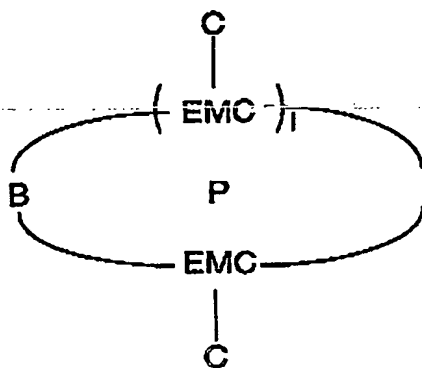


24. An encapsulated protein according to any of the claims 1 to 23, wherein the protein is selected from ACTH, adenosine deaminase, agalsidase, albumin, alfa-1 antitrypsin (AAT), alfa-1 proteinase inhibitor (API), alteplase, anistreplase, ancrod serine protease, antibodies (monoclonal or polyclonal, and fragments or fusions), antithrombin III, antitrypsins, aprotinin, asparaginases, biphalin, bone-morphogenic proteins, calcitonin (salmon), collagenase, DNase, endorphins, enfuvirtide, enkephalins, erythropoietins, factor VIIa, factor VIII, factor VIIIa, factor IX, fibrinolysin, fusion proteins, follicle-stimulating hormones, granulocyte colony stimulating factor (G-CSF), galactosidase, glucagon, glucocerebrosidase, granulocyte macrophage colony stimulating factor (GM-CSF), gonadotropin chorionic (hCG), hemoglobins, hepatitis B vaccines, hirudin, hyaluronidases, iduronidase, immune globulins, influenza vaccines, interleukins (1 alfa, 1 beta, 2, 3, 4, 6, 10, 11, 12), IL-1 receptor antagonist (rhIL-1ra), insulins, interferons (alfa 2a, alfa 2b, alfa 2c, beta 1a, beta 1b, gamma 1a, gamma 1b), keratinocyte growth factor (KGF), lactase, leuprolide, levothyroxine, luteinizing hormone, lyme vaccine, natriuretic peptide, pancrelipase, papain, parathyroid hormone, PDGF, pepsin, platelet activating factor acetylhydrolase (PAF-AH), prolactin, protein C, octreotide, secretin, sermorelin, superoxide dismutase (SOD), somatropins (growth hormone), somatostatin, streptokinase, sucrase, tetanus toxin fragment, tilactase, thrombins, thymosin, thyroid stimulating hormone, thyrotropin, tumor necrosis factor (TNF), TNF receptor-IgG Fc, tissue plasminogen activator (tPA), TSH, urate oxidase, urokinase, vaccines.
25. An encapsulated protein according to any of claims 1 to 24, wherein the protein carries functional groups Pfg which have formed a chemical bond with functional groups Lfg present on the linker, and which functional groups are selected from amino, thiol, hydroxyl, phenol, imidazole, amide, indole, carboxylic acid and guanidino groups, preferably amino, imidazole and thiol groups.
26. An encapsulated protein according to any of claims 1 to 25, wherein the protein is not connected to the linker or any other part of the EOC by a chemical bond.

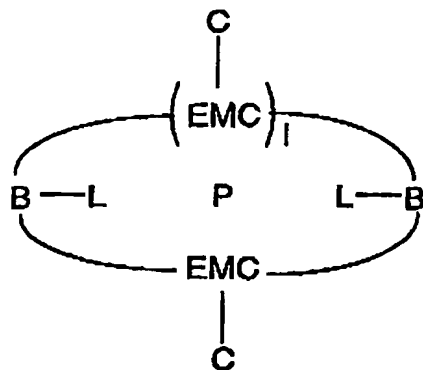
27. An encapsulated protein according to claim 26, wherein parts of the linker remain at the EOC, or the linker has been removed without leaving a residue.
28. An encapsulated protein according to claim 26 or 27, wherein the encapsulated protein is in accordance with any of the formula (XVII) to (XX)



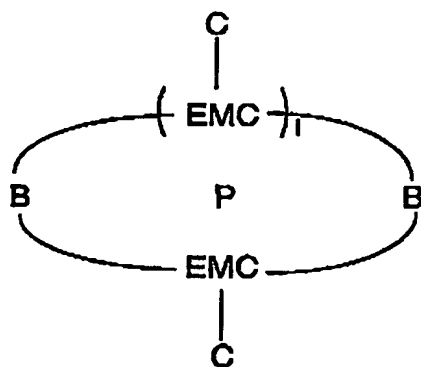
(XVII)



(XVIII)

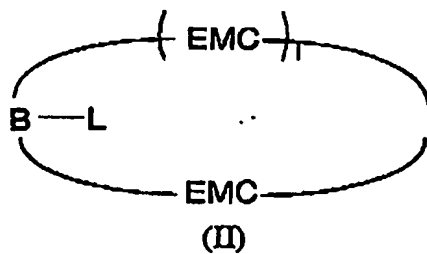


(XIX)

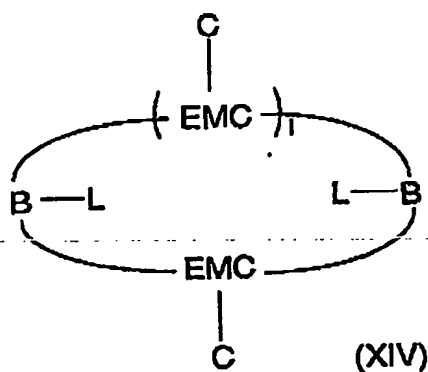
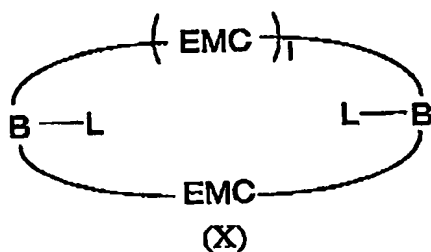
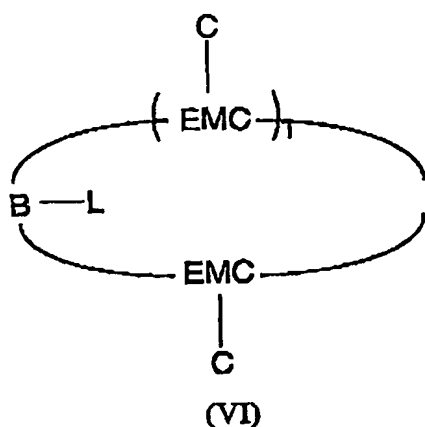


(XX)

29. An encapsulating organic compound EOC according to any of the formulae



(II)



wherein the symbols in formulae (II), (VI), (X), and (XIV) have the following meanings:

B: branching unit containing at least one branching center Bc and at least two branching functional groups Bg connected to or capable of reacting with an

P: pharmacologically active protein.

C: capping group

30. The use of an encapsulating organic compound according to claim 29 for the encapsulation of a protein.
31. A method for the synthesis of an encapsulated protein according to any of the claims 1 to 28, which method comprises
 - providing an EOC according to claim 29;
 - optionally modifying the protein to be encapsulated by introducing functional groups
 - linking the protein and the EOC
 - optionally cleaving the linker
32. A method for selectively delivering a protein to a target, which method comprises
 - providing an encapsulated protein according to any of claims 1 to 28;
 - bringing the encapsulated protein into contact with a body liquid containing the target.
33. A drug containing an encapsulated organic compound according to any of claims 1 to 28.

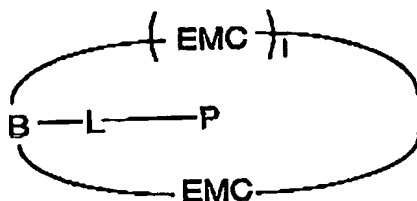
Complex Biosystems GmbH

October 02, 2003
CPX62383EP HN/ht

Abstract

The present invention relates to a protein encapsulated by an encapsulating organic compound (EOC) or proteophore wherein the protein and the encapsulating organic compound are present in 1:1 stoichiometry.

In one embodiment of the present invention, the encapsulation is realized by an EOC protein inclusion compound EPIC according to the formula (I) in which the EMCs are connected to each other by one branching unit B under formation of a cavity which is vertically and horizontally open



(I)

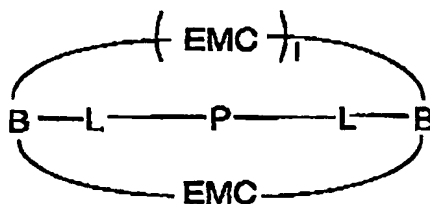
and wherein the symbols have the following meanings:

B: branching unit containing at least one branching center Bc and at least two branching functional groups Bfg connected to or capable of reacting with an encapsulating molecular chain EMC;

EMC: encapsulating molecular chain;

L: linker containing at least one functional group Lfg which is connected to the protein P or capable of connecting with functional groups present on the protein P under formation of a chemical bond.

In a further embodiment of the present invention, the encapsulation is realized by a macrocycle-protein inclusion compound MPIC according to the formula (IX)



(IX)

containing a protein P and a macrocyclic structure, and wherein the symbols have the meanings given above for formula (I).

The EMCs can comprise substituents (capping groups), which can result in dendritic structures.

The encapsulation assures a controlled release of a protein.

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